



Two new records of the genus *Trioxys* (Hymenoptera, Braconidae, Aphidiinae) parasitic on bamboo aphids from South Korea

Sangjin Kim[‡], Juhyeong Sohn[‡], Hyojoong Kim[‡]

[‡] Kunsan National University, Gunsan, Republic of Korea

Corresponding author: Hyojoong Kim (hkim@kunsan.ac.kr)

Academic editor: Jose Fernandez-Triana

Received: 11 Jan 2024 | Accepted: 29 Feb 2024 | Published: 14 Mar 2024

Citation: Kim S, Sohn J, Kim H (2024) Two new records of the genus *Trioxys* (Hymenoptera, Braconidae, Aphidiinae) parasitic on bamboo aphids from South Korea. Biodiversity Data Journal 12: e118599.

<https://doi.org/10.3897/BDJ.12.e118599>

Abstract

Background

The genus *Trioxys* Haliday, 1833 consists of more than 80 species worldwide with three species being recorded in South Korea. In this study, we report the first observation of the two additional species, *T. liui* Chou & Chou, 1993 from *Takecallis arundinariae* (Essig, 1917) on *Phyllostachys bambusoides* Siebold & Zucc., 1843 and *T. remaudierei* Stary & Rakhshani, 2017 from *T. taiwana* (Takahashi, 1926) on *Sasa borealis* (Hack.) Makino & Shibata, 1901.

New information

Trioxys liui and *T. remaudierei* are described and reported with photographs of the diagnostic morphological characters and the mitochondrial *cytochrome c oxidase subunit I* (COI) data (barcode region) and Bayesian tree of the phylogenetic analysis amongst the closely-related taxa are provided.

Keywords

DNA barcoding, natural enemy, parasitoid wasps, systematics, taxonomy

Introduction

The genus *Trioxys* Haliday, 1833 (Hymenoptera, Braconidae, Aphidiinae) consists of more than 80 known species around the world (Rakhshani et al. 2012, Yu et al. 2016, Rakhshani et al. 2017, Črkić et al. 2021, Davidian 2021, Petrović et al. 2021), with three species being recorded in South Korea (National Institute of Biological Resources 2022). As a key character for classification to the two morphologically similar genera, *Trioxys* Haliday, 1833 and *Binodoxys* Mackauer, 1960, a pair of accessory prongs are evident on the abdominal sternite. The former has only a spiracular (primary) tubercle, while the latter has not only a spiracular tubercle, but also has a secondary tubercle (Mackauer 1961, Starý 1981).

The plant subfamily Bambusoideae Luer., (1893) (Cyperales, Poaceae), commonly known as Bamboo, consists of 120 genera with more than 1600 species in the world (Soreng et al. 2017) and five genera with 13 species being recorded in South Korea (National Institute of Biological Resources 2022). Bamboo, an evergreen perennial flowering plant, is represented by three main species in Korea: *Phyllostachys bambusoides* Siebold & Zucc., 1843, *P. nigra* var. *henonis* (Mitford) Stapf ex Rendle, 1904 and *Pseudosasa japonica* (Siebold & Zucc. ex Steud.) Makino ex Nakai, 1920, these being the most widely distributed in the region (Kelchner 2013, Zhao et al. 2019).

The genus *Takecallis* Mastumura, 1917 (Hemiptera, Aphididae, Calaphidinae) consists of eight valid species in the world (Remaudière and Remaudière 1997, Lee and Lee 2018, Wieczorek and Sawka-Gądek 2023) with five species recorded in South Korea (National Institute of Biological Resources 2022). *Takecallis* species was regarded as a pest in various bamboos, *Arundinaria* spp., *Bambusa* spp., *Dendrocalamus* spp., *Phyllostachys* spp., *Pleioblastus* spp., *Pseudosasa* spp., *Sasa* spp. and *Yushania* sp. (Quednau 2003, Qiao and Zhang 2004). In this study, we provide the diagnostic characters for two species, *Trioxys liui* and *T. remaudierei*, from the *Takecallis* species in bamboo spp. and analyse their phylogenetic amongst closely-related congeneric species, using the *COI* barcode region.

Materials and methods

Field and Taxonomic works

Samples were collected by searching for *Takecallis* mummies (*T. taiwana* (Takahashi, 1926) and *T. arundinariae* (Essig, 1917)) on various bamboo species (*Phyllostachys bambusoides* and *Sasa borealis*). Leaves containing mummified aphids were then collected and placed in a clean insect breeding dish (SPL Life Sciences, Korea). To ensure a sufficient number of samples, these dishes were kept in the laboratory at room

temperature. The emergence of parasitoid wasps was monitored daily and they were collected using an insect aspirator. Subsequently, the collected wasps were preserved in 80% ethyl alcohol at -19°C .

Morphological identification of *Trioxys* species was referred from Chou and Chou (1993), Davidian (2007), Yu et al. (2016), Das and Chakrabarti (2017), Rakhshani et al. (2017), Rakhshani et al. (2020) and that of *Takecallis* species from Lee and Lee (2018) and Wieczorek and Sawka-Gądek (2023). Terminology of morphological characters for the Aphidiinae follows Wharton et al. (1997) (for the venation of wing) and Takada (1968). We first performed morphological sorting of similar phenotypes and labelling of this sample using a stereomicroscope (OLYMPUS SZX16, Leica M205C), after which DNA extraction was performed.

After conducting both morphological and molecular identification, measurements of unrecorded species were carried out. A LEICA DMC2900 digital camera and a LEICA M205 C microscope (Leica Geosystems AG) were utilised for photography and characterisation. Multiple pictures were taken at various heights using multifocusing technology. LAS V4.11 (Leica Geosystems AG) and HeliconFocus 7 (Helicon Soft) software were used for the stacking process. After the stacking procedure, illustrations were generated using Adobe Photoshop CS6. LAS V.4.11 (Leica Geosystems AG) was utilised to determine the shape of the specimens (Berkovitch et al. 2009).

Molecular analysis

Total genomic DNA extraction was performed using a LaboPass Tissue Kit (COSMOgenetech, Korea) following the manufacturer's protocol. To preserve a morphologically complete specimen, the DNA extraction method was slightly modified from the "freezing method" used by Yaakop et al. (2013). In the original protocol, the sample was incubated for 30 minutes at 56°C with 200 μl of TL buffer + 20 μl of proteinase K. In the slightly modified DNA extraction methods, a 2 hour incubation period at the same temperature was used. Genomic DNA was extracted individually from each sample.

The target site for molecular identification was the front partial region of mitochondrial *COI*, a 658-bp fragment, amplified using primers, LCO1490 (forward) 5'-GGTCAACAAATCA TAAAGATATTGG-3' and HCO2198 (reverse) 5'-TAAACTTCAGGGTGACCAAAAAATCA-3' (Folmer et al. 1994), with AccuPower PCR PreMix (Bioneer Corp., Daejeon, Korea). Polymerase chain reaction (PCR) amplification was conducted with 20 μl of a reaction mixture consisting of 3 μl of DNA extract, 2 μl of primer and 15 μl of ddH_2O . It was carried out as follows: denaturation for 5 min at 95°C ; 4 cycles of 20 s at 95°C , 30 s at 55°C (decreasing incrementally by 2°C per cycle) and 40 s at 72°C , 31 cycles of 20 s at 95°C , 30 s at 48°C and 40 s at 72°C ; and final extension at 72°C for 5 min. PCR products were visualised by electrophoresis on agar gel and, if a band existed, we commissioned Macrogen (Daejeon, Korea) for purification and sequencing analysis.

Using MEGA version 7.0 (Kumar et al. 2016), sequences were aligned by ClustalW default settings and their frame-shifts checked to avoid pseudogenes. Alignments were translated

to amino acids using MEGA version 7.0. We calculated sequence divergences using the 'p-distance' model commonly with 1,000 bootstrapping replications and complete deletion in data gaps.

A phylogenetic tree was constructed with the Bayesian method using BEAST2 (Bouckaert et al. 2014). To produce dated phylogenies, we used an optimised relaxed clock model (Drummond et al. 2006) in BEAUti and other options were set at default. MCMC analysis was performed and checked using Tracer, DensiTree. After that, we constructed the consensus tree using TreeAnnotator with posterior probability limit 1.0 setting.

Molecular identification was based on Ratnasingham and Herbert (2007). The front partial region 658 bp of the *COI* fragment was sequenced from *T. liui* and *T. remaudierei* and deposited in GenBank. Altogether, 14 sequences of six species, containing the outgroup, were retrieved from GenBank and BOLD (<http://www.boldsystems.org>) and were used to compare them with *T. liui* and *T. remaudierei* (Table 1).

Table 1.

Analysis sample list from GenBank (1-11, 16-18) with our own sample (12-15).

No	Species	NCBI accession number	BOLD ID
1	<i>Trioxys complanatus</i>	KJ848479.1	-
2	<i>T. liui</i>	MT324249.1	-
3	<i>T. liui</i>	MT324250.1	-
4	<i>T. pallidus</i>	KM973397.1	GBAHB12494-15
5	<i>T. pallidus</i>	KM973342.1	GBAHB1349-15
6	<i>T. pallidus</i>	KM973234.1	GBAHB1457-15
7	<i>T. sunnysidensis</i>	KR789189.1	JSJUN008-11
8	<i>T. sunnysidensis</i>	MG438589.1	BARSE352-16
9	<i>T. ulmi</i>	MT873046.1	-
10	<i>T. ulmi</i>	MT873047.1	-
11	<i>T. ulmi</i>	MT873048.1	-
12	<i>T. liui</i>	PP373116.1	-
13	<i>T. liui</i>	PP373117.1	-
14	<i>T. remaudierei</i>	PP373118.1	-
15	<i>T. remaudierei</i>	PP373119.1	-
16	<i>Aphidius uzbekistanicus</i>	ON759206.1	-
17	<i>Aphidius uzbekistanicus</i>	ON827042.1	-
18	<i>Aphidius uzbekistanicus</i>	ON827045.1	-

Taxon treatments

Trioxys liui Chou & Chou, 1993

- GenBank [PP373116](#)
- GenBank [PP373117](#)

Nomenclature

Trioxys liui Chou & Chou, 1993 - Chou and Chou (1993): 375-378.

Trioxys sp. Starý and Schlinger (1967), 1967: 127.

Trioxys bambusa Liu, 1975 - Liu (1975): 69 (nomen nudum).

Material

- scientificName: *Trioxys liui* Chou & Chou, 1993; kingdom: Animalia; phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Braconidae; taxonRank: species; genus: *Trioxys*; specificEpithet: *liui*; scientificNameAuthorship: Chou & Chou, 1993; higherGeography: East Asia; country: South Korea; countryCode: KR; stateProvince: Jeollabuk-do; municipality: Gunsan-si; locality: 290-2, Singwan-dong; eventDate: 08-11-2022; year: 2022; month: 11; day: 8; individualCount: 6; sex: female; lifeStage: adult; preparations: in 80% ethanol and dry-specimen; occurrenceStatus: present; identifiedBy: Sangjin Kim, JuHyeong Sohn, Hyojoong Kim; institutionCode: KSNU; basisOfRecord: preserved specimen; occurrenceID: 1A829D92-1D9D-5A03-BB52-E23958988DE4

Description

Female. Length of body about 1.99 mm (Figure Fig. 1A). Length of forewing 1.67 mm (Fig. 1K).

Head. Eyes oval, sparsely setose. Tentorial index 0.20 (Fig. 1D). Clypeus oval with 6 setae. Malar space 0.09 times as long as longitudinal eye diameter. Antenna 11-segmented (Fig. 1B). F1 equal with F2 (Fig. 1C). F1 and F2 4.1 and 3.4 times as long as their width at the middle, respectively. F1 and F2 are with two and three longitudinal placodes, respectively. Maxillary palp with four palpomeres, labial palp with two palpomeres. Ratio of eye to temple in dorsal view 0.7. Face width/height ratio 1.3 (Fig. 1D).

Mesosoma. Mesoscutum with notaulices on anterior part, dorsal surface smooth, scarcely setose (Fig. 1E). Head width/mesoscutum width ratio 1.4. Propodeum areolated, areola length/width ratio subequal (1.01×) (Fig. 1F). Pterostigma 3.6 times as long as width. Ratio of pterostigma length to R1 vein (= metacarpus) length 1.4. r and RS veins extended (Fig. 1D).

Metasoma. Petiole 1.5 times as long as wide at spiracles (Fig. 1I and J). Ovipositor sheath stout, concave on ventral margin. Ratio of ovipositor sheath width/length 2.0 at

base (Fig. 1G). Anal prongs (= accessory prongs) almost straight, slightly curved upwards at apex. Dorsal side of prongs four setae, with one claw-like apical bristle and two setae at apex (Figure 2H).

Colour. Antenna brown; scape, pedicel and F1 yellowish-brown. Head and face dark brown, clypeus with mouth-parts yellowish-brown. Mesosoma and metasoma brown; Petiole yellowish-brown. Legs yellowish-brown with dark apices.

Parasite of: *Takecallis taiwana* on *Sasa borealis*

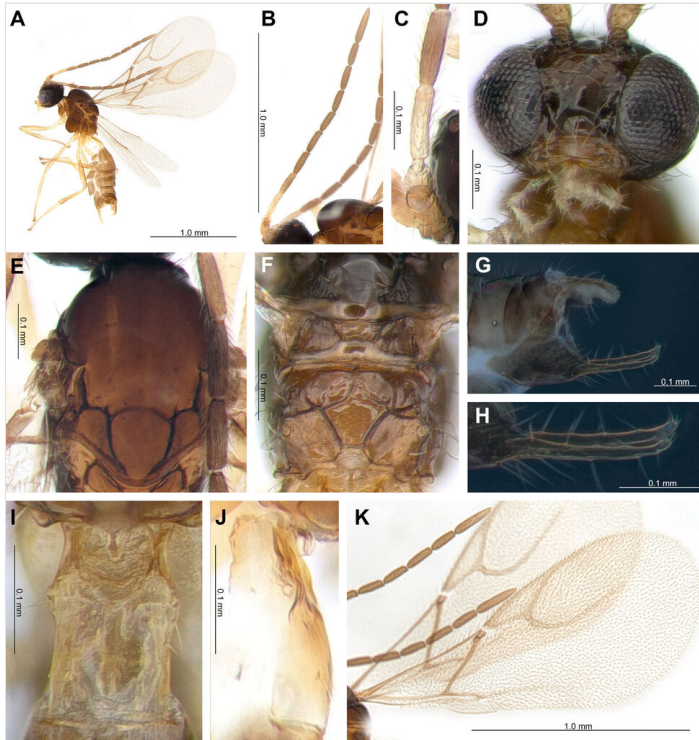


Figure 1. [doi](#)

Trioxys liui: **A** Habitus; **B** Antenna; **C** F1 and F2; **D** Head; **E** Mesoscutum; **F** Propodeum; **G** Ovipositor; **H** Prong; **I** Dorsal view of petiole; **J** Lateral view of petiole; **K** Forewing.

Trioxys remaudierei Starý & Rakhshani, 2017

- GenBank [PP373118](#)
- GenBank [PP373119](#)

Nomenclature

Trioxys remaudierei Starý & Rakhshani, 2017 - Rakhshani et al. (2017): 1237-1248.

Material

- a. scientificName: *Trioxys remaudierei* Stary & Rakhshani, 2017; kingdom: Animalia; phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Braconidae; genus: *Trioxys*; specificEpithet: *remaudierei*; scientificNameAuthorship: Stary & Rakhshani, 2017; higherGeography: East Asia; country: South Korea; countryCode: KR; stateProvince: Jeollabuk-do; municipality: Gimje-si; locality: Cheongha-myeon; verbatimLocality: 119, Jangsan-ri; eventDate: 07-11-2022; year: 2022; month: 11; day: 7; individualCount: 7; sex: female; lifeStage: adult; preparations: in 80% ethanol, dry-specimen; occurrenceStatus: present; identifiedBy: Sangjin Kim, JuHyeong Sohn, Hyojoong Kim; language: en; institutionCode: KSNU; occurrenceID: 98714FFC-467A-5148-97A4-D142E794F19F

Description

Female. Length of body about 2.3 mm (Fig. 2A). Length of forewing 1.9 mm (Fig. 2K).

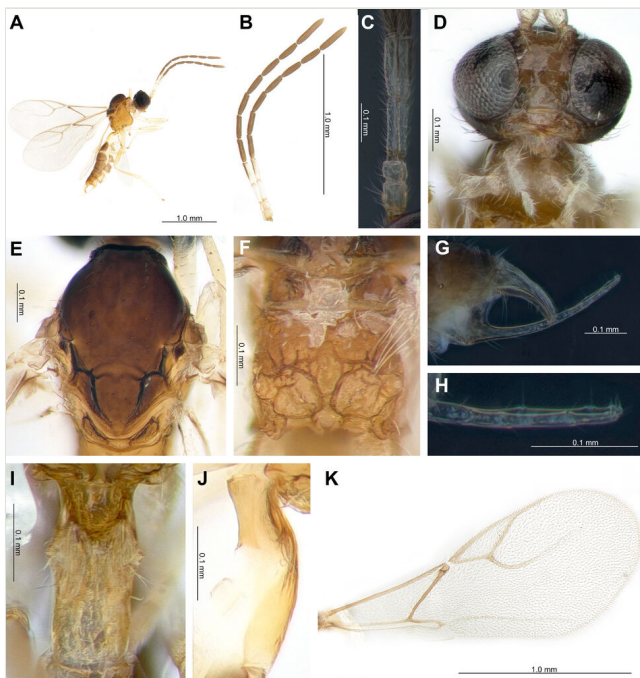


Figure 2. [doi](#)

Trioxys remaudierei: **A** Habitus; **B** Antenna; **C** F1 and F2; **D** Head; **E** Mesoscutum; **F** Propodeum; **G** Ovipositor; **H** Prong; **I** Dorsal view of petiole; **J** Lateral view of petiole; **K** Forewing.

Head. Eyes oval, sparsely setose. Tentorial index 0.14 (Fig. 2D). Clypeus oval with seven setae. Malar space 0.08 times as long as longitudinal eye diameter. Antenna 11-segmented (Fig. 2B). F1 slightly shorter than F2 (F2 1.07 times as long as F1) (Fig. 2C). F1 and F2 5.0 and 4.1 times as long as their width at the middle, respectively. F1

without longitudinal placodes and F2 with 3-4 longitudinal placodes. Maxillary palp with four palpomeres, labial palp with two palpomeres. Ratio of eye to temple in dorsal view 0.6. Face width/height ratio 1.2 (Fig. 2D).

Mesosoma. Mesoscutum with notaulices on anterior part, dorsal surface smooth, two rows of scarcely setose start at end of notaulus (Fig. 2E). Head width/mesoscutum width ratio 1.3. Propodeum areolated, areola length/width ratio 1.2 (Fig. 2F). Pterostigma 3.5 times as long as width. Ratio of pterostigma length to R1 vein (= metacarpus) length 2.5 (Fig. 2K). r and RS veins extended.

Metasoma. Petiole 2.0 times as long as wide at spiracles (Fig. 2I and J). Ovipositor sheath elongate, gently concave on ventral margin. Ratio of ovipositor sheath width/length 2.8 at base (Fig. 2G). Anal prongs (= accessory prongs) long and straight upwards, bifurcated on apical one-third to two-thirds (apical one-third are four, apical half is two, apical two-third is one), each having one claw-like apical bristle and one seta at apex with two or three setae at dorsal side (Fig. 2H).

Colour. Antenna brown; scape, pedicel, F1 and F2 yellowish-brown, sometimes dorsal part of F2 brown; Head, face and clypeus with mouth-parts dark brown. Dorsal side of mesoscutum and metasoma dark brown, except light brown propodeum and yellowish-brown petiole. Legs yellowish-brown with dark apices.

Parasite of: *T. arundinariae* on *Phyllostachys bambusoides*.

Notes

In the original description, the antenna of this species was 13-segmented. However, it was a mistake in the email from the author. This species is really 11-segmented, but two segments are artificially repeated in the line drawing.

Analysis

A Bayesian tree was constructed with twelve sequences from seven species, including an outgroup. Three clades were identified: Clade A represented *Trioxys sunnysidensis*, Clade B included *T. remaudierei*, both identified as monophyletic and the remaining species formed Clade C (Fig. 3). Within Clade C, *T. liui* was observed as the sister group to *T. pallidus*, *T. companatus* and *T. ulmi*. In comparison to a previous study by Črkrić et al. (2021), *T. liui* exhibited a consistent pattern, but in this study, it was positioned between *T. ulmi* and the *T. pallidus-complanatus* group. *Trioxys remaudierei* was situated between Clade A and Clade C (Fig. 3).

Intraspecific and interspecific distances ranged from 0.000 to 0.011 (averaging 0.003) and 0.057 to 0.128 (averaging 0.108), respectively (Table 2). In Clade C, interspecific genetic distances ranged from 0.102 to 0.120 (averaging 0.113) between *T. liui* and the other species (*T. pallidus*, *T. complanatus* and *T. ulmi*). Genetic distances between *T.*

remaudierei and *T. sunnysidensis* were 0.105 and genetic distances within Clade C were 0.111 (Table 2).

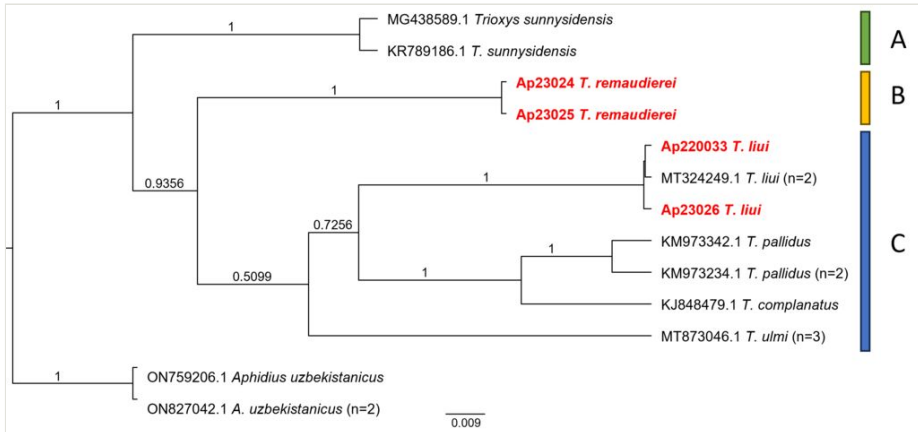


Figure 3. [doi](#)

Phylogenetic tree of six *Trioxys* spp. estimated by the Bayesian method using their COI DNA barcode data. *Aphidius uzbekistanicus* was used as an outgroup. Bootstrap support values more than 50% are indicated above branches. Scale-bar means the expected rate of a nucleotide substitution.

Table 2.

Calculated genetic distances, based on COI sequences between *Trioxys* spp. used in the analysis.

	<i>T. liui</i> (n = 4)	<i>T. remaudierei</i> (n = 2)	<i>T. complanatus</i>	<i>T. pallidus</i> (n = 3)	<i>T. sunnysidensis</i> (n = 2)	<i>T. ulmi</i> (n = 3)
<i>Trioxys liui</i>	(0.000)	-	-	-	-	-
<i>T. remaudierei</i>	0.111	(0.000)	-	-	-	-
<i>T. complanatus</i>	0.120	0.120	(0.000)	-	-	-
<i>T. pallidus</i>	0.118	0.113	0.057	(0.011)	-	-
<i>T. sunnysidensis</i>	0.110	0.105	0.121	0.128	(0.005)	-
<i>T. ulmi</i>	0.102	0.101	0.115	0.106	0.100	(0.000)

Discussion

Most of the *Trioxys* species typically exhibit morphology characterised by two prongs (Mackauer 1960, Mackauer 1961). However, *T. remaudierei* stands out as it starts with a single prong that gradually bifurcates into two towards the apex (Fig. 2G and H). This morphology is not unique to *T. remaudierei* alone, other species like *T. tenuicaudus* Starý, 1978, *T. udalovi* Davidian, 2005, *T. betulae* Marshall, 1896 and *T. artistigma* Telenga, 1953 also exhibit this morphology (Davidian 2005, Davidian 2007). The genus *Trioxys* uses anal prongs (= accessory prongs) for grasping and restraining host movement (Eidmann 1924,

Schlinger and Hall 1961, Shaw and Huddleston 1991) and may have evolved into two strands to maximise it. Therefore, such prong morphology was considered a potential indicator of evolutionary ancestral traits even although it seemed to be not a clear correlation between taxa due to the limited number of samples in this study.

In case of *T. liui*, it is parasitic on *Cranaphis formosanus* (Takahashi) (Liu, 1975) and parasitic on *Phyllaphoides bambusicola* Takahashi, 1921 on *Phyllostachys makinoi* Hayata, 1915 in China (Chou and Chou 1993). Moreover, *Takecallis* sp. has been recorded on *Phyllostachys aurea* Carrière ex. Rivière & C. Rivière, 1878, *T. taiwana* and *T. arundinariae* on *Phyllostachys* sp. and captured on *Indocalamus tessellatus* (Munro) Keng f., 1957 in Spain (Rakhshani et al. 2020). Similarly, *T. remaudierei* parasite *T. taiwana* on *Phyllostachys* sp. in France and *T. arundinariae* on *Phyllostachys* sp. in Spain (Rakhshani et al. 2017). This study newly records of *T. liui* is parasitic on *T. taiwana* on *Sasa borealis*, and *T. remaudierei* is parasitic on *T. arundinariae* on *P. bambusoides* in South Korea.

Both species have already been recorded in Europe and are considered invasive or subsequent adaptation into western Europe (Rakhshani et al. 2020). Aphidiinae species exhibit strong host specificity (Starý 1978) and both species show a strong host specificity for *Takecallis* species. Furthermore, since *Takecallis* species have been recorded only on bamboo species (Lee and Lee 2018), the distribution of *T. liui* and *T. remaudierei* parasitising on it seems to be limited to areas where bamboo is located. In the natural range of *Takecallis* species, all known species of this genus are restricted to East Asia (China, India, Japan, Korea, Manchurian subregion, Taiwan), *T. arundicolens* (Clarke, 1903), *T. arundinariae* and *T. taiwana*, *T. nigroantennatus* Wieczorek, 2023 are now widely distributed and introduced to other countries, including Africa (Algeria), USA (California), England, Netherlands, Hungary, Madeira, Australia, New Zealand, Argentina, Spain and Poland (Blackman and Eastop 2023, Wieczorek and Sawka-Gądek 2023). Consequently, parasitoids of *Takecallis* species are likely to be invasive species outside East Asia, with invasive countries potentially hosting distributions of these species.

Acknowledgements

This work was funded by the Ministry of Environment (MOE) of the Republic of Korea (NIBR202304203). It was also supported by the Basic Science Research Program through the National Research Foundation of Korea funded by the Ministry of Education (2022R1A2C1091308).

Author contributions

Sangjin Kim wrote the text; Sangjin Kim, JuHyeong Sohn and Hyjooing Kim collected material, carried out the DNA experiment and provided photographs; all authors revised the manuscript.

References

- Berkovitch A, Belfer I, Hassin Y, Landa E (2009) Diffraction imaging by multifocusing. *Geophysics* 74 (6): WCA74-WCA81. <https://doi.org/10.1190/1.3198210>
- Blackman RL, Eastop VF (2023) Aphids of the World's Plants: An Online Identification and Information Guide. <http://www.aphidsonworldsplants.info>. Accessed on: 2024-2-22.
- Bouckaert R, Heled J, Kühnert D, Vaughan T, Wu C, Xie D, Suchard M, Rambaut A, Drummond A, et al. (2014) BEAST 2: A software platform for Bayesian evolutionary analysis. *PLOS Computational Biology* 10 (4): e1003537. <https://doi.org/10.1371/journal.pcbi.1003537>
- Chou LY, Chou KC (1993) A new species of *Trioxys* (Hymenoptera: Braconidae) from Taiwan. *Chinese Journal of Entomology* 13 (4): 375-378. <https://doi.org/10.6660/TESE.1993036>
- Črkrić J, Petrović A, Kocić K, Tomanović Ž (2021) Insights into phylogenetic relationships between *Trioxys* Haliday, 1833 and *Binodoxys* Mackauer, 1960 (Hymenoptera, Braconidae, Aphidiinae), with a description of a new species of the genus *Trioxys*. *Zoosystema* 43 (8): 145-154. <https://doi.org/10.5252/zoosystema2021v43a8>
- Das BC, Chakrabarti S (2017) Aphidiid parasitoids (Hymenoptera: Aphidiidae) of graminaceous aphids in Garhwal, Western Himalaya. *Oriental Insects* 23 (1): 365-372. <https://doi.org/10.1080/00305316.1989.11835526>
- Davidian EM (2005) A review of species of the subgenus *Trioxys* s. str., genus *Trioxys* Haliday (Hymenoptera, Aphidiidae) of Russia and adjacent countries. *Entomologicheskoe Obozrenie* 84 (3): 579-609.
- Davidian EM (2007) Family Aphidiidae. In: Lelej AS (Ed.) *Key to the insects of Russian Far East*. 4. Vladivostok. [In Russian].
- Davidian EM (2021) A new species of the aphidiid wasp genus *Trioxys* Haliday (Hymenoptera, Aphidiidae) from Orenburg Province of Russia. *Entomological Review* 100 (7): 1029-1032. <https://doi.org/10.1134/s0013873820070106>
- Drummond AJ, Ho SW, Phillips MJ, Rambaut A (2006) Relaxed phylogenetics and dating with confidence. *PLOS Biology* 4 (5): e88. <https://doi.org/10.1371/journal.pbio.0040088>
- Eidmann H (1924) Die Eiablage von *Trioxys* Hal. (Hym., Braconidae). *Zeitschrift für Angewandte Entomologie* 10 (2): 353-363. <https://doi.org/https://doi.org/10.1111/j.1439-0418.1924.tb01533.x>
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology Biotechnology* 3 (5): 294-299. URL: https://www.mbari.org/wp-content/uploads/2016/01/Folmer_94MMBB.pdf
- Kelchner S (2013) Higher level phylogenetic relationships within the bamboos (Poaceae: Bambusoideae) based on five plastid markers. *Molecular Phylogenetics and Evolution* 67 (2): 404-413. <https://doi.org/10.1016/j.ympev.2013.02.005>
- Kumar S, Stecher G, Tamura K (2016) MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution* 33 (7): 1870-1874. <https://doi.org/10.1093/molbev/msw054>

- Lee Y, Lee S (2018) A review of the genus *Takecallis* Mastumura in Korea with the description of a new species (Hemiptera, Aphididae). Plazi.org taxonomic treatments database 748: 131-149. <https://doi.org/10.3897/zookeys.748.23140>
- Liu CS (1975) Aphid parasites and their propagation in Taiwan. Journal of Agricultural Research of China 24 (3-4): 62-84. [In Chinese].
- Mackauer M (1960) Zur Systematik der Gattung *Trioxys* Haliday (Hymenoptera: Braconidae, Aphidiinae). Contributions to Entomology = Beiträge zur Entomologie 10 (1-2): 137-160. <https://doi.org/10.21248/contrib.entomol.10.1-2.137-160>
- Mackauer M (1961) Die gattungen der Familie Aphidiidae und ihre verwandtschaftliche zuordnung (Hymenoptera: Ichneumonoidea). Beiträge Zur Entomologie 11: 792-803.
- National Institute of Biological Resources (2022) National list of species of Korea. <https://kbr.go.kr/>. Accessed on: 2023-12-26.
- Petrović A, Kocić K, Črkić J, Tomanović Ž (2021) Additional data on Aphidiinae (Hymenoptera, Braconidae) fauna of Kyrgyzstan, with description of a new species. Journal of Hymenoptera Research 82: 221-235. <https://doi.org/10.3897/jhr.82.64968>
- Qiao GX, Zhang GX (2004) Review of the genus *Takecallis* Matsumura (Homoptera: Aphididae: Myzocallidinae) from China and description of one new species. The Raffles Bulletin of Zoology 52 (2): 373-378.
- Quednau FW (2003) Atlas of the drepanosiphine aphids of the World: Panaphidini Oestlund, 1923-Panaphidina Oestlund, 1923 (Hemiptera: Aphididae: Calaphidinae). American Entomological Institute, Gainesville. [ISBN 9781887988162]
- Rakhshani E, Kazemzadeh S, Starý P, Barahoei H, Kavallieratos N, Četković A, Popović A, Bodlah I, Tomanović Ž (2012) Parasitoids (Hymenoptera: Braconidae: Aphidiinae) of Northeastern Iran: aphidiine-aphid-plant associations, key and description of a new species. Journal of Insect Science 12 (143): 1-26. <https://doi.org/10.1673/031.012.14301>
- Rakhshani E, Pons X, Lumbierres B, Havelka J, Pérez Hidalgo N, Tomanović Ž, Starý P (2017) A new parasitoid (Hymenoptera: Braconidae: Aphidiinae) of the invasive bamboo aphids *Takecallis* spp. (Hemiptera: Aphididae) from Western Europe. Journal of Natural History 51: 1237-1248. <https://doi.org/10.1080/00222933.2017.1327622>
- Rakhshani E, Saval JM, Hidalgo NP, Pons X, Kavallieratos N, Starý P (2020) *Trioxys liui* Chou & Chou, 1993 (Hymenoptera, Braconidae, Aphidiinae): an invasive aphid parasitoid attacking invasive *Takecallis* species (Hemiptera, Aphididae) in the Iberian Peninsula. ZooKeys 944: 99-114. <https://doi.org/10.3897/zookeys.944.51395>
- Ratnasingham S, Herbert PN (2007) Bold: The Barcode of Life Data System (<http://www.barcodinglife.org>). Molecular Ecology Notes 7 (3): 355-364. <https://doi.org/10.1111/j.1471-8286.2007.01678.x>
- Remaudière G, Remaudière M (1997) Catalogue des Aphididae du monde (Homoptera Aphidoidea). Institut National de la Recherche Agronomique
- Schlinger E, Hall J (1961) The Biology, Behavior, and Morphology of *Trioxys* (*Trioxys*) *utilis*, an Internal Parasite of the Spotted Alfalfa Aphid, *Therioaphis maculata* (Hymenoptera: Braconidae, Aphidiinae). Annals of the Entomological Society of America 54 (1): 34-45. <https://doi.org/10.1093/aesa/54.1.34>
- Shaw M, Huddleston T (1991) Classification and biology of braconid wasps. Handbooks for the identification of British insects 7 (11): 1-126.
- Soreng RJ, Peterson PM, Romaschenko K, Davidse G, Teisher JK, Clark LG, Barberá P, Gillespie LJ, Zuloaga FO (2017) A worldwide phylogenetic classification of the

- Poaceae (Gramineae) II: an update and a comparison of two 2015 classifications. *Journal of Systematics and Evolution* 55: 259-290. <https://doi.org/10.1111/jse.12262>
- Starý P, Schlinger EI (1967) A revision of the far east Asian Aphidiidae (Hymenoptera). Springer, Dordrecht. <https://doi.org/10.1007/978-94-017-6337-0>
 - Starý P (1978) Parasitoid spectrum of the arboricolous callaphidid aphids in Europe (Hymenoptera, Aphidiidae; Homoptera, Aphidoidea, Callaphididae). *Acta Entomologica Bohemoslovaca* 75 (3): 164-177.
 - Starý P (1981) Biosystematical classification of *Trioxys* Hal. and related genera (Hymenoptera, Aphidiidae). *Bollettino del Laboratorio di Entomologia Agraria/Filippo Silvestri* 38: 85-93.
 - Takada H (1968) Aphidiidae of Japan (Hymenoptera). *Insecta Matsumurana* 30 (2): 92-102.
 - Wharton RA, Marsh PM, Sharkey MJ (1997) Manual of the New World genera of the family Braconidae (Hymenoptera). 1. The International Society of Hymenopterists, Washington, DC.
 - Wieczorek K, Sawka-Gądek N (2023) DNA barcoding and molecular phylogenetics revealed a new cryptic bamboo aphid species of the genus *Takecallis* (Hemiptera: Aphididae). *Applied Sciences* 13 (13): 7798. <https://doi.org/10.3390/app13137798>
 - Yaakop S, Achterberg C, Ghani I, Zuki A (2013) Freezing method as a new non-destructive modification of DNA extraction. *Pertanika Journal of Tropical Agricultural Science* 36: 373-392.
 - Yu D, van Achterberg C, Horstmann K (2016) Taxapad 2016. Ichneumonoidea 2015 (Biological and taxonomical information), Taxapad interactive catalogue database on flash-drive.
 - Zhao Y, Zhao Y, Du Y, Kang JS (2019) Characterization and classification of three common Bambusoideae species in Korea by an HPLC-based analytical platform coupled with multivariate statistical analysis. *Industrial Crops and Products* 130: 389-397. <https://doi.org/10.1016/j.indcrop.2018.12.096>