

Length–weight relations of 44 fish species (Actinopterygii) inhabiting an unprotected tropical coastal biological corridor of Yucatan, Mexico

María Eugenia VEGA-CENDEJAS¹, Mirella HERNÁNDEZ DE SANTILLANA¹,
Sonia PALACIOS-SÁNCHEZ¹

¹ Centro de Investigación y de Estudios Avanzados del Instituto Politécnico Nacional, Unidad Mérida, Mérida (CINVESTAV), Yucatán, Mexico

<https://zoobank.org/A865B16B-3505-4CEF-8E7F-A863D292A3B3>

Corresponding author: María Eugenia Vega-Cendejas (maruvega@cinvestav.mx)

Academic editor: Rodolfo Reyes ♦ **Received** 3 August 2023 ♦ **Accepted** 23 October 2023 ♦ **Published** 24 November 2023

Citation: Vega-Cendejas ME, Hernández de Santillana M, Palacios-Sánchez S (2023) Length–weight relations of 44 fish species (Actinopterygii) inhabiting an unprotected tropical coastal biological corridor of Yucatan, Mexico. *Acta Ichthyologica et Piscatoria* 53: 201–207. <https://doi.org/10.3897/aiep.53.110519>

Abstract

Length–weight relations (LWRs) were estimated for 44 fish species, representing 23 families, collected from an unprotected coastal biological corridor of the Yucatan Peninsula. The following species were studied (in alphabetical order): *Acanthostracion quadricornis* (Linnaeus, 1758); *Albula vulpes* (Linnaeus, 1758); *Anchoa hepsetus* (Linnaeus, 1758); *Anchoa lamprotaenia* Hildebrand, 1943; *Anchoa lyolepis* (Evermann et Marsh, 1900); *Anchoa mitchilli* (Valenciennes, 1848); *Archosargus rhomboidalis* (Linnaeus, 1758); *Ariopsis felis* (Linnaeus, 1766); *Bagre marinus* (Mitchill, 1815); *Bairdiella chrysoura* (Lacepède, 1802); *Caranx latus* Agassiz, 1831; *Chaetodipterus faber* (Broussonet, 1782); *Chriodorus atherinoides* Goode et Bean, 1882; *Cynoscion arenarius* Ginsburg, 1930; *Elops saurus* Linnaeus, 1766; *Eucinostomus argenteus* Baird et Girard, 1855; *Eucinostomus gula* (Quoy et Gaimard, 1824); *Eucinostomus harengulus* Goode et Bean, 1879; *Harengula jaguana* Poey, 1865; *Hyporhamphus unifasciatus* (Ranzani, 1841); *Lagodon rhomboides* (Linnaeus, 1766); *Lutjanus griseus* (Linnaeus, 1758); *Menticirrhus americanus* (Linnaeus, 1758); *Menticirrhus littoralis* (Holbrook, 1847); *Menticirrhus saxatilis* (Bloch et Schneider, 1801); *Mugil curema* Valenciennes, 1836; *Mugil trichodon* Poey, 1875; *Oligoplites saurus* (Bloch et Schneider, 1801); *Opisthonema oglinum* (Lesueur, 1818); *Opsanus beta* (Goode et Bean, 1880); *Orthopristis chrysoptera* (Linnaeus, 1766); *Prionotus tribulus* Cuvier, 1829; *Rypticus maculatus* Holbrook, 1855; *Selene vomer* (Linnaeus, 1758); *Sphoeroides spengleri* (Bloch, 1785); *Sphoeroides testudineus* (Linnaeus, 1758); *Strongylura notata* (Poey, 1860); *Strongylura timucu* (Walbaum, 1792); *Symphurus plagiusa* (Linnaeus, 1766); *Synodus foetens* (Linnaeus, 1766); *Trachinotus carolinus* (Linnaeus, 1766); *Trachinotus falcatus* (Linnaeus, 1758); *Trachinotus goodei* Jordan et Evermann, 1896; *Urobatis jamaicensis* (Cuvier, 1816). A new maximum standard length (SL) was recorded for *Anchoa lamprotaenia*. Positive allometric growth was reported in ten species, negative allometric growth in sixteen species, and isometric growth in eighteen species.

Keywords

Length–weight relations, nursery habitats, wetlands, Yucatan Peninsula

Introduction

Length–weight relations (LWRs) of fishes are a key element for the study of their biology, taxonomy, physiology, ecology (Vega-Cendejas et al. 2017), and fish population

dynamics (Kohler et al. 1995). They are useful to calculate the expected weight from the known length of fish and vice versa (Xie et al. 2015; Kuriakose 2017), to estimate the isometric or allometric growth (Teixeira-de Mello et al. 2006), as an indicator of fatness and the relative well-be-

ing of the fish population, the standing stock biomass and comparing the ontogeny of fish populations from different regions (Petraakis and Stergiou 1995). This relation has also been used for species-specific life history comparisons between regions (Wotton 1990), and evaluations of parasite effects (Teixeira-de Mello and Eguren 2008).

The presently reported study was an effort to determine LWRs for 44 fish species from Yucatan, southern Gulf of Mexico. The fish were collected from a chain of unprotected littoral habitats which we will later refer to as the Yucatan Coastal Biological Corridor (YCBC). A biological corridor is a delimited geographic space that promotes connectivity between landscapes, ecosystems, and natural or modified habitats and ensures the maintenance of biological diversity and ecological processes. It also allows genetic exchange between fragmented populations and the integration of these areas into land use planning plans. Studying these areas provides valuable information to propose new locations that require protection, as well as to identify high-priority network linkages between existing marine protected areas (Pendoley et al. 2014) and to define essential habitats for target species (Turk-Boyer et al. 2014).

Biological corridors emerge as a mechanism that attempts to give greater variability to the conservation of species found in wild areas, allowing the movement of biota from one protected area to another or between fragments of ecosystems (Moran et al. 2019). The YCBC unites ecologically protected natural areas through regions with various productive activities and different land uses. Its importance lies in the fact that this system is unique in the association of the species with the habitat and its ecological process, in the way in which the populations

that inhabit the coast using their natural resources. This area has been recognized for having great biodiversity, characterized by the heterogeneity of its habitats with the presence of wetlands, coastal lagoons, and petenes on its coastline. However, the YCBC, which stretches 128 km, has been modified by various anthropogenic activities such as the construction of docks and ports, as well as by artisanal and industrial fishing, aquaculture, and ecotourism (Herrera-Silveira and Morales-Ojeda 2009). Studies in this area have indicated that diversity and abundance of fishery resources increase inside protected areas. However, the surrounding unprotected areas require strategies to allow the free flow of species from one protected area to the other (Palacios-Sánchez et al. 2019).

Material and methods

The YCBC, as part of the Mesoamerican Corridor is located in the tropical region of the southeastern Gulf of Mexico ($21^{\circ}02'48.66''$ – $21^{\circ}21'28.20''$ N, $89^{\circ}07'8.04''$ – $90^{\circ}16'45.84''$ W), which includes 350 km of coastal zone habitats (Euán-Avila et al. 2014) and connects two important reserves in the Yucatan Peninsula—Celestun in the West and Ria Lagartos in the East (Palacios-Sánchez et al. 2019) (Fig. 1).

Sampling of the fish specimens was carried out monthly for three years (October 2001 through April 2004) in 24 localities of the YCBC. Fish samples were collected during the first six hours of light of the day using a benthic trawling net (15 m long \times 1.5 m, 2.54 cm mesh) in all the habitats (wetlands, coastal lagoons, coastline). At each station, to carry out sampling, we walked perpendicular

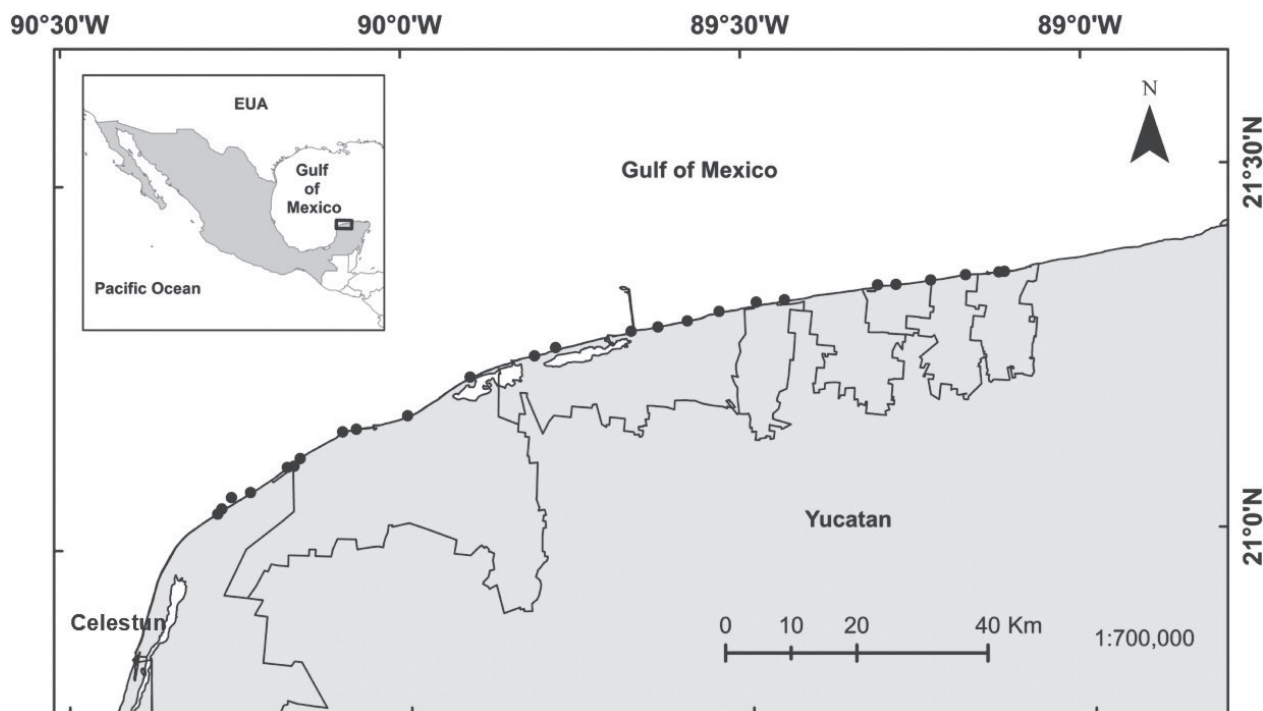


Figure 1. Sampling stations in the study area in the unprotected Yucatan Coastal Biological Corridor, southern Gulf of Mexico.

to the coast, measuring the distance from the shore to record the sampled area. The trawl net was dragged manually, making two replicates per station, separated by 10 m. Collected fishes were euthanized in ice slurry, preserved (70% ethanol), and transported to the laboratory where they were identified using specialized references (Allen 1985; Carpenter 2002a, 2002b; McEachran and Fechhelm 1998, 2005, among others), measured for standard length (SL) (to the nearest 0.1 cm), and weighed (to the nearest 0.01 g). A representative sample of each species was deposited and cataloged in the Ichthyology Collection of the Centro de Investigación y de Estudios Avanzados del Instituto Politécnico Nacional, Mérida, Mexico (CINV-NEC), reference number YUC-PEC.084.0999.

We calculated the LWRs using the allometric formula

$$W = aL^b$$

where W is the weight of the fish [g], L is the standard length [cm], a is the intercept, and b is the allometric coefficient/slope. The values of a and b were calculated with Statgraphics software (Centurion XV, Version 15.1.02, Copyright 1982–2006 StatPoint, Inc.) with a linear least squares regression using a logarithmic scale. Based on the value of the slope (b), the growth of a fish species was considered negative allometric ($b < 3$), positive allometric ($b > 3$), or isometric ($b = 3$) (Froese et al. 2011). Outliers were removed using logarithmic plots, and limits for a and b were estimated by a student's t -test with a 95% confidence (Froese 2006). In some cases, when the number of specimens was too small or the size range was too narrow to estimate the LWRs parameters a and b , we assumed an isometric relation ($b = 3$) (Froese 2006; Hay et al. 2020) and the value of the intercept a will be obtained with the following formula:

$$a = \frac{\sum_{i=1}^n \frac{W}{L^3}}{n}$$

where W refers to the weight [g], L to the standard length [cm], and n to the number of specimens.

Results

The descriptive statistics and the estimated LWRs parameters for 44 species which represents 23 families are summarized in Table 1, including *Urobatis jamaicensis* (Cuvier, 1816) [Urotrygonidae]; *Albula vulpes* (Linnaeus, 1758) [Albulidae]; *Elops saurus* Linnaeus, 1766 [Elopidae]; *Anchoa hepsetus* (Linnaeus, 1758), *Anchoa lamprotaenia* Hildebrand, 1943, *Anchoa lyolepis* (Evermann et Marsh, 1900), *Anchoa mitchilli* (Valenciennes, 1848) [Engraulidae]; *Harengula jaguana* Poey, 1865, *Opisthonema oglinum* (Lesueur, 1818) [Dorosomatidae]; *Ariopsis felis* (Linnaeus, 1766), *Bagre marinus* (Mitchill, 1815) [Ariidae]; *Synodus foetens* (Linnaeus, 1766) [Synodontidae]; *Opsanus beta* (Goode et Bean, 1880) [Batrachoididae]; *Mugil curema* Valenciennes, 1836, *Mugil trichodon*

Poey, 1875 [Mugilidae]; *Strongylura notata* (Poey, 1860), *Strongylura timucu* (Walbaum, 1792) [Belontiidae]; *Chriodorus atherinoides* Goode et Bean, 1882, *Hyporhamphus unifasciatus* (Ranzani, 1841) [Hemiramphidae]; *Caranx latus* Agassiz, 1831, *Oligoplites saurus* (Bloch et Schneider, 1801), *Selene vomer* (Linnaeus, 1758), *Trachinotus carolinus* (Linnaeus, 1766), *Trachinotus falcatus* (Linnaeus, 1758), *Trachinotus goodei* Jordan et Evermann, 1896 [Carangidae]; *Symphurus plagiusa* (Linnaeus, 1766) [Cynoglossidae]; *Eucinostomus argenteus* Baird et Girard, 1855, *Eucinostomus gula* (Quoy et Gaimard, 1824), *Eucinostomus harengulus* Goode et Bean, 1879 [Gerreidae]; *Rypticus maculatus* Holbrook, 1855 [Grammistidae]; *Orthopristis chrysoptera* (Linnaeus, 1766) [Haemulidae]; *Lutjanus griseus* (Linnaeus, 1758) [Lutjanidae]; *Prionotus tribulus* Cuvier, 1829 [Triglidae]; *Chaetodipterus faber* (Broussonet, 1782) [Ephippidae]; *Bairdiella chrysoura* (Lacepède, 1802), *Cynoscion arenarius* Ginsburg, 1930, *Menticirrhus littoralis* (Holbrook, 1847), *Menticirrhus americanus* (Linnaeus, 1758), *Menticirrhus saxatilis* (Bloch et Schneider, 1801) [Sciaenidae]; *Archosargus rhomboidalis* (Linnaeus, 1758), *Lagodon rhomboides* (Linnaeus, 1766) [Sparidae]; *Acanthostracion quadricornis* (Linnaeus, 1758) [Ostraciidae]; *Spherooides spengleri* (Bloch, 1785), *Spherooides testudineus* (Linnaeus, 1758) [Tetraodontidae].

All LWRs estimates were statistically significant ($P < 0.05$). New maximum lengths are reported for one species *Anchoa lamprotaenia* (12.2 cm SL). The scaled herring, *Harengula jaguana*, was the most abundant fish species (3769 specimens), followed by the broad-striped anchovy, *Anchoa hepsetus* (3559 specimens). However, even though the sampling continued for three years, a lower number of specimens was obtained (10–12 specimens) for some of the species (*Urobatis jamaicensis*, *Bagre marinus*, *Mugil curema*, *Strongylura timucu*, *Selene vomer*, *Rypticus maculatus*, *Prionotus tribulus*, *Chaetodipterus faber*), due to their low abundance and occurrence in these coastal ecosystems. Estimates of a and b for the LWRs, the coefficient of determination R^2 , and 95% confidence limits for b are given in Table 1. A negative allometric growth was recorded in 16 species, a positive allometric growth in 10 species, and isometric growth in 18 species.

Discussion

The coefficient of determination (R^2) ranged from 0.861 (*Anchoa hepsetus*) to 0.996 (*Albula vulpes*, *Archosargus rhomboidalis*). This low value is related to the high dispersion of the length data. It is important also to mention that the length range of specimens is not the only magnitude that influences the value of R^2 . Other factors such as the size of the sample, the length of the specimens, the gonad maturity, and diet are of importance. The exact relation between length and weight differs among species according to their inherited body shape, and within a species according to the condition (robustness) of individual fish. The condition fac-

Table 1. Length–weight relations for 44 species of the unprotected Yucatan Coastal Biological Corridor, Mexico.

Species	n	SL [cm]	Weight [g]	a	CI 95% a	b	CI 95% b	Growth type	R ²	Reference data	
										L _m [cm]	L _{max} [cm]
Urotrygonidae											
<i>Urobatis jamaicensis</i>	10	12.5–29.9	22.0–239.9	0.010	0.009–0.0011	3.000	—	I	—	20.0 _{TL,1}	76.0 _{TL}
Albulidae											
<i>Albula vulpes</i>	13	3.2–13.1	0.3–32.8	0.008	0.006–0.010	3.229	3.085–3.373	+A	0.996	21.0 _{FL}	104.0 _{TL}
Elopidae											
<i>Elops saurus</i>	20	12.7–31.5	24.1–196.5	0.007	0.006–0.008	3.000	—	I	—	32.5 _{SL}	100.0 _{TL}
Engraulidae											
<i>Anchoa hepsetus</i>	3559	3.4–6.7	0.4–2.4	0.019	0.018–0.020	2.508	2.475–2.542	–A	0.861	4.3 _{TL,2}	15.3 _{TL}
<i>Anchoa lamprotaenia</i>	360	2.9–12.2	0.12–34.5	0.005	0.005–0.006	3.315	3.278–3.352	+A	0.989	5.0 _{SL,3}	12.0 _{TL}
<i>Anchoa lyolepis</i>	39	3.9–6.3	0.5–2.2	0.009	0.008–0.009	3.000	—	I	—	8.2 _{SL,28}	12.0 _{TL}
<i>Anchoa mitchilli</i>	1232	2.3–6.1	0.1–2.4	0.009	0.009–0.010	2.999	2.944–3.055	–A	0.905	4.0 _{SL,4}	10.0 _{TL}
Dorosomatidae											
<i>Harengula jaguana</i>	3769	2.1–12.8	0.1–36.3	0.008	0.007–0.008	3.381	3.366–3.397	+A	0.979	8.0 _{SL,5}	21.2 _{TL}
<i>Opisthonema oglinum</i>	92	3.8–17.1	0.8–86.2	0.011	0.009–0.012	3.122	3.020–3.224	+A	0.976	11.5 _{FL}	38.0 _{TL}
Ariidae											
<i>Ariopsis felis</i>	1388	4.0–26.3	0.9–240.7	0.016	0.015–0.017	2.948	2.924–2.972	–A	0.977	15.0 _{SL,6}	70.0 _{TL}
<i>Bagre marinus</i>	12	7.2–15.3	5.0–50.2	0.015	0.014–0.016	3.000	—	I	—	32.8 _{FL,8}	100 _{TL,7}
Synodontidae											
<i>Synodus foetens</i>	52	3.9–41.4	0.4–166.0	0.016	0.011–0.022	2.751	2.620–2.882	–A	0.974	19.0 _{SL}	53.8 _{TL}
Batrachoididae											
<i>Opsanus beta</i>	23	4.5–10.4	1.9–24.8	0.012	0.007–0.019	3.301	3.032–3.571	+A	0.972	7.6 _{SL}	32.4 _{SL,9}
Mugilidae											
<i>Mugil curema</i>	10	2.0–23.9	0.1–153.4	0.016	0.014–0.018	3.000	—	I	—	16.4 _{TL,10}	91.0 _{TL}
<i>Mugil trichodon</i>	20	2.1–15.3	0.1–64.6	0.013	0.009–0.018	3.180	3.034–3.326	+A	0.991	16.0 _{FL}	46.0 _{TL}
Belonidae											
<i>Strongylura notata</i>	104	24.0–46.0	23.7–124.8	0.009	0.005–0.015	2.524	2.364–2.683	–A	0.909	22.6 _{TL}	61.0 _{TL}
<i>Strongylura timucu</i>	10	7.2–36.5	0.3–72.0	0.0012	0.001–0.0014	3.000	—	I	—	—	61.0 _{TL}
Hemiramphidae											
<i>Chriodorus atherinoides</i>	36	3.8–17.8	0.2–40.2	0.008	0.005–0.011	3.312	2.971–3.652	+A	0.933	—	26.0 _{TL}
<i>Hyporhamphus unifasciatus</i>	173	5.0–25.5	0.4–86.7	0.003	0.003–0.0033	3.000	—	I	—	18.5 _{FL,28}	30.0 _{TL}
Carangidae											
<i>Caranx latus</i>	14	7.0–14.9	7.5–82.0	0.021	0.020–0.022	3.000	—	I	—	37.0 _{FL}	101.0 _{FL}
<i>Oligoplites saurus</i>	28	2.2–23.8	0.1–145.3	0.010	0.010–0.011	3.000	—	I	—	19.8 _{SL,11}	42.5 _{SL,13}
<i>Selene vomer</i>	11	2.3–9.2	0.4–23.4	0.049	0.031–0.079	2.700	2.374–3.025	–A	0.982	24.1 _{TL,29}	48.3 _{TL}
<i>Trachinotus carolinus</i>	123	1.5–9.5	0.5–20.7	0.026	0.025–0.027	3.000	—	I	—	25.0 _{FL,12}	64.0 _{TL}
<i>Trachinotus falcatus</i>	491	2.0–14.5	0.4–104.1	0.045	0.042–0.049	2.850	2.800–2.900	–A	0.963	48.6 _{FL}	122.0 _{TL}
<i>Trachinotus goodei</i>	34	2.8–16.9	0.8–119.4	0.029	0.021–0.041	2.927	2.778–3.078	–A	0.983	26.0 _{TL,12}	50.0 _{TL}
Cynoglossidae											
<i>Symphurus plagiusa</i>	14	7.5–14.4	3.6–28.7	0.009	0.008–0.009	3.000	—	I	—	10.1 _{TL}	21.0 _{TL}
Gerreidae											
<i>Eucinostomus argenteus</i>	347	2.0–14.5	0.2–45.1	0.022	0.020–0.024	3.006	2.954–3.058	I	0.975	12.0 _{TL,14}	21.2 _{TL}
<i>Eucinostomus gula</i>	388	2.6–9.1	0.4–20.7	0.016	0.015–0.018	3.219	3.161–3.277	+A	0.970	9.0 _{FL,14}	25.5 _{TL}
<i>Eucinostomus harengulus</i>	19	5.5–8.2	3.0–12.5	0.021	0.019–0.022	3.000	—	I	—	12.0 _{SL}	15.0 _{SL}
Grammistidae											
<i>Rypticus maculatus</i>	10	6.4–8.9	5.2–13.6	0.019	0.018–0.020	3.000	—	I	—	8.9 _{TL,16}	24.0 _{TL,15}
Haemulidae											
<i>Orthopristis chrysoptera</i>	15	4.2–20.5	1.5–158.7	0.022	0.021–0.023	2.844	2.601–3.086	–A	0.992	20.0 _{SL}	46.0 _{FL}
Lutjanidae											
<i>Lutjanus griseus</i>	42	4.8–18.5	2.9–128.4	0.034	0.025–0.045	2.891	2.766–3.015	–A	0.984	18.0 _{SL,17}	89.0 _{TL}
Triglidae											
<i>Prionotus tribulus</i>	10	2.6–14.3	0.6–61.6	0.026	0.024–0.028	3.000	—	I	—	8.4 _{TL,18}	35.0 _{TL}
Ephippidae											
<i>Chaetodipterus faber</i>	10	2.8–7.4	1.5–23.8	0.064	0.060–0.068	3.000	—	I	—	9.9 _{TL,19}	91.0 _{TL}
Sciaenidae											
<i>Bairdiella chrysoura</i>	114	3.3–17.7	0.7–111.2	0.021	0.018–0.024	2.966	2.909–3.022	–A	0.990	9.1 _{SL,20}	30.0 _{TL}
<i>Cynoscion arenarius</i>	64	2.6–20.9	0.3–109.3	0.018	0.015–0.021	2.914	2.853–2.976	–A	0.994	14.0 _{SL,21}	63.5 _{TL}
<i>Menticirrhus littoralis</i>	69	2.6–15.7	0.2–63.2	0.014	0.012–0.017	2.943	2.856–3.031	–A	0.984	19.8 _{TL,23}	60.0 _{SL,22}
<i>Menticirrhus americanus</i>	104	2.4–14.8	0.2–57.3	0.010	0.009–0.012	3.149	3.093–3.206	+A	0.992	15.0 _{TL,24}	60.0 _{TL,25}
<i>Menticirrhus saxatilis</i>	57	2.5–19.0	0.4–102.2	0.014	0.012–0.017	2.997	2.908–3.086	–A	0.986	25.6 _{TL}	46.0 _{TL}

Table continues on next page.

Table 1. Continued.

Species	n	SL [cm]	Weight [g]	a	CI 95% a	b	CI 95% b	Growth type	R ²	Reference data	
										L _m [cm]	L _{max} [cm]
Sparidae											
<i>Archosargus rhomboidalis</i>	139	2.7–21.0	0.5–327.0	0.023	0.021–0.024	3.148	3.116–3.179	+A	0.996	8.0 _{SL}	33.0 _{TL}
<i>Lagodon rhomboides</i>	230	4.7–13.0	2.6–62.4	0.041	0.032–0.052	2.846	2.740–2.953	–A	0.929	8.0 _{SL,26}	40.0 _{TL}
Ostraciidae											
<i>Acanthostracion quadricornis</i>	16	12.9–21.7	79.9–283.6	0.033	0.031–0.035	3.000	—	I	—	19.8 _{TL}	55.0 _{TL}
Tetraodontidae											
<i>Sphoeroides spengleri</i>	19	3.8–6.3	1.7–6.9	0.029	0.028–0.030	3.000	—	I	—	18.8 _{SL,28}	30.0 _{TL}
<i>Sphoeroides testudineus</i>	110	2.3–20.0	3.8–378.7	0.055	0.039–0.077	2.880	2.753–3.008	–A	0.925	10.0 _{TL,27}	38.8 _{TL}

n = number of individuals, SL = standard length, TL = total length, a = intercept (equation parameter), b = slope (allometry coefficient), 95% CI = 95% confidence limits (for both equation parameters), R² = coefficient of determination, L_m = size at first maturity, L_{max} = maximum length. Species in bold denote new maximum length. I = isometric growth, –A = negative allometric growth, +A = positive allometric growth. Isometric growth is assumed in the species with low number of specimens and/or narrow range sizes (no value for 95% CI b) (Froese 2006; Hay et al. 2020). Length values without references subscripts comprise information from FishBase. Subscript references: 1 = Yáñez-Arancibia and Amezcua (1979), 2 = Munroe et al. (2015a), 3 = Munroe et al. (2015b), 4 = Vega-Cendejas et al. (2017), 5 = Munroe et al. (2019), 6 = Betancur (2015), 7 = Chao et al. (2015), 8 = Caballero-Chávez (2013), 9 = Collette et al. (2019), 10 = Yago-Bruno et al. (2020), 11 = Duque-Nivia et al. (1995), 12 = Alvarez-Lajonchere and Ibarra Castro (2012), 13 = Ospina-Arango et al. (2008), 14 = Mexicano-Cintora (1999), 15 = Anderson et al. (2015), 16 = Bullock and Smith (1991), 17 = Allen (1985), 18 = Hoff (1992), 19 = Soeth et al. (2018), 20 = Grammer et al. (2009), 21 = Nemeth et al. (2006), 22 = Chao et al. (2020a), 23 = Aloisio and Nelson (2004), 24 = Chao et al. (2020b), 25 = McEachran and Fechhelm (2005), 26 = Russell et al. (2014), 27 = Shao et al. (2014), 28 = Bouchon-Navaro et al. (2006); 29 = Becerra et al. (2013).

tor sometimes reflects food availability and growth within the weeks before sampling. But the condition is variable and dynamic. Individual fish within the same sample vary considerably, and the average condition of each population varies seasonally and yearly (Kuriakose 2017).

The exponent b presented a mean value of 2.997 (DE: 0.18) with values ranging from 2.508 estimated for *Anchoa hepsetus* to 3.381 for *Harengula jaguana*. The lower values may have resulted from the fact that the majority of the specimens analyzed were juveniles (<4.3 cm) due to their type of habitat (wetlands, petenes, swamps), while in the case of *H. jaguana* it is attributed to its maturity stage. The LWRs parameters of *Rypticus maculatus* (Grammitidae) and *Anchoa lamprotaenia* (Engraulidae) are herein published for the first time in both the scientific literature and databases, such as FishBase (Froese and Pauly 2023) (Table 1). A new maximum length was recorded for *A. lamprotaenia* (12.2 cm SL). Overall, LWRs were highly significant for all species ($P < 0.001$). Changes in b reflect mostly the species morphology and environmental factors such as temperature, salinity, food (quantity, quality, and size), sex, health, and developmental stage (Sparre 1992). In the case of *Sphoeroides testudineus* a (0.055) and b (2.880) were very similar to those previously reported in a hyperhaline coastal lagoon located near this unprotected coastal region (Vega-Cendejas et al. 2017). The number of explanatory variables considered in the model also conditions the value of this coefficient. Carlander (1977) demonstrated that values of $b < 2.5$ or > 3.5 are often derived from samples with narrow size ranges. The mean condition of specimens as well as the difference in condition between small and large specimens vary between season localities and years, resulting in different weight relations. The influence of extreme values of b on mean b decreases with the number of estimates (Froese 2006).

For species with low numbers or low size ranges (Carlander 1997) (*Urobatis jamaicensis*, *Elops saurus*, *Anchoa lyolepis*, *Bagre marinus*, *Mugil curema*, *Strongylura timucu*, *Chriodorus atherinoides*, *Hyporhamphus unifasciatus*, *Caranx latus*, *Oligoplites saurus*, *Trachinotus carolinus*, *Symphurus plagiusa*, *Eucinostomus harengulus*, *Rypticus maculatus*, *Prionotus tribulus*, *Chaetodipterus faber*, *Acanthostracion quadricornis*, *Sphoeroides spengleri*), LWRs were calculated assuming $b = 3.0$, being the value of the slope considered by the formula of Hay et al. (2020).

Conclusions

The results provided in this study can be very useful for the management of coastal ecosystems, including wetlands, which are required to maintain their diversity due to the increase in human activity in this unprotected coastal region (tourism, fisheries, habitat degradation). Additionally, this information is very useful for the development of trophic models using ECOPATH, which are of significant value in making predictions about the conservation status of this critical habitat for fishery and ecologically important species that use the ecosystem in the juvenile stage.

Acknowledgments

We are very grateful to the Comisión Nacional de Biodiversidad, México (CONABIO) for financial support for the research project (CONABIO-027) and to Alex Acosta, Walter Canto, Daniel Arceo, Víctor Castillo, Oscar Reyes, Orlando Cervantes, and M. Angel Villalobos for field assistance and support in sample processing.

References

- Allen GR (1985) Snappers of the world: An annotated and illustrated catalogue of lutjanid species known to date. FAO Fisheries Synopsis. FAO, Rome, 6, 207 pp.
- Aloisio SB, Nelson FF (2004) Reproductive biology of *Menticirrhus littoralis* in southern Brazil (Actinopterygii: Perciformes: Sciaenidae). Neotropical Ichthyology 2(1): 31–36. <https://doi.org/10.1590/S1679-62252004000100005>
- Alvarez-Lajonchere L, Ibarra-Castro L (2012) Relationships of maximum length, length at first sexual maturity, and growth performance index in nature with absolute growth rates of intensive cultivation of some tropical marine fish. Journal of the World Aquaculture Society 43(5): 607–620. <https://doi.org/10.1111/j.1749-7345.2012.00591.x>
- Anderson W, Carpenter KE, Gilmore G, Milagrosa Bustamante G, Polanco Fernandez A, Robertson R (2015) *Rypticus maculatus*. The IUCN Red List of threatened species 2015. e.T16759353A16781863. <https://doi.org/10.2305/IUCN.UK.2015-2.RLTS.T16759353A16781863.en>
- Becerra da Silva C, Araújo M, Vieira C (2013) Sustainability of capture of fish bycatch in the prawn trawling in northeastern Brazil. Neotropical Ichthyology 11(1): 133–142. <https://doi.org/10.1590/S1679-62252013000100016>
- Betancur R (2015) *Ariopsis felis*. The IUCN Red List of threatened species 2015: e.T190456A1952682. <https://doi.org/10.2305/IUCN.UK.2015-2.RLTS.T190456A1952682.en>
- Bouchon-Navaro Y, Bouchon C, Kopp D, Louis M (2006) Weight–length relationships for 50 fish species collected in seagrass beds of the Lesser Antilles. Journal of Applied Ichthyology 22(4): 322–324. <https://doi.org/10.1111/j.1439-0426.2006.00715.x>
- Bullock H, Smith GB (1991) Sea basses (Pisces: Serranidae). Memoirs of the Hourglass Cruises. Marine Research Laboratory, Florida Department of Natural Resources, St. Petersburg, Florida, 205 pp.
- Caballero-Chávez V (2013) Madurez y reproducción de bagre bandera *Bagre marinus* en el sudeste de Campeche. Ciencia Pesquera 21(2): 13–19.
- Carlander KD (1977) Handbook of freshwater fishery biology. Vol. 2. Life history data on centrarchid fishes of the United States and Canada. Iowa State University Press, Ames, IA, USA.
- Carpenter KE (Ed.) (2002a) The living marine resources of the Western Central Atlantic. Volume 2: Bony fishes part 1 (Acipenseridae to Grammatidae). FAO Species Identification Guide for Fishery Purposes and American Society of Ichthyologists and Herpetologists Special Publication No. 5. FAO, Rome, 601–1374.
- Carpenter KE (Ed.) (2002b) The living marine resources of the Western Central Atlantic: volume 3 -Bony fishes part 2 -(Opistognathidae to Molidae), sea turtles and marine mammals. FAO Species Identification Guide for Fishery Purposes and American Society of Ichthyologists and Herpetologists Special Publication No. 5. FAO, Rome, 1375–2127.
- Chao L, Vega-Cendejas M, Tolan J, Jelks H, Espinosa-Perez H (2015) *Bagre marinus*. The IUCN Red List of threatened species 2015: e.T196806A2476570. <https://doi.org/10.2305/IUCN.UK.2015-2.RLTS.T196806A2476570.en>
- Chao L, Espinosa-Perez H, Aguilera Socorro O, Haimovici M (2020a) *Menticirrhus littoralis*. The IUCN Red List of threatened species 2020: e.T46105545A82677213. <https://doi.org/10.2305/IUCN.UK.2020-2.RLTS.T46105545A82677213.en>
- Chao L, Espinosa-Perez H, Aguilera Socorro O, Haimovici M (2020b) *Menticirrhus americanus*. The IUCN Red List of threatened species 2020: e.T195075A82668543. <https://doi.org/10.2305/IUCN.UK.2020-2.RLTS.T195075A82668543.en>
- Collette BB, Aiken KA, Polanco Fernandez A, Vega-Cendejas M (2019) *Opsanus beta*. The IUCN Red List of threatened species 2019. e.T190257A86399458. <https://doi.org/10.2305/IUCN.UK.2019-2.RLTS.T190257A86399458.en>
- Duque-Nivia G, Acero AP, Santos-Martinez A (1995) Aspectos reproductivos de *Oligoplites saurus* y *O. palometa* (Pisces: Carangidae) en la Ciénega Grande de Santa Marta, Caribe Colombiano. Caribbean Journal of Science 31(3–4): 317–326.
- Euán-Avila J, García de Fuentes A, Liceaga Correa MA, Munguia Gil A (Eds.) (2014) La costa del Estado de Yucatán, un espacio de reflexión sobre la sociedad-naturaleza, en el contexto del ordenamiento territorial. Tomo II. Plaza y Valdés.
- Froese R (2006) Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations. Journal of Applied Ichthyology 22(4): 241–253. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- Froese R, Pauly D (Eds.) (2022) FishBase. [Version 04/2022] <http://www.fishbase.org>
- Froese R, Tsikliras AC, Stergiou KI (2011) Editorial note on weight–length relations of fishes. Acta Ichthyologica et Piscatoria 41(4): 261–263. <https://doi.org/10.3750/AIP2011.41.4.01>
- Grammer GL, Brown-Peterson NJ, Peterson MS, Comyns BH (2009) Life history of silver perch *Bairdiella chrysoura* (Lacepède, 1803) in north-central Gulf of Mexico estuaries. Gulf of Mexico Science 27(1): 62–73. <https://doi.org/10.18785/goms.2701.07>
- Hay A, Xian W, Bailly N, Liang C, Pauly D (2020) The why and how of determining length–weight relationships of fish from preserved museum specimens. Journal of Applied Ichthyology 36(3): 373–379. <https://doi.org/10.1111/jai.14014>
- Herrera-Silveira JA, Morales-Ojeda SM (2009) Evaluation of the health status of a coastal ecosystem in southeast Mexico: Assessment of water quality, phytoplankton and submerged aquatic vegetation. Marine Pollution Bulletin 59(1–3): 72–86. <https://doi.org/10.1016/j.marpolbul.2008.11.017>
- Hoff JG (1992) Comparative biology and population dynamics of searobins (genus *Prionotus*) with emphasis on populations in the north-western Gulf of Mexico. Dissertations, Theses, and Masters Projects. Paper 1539616697. <https://doi:10.25773/v5-c67f-dt79>
- Kohler N, Casey J, Turner P (1995) Length–weight relationships for 13 species of sharks from the western North Atlantic. Fish Bulletin 93: 412–418.
- Kuriakose S (2017) Estimation of length weight relationship in fishes. Summer School on Advanced Methods for Fish Stock Assessment and Fisheries Management. Lecture Note Series No. 2/2017. CMFRI; Kochi, Kochi, 215–220.
- McEachran JD, Feckhelm JD (1998) Fishes of the Gulf of Mexico, Volumen 1: Myxiniiformes to Gasterosteiformes. University of Texas Press, Austin, TX, USA.
- McEachran JD, Feckhelm JD (2005) Fishes of the Gulf of Mexico, Volumen 2: Scorpaeniformes to Tetraodontiformes. University of Texas Press, Austin, TX, USA. <https://doi.org/10.7560/706347>

- Mexicano-Cintora G (1999) Crecimiento y reproducción de la mojarra, *Eucinostomus gula* de Celestún, Yucatán, México. Proceedings of the Gulf and Caribbean Fisheries Institute 45: 524–536.
- Moran M, Monroe A, Stallcup L (2019) A proposal for practical and effective biological corridors to connect protected areas in north-west Costa Rica. Nature Conservation 36: 113–137. <https://doi.org/10.3897/natureconservation.36.27430>
- Munroe T, Aiken KA, Brown J, Grijalba Bendeck L (2015a) *Anchoa hepsetus*. The IUCN Red List of threatened species 2015: e.T16406327A16510237. <https://doi.org/10.2305/IUCN.UK.2015-4.RLTS.T16406327A16510237.en>
- Munroe T, Aiken KA, Brown J, Grijalba Bendeck L (2015b) *Anchoa lamprotaenia*. The IUCN Red List of threatened species 2015: e.T16406525A16509907. <https://doi.org/10.2305/IUCN.UK.2015-4.RLTS.T16406525A16509907.en>
- Munroe TA, Aiken KA, Brown J, Grijalba Bendeck L, Vega-Cendejas M (2019) *Harengula jaguana*. The IUCN Red List of threatened species 2019: e.T190478A86377366. <https://doi.org/10.2305/IUCN.UK.2019-2.RLTS.T190478A86377366.en>
- Nemeth DJ, Jackson JB, Knapp AR, Purtlebaugh CH (2006) Age and growth of sand seatrout (*Cynoscion arenarius*) in the estuarine waters of the eastern Gulf of Mexico. Gulf of Mexico Science 24(1): 45–60. <https://doi.org/10.18785/goms.2401.07>
- Ospina-Arango JF, Pardo-Rodríguez FI, Álvarez-León R (2008) Madurez gonadal de la ictiofauna presente en la Bahía de Cartagena, Caribe Colombiano. Boletín Científico Centro de Museo de Historia Natural 12: 117–140.
- Palacios-Sánchez SE, Vega-Cendejas ME, Hernández-de-Santillana M, Aguilar-Medrano R (2019) Anthropogenic impacts in the nearshore fish community of the Yucatan Coastal Corridor. A comparison of protected and unprotected areas. Journal for Nature Conservation 51: 125721. <https://doi.org/10.1016/j.jnc.2019.125721>
- Pendoley KL, Schofield G, Whittock PA, Ierodiaconou D, Hays GC (2014) Protected species use of a coastal marine migratory corridor connecting marine protected areas. Marine Biology 161(6): 1455–1466. <https://doi.org/10.1007/s00227-014-2433-7>
- Petrakis G, Stergiou KI (1995) Weight–length relationships for 33 fish species in Greek waters. Fisheries Research 21(3–4): 465–469. [https://doi.org/10.1016/0165-7836\(94\)00294-7](https://doi.org/10.1016/0165-7836(94)00294-7)
- Russell B, Carpenter KE, MacDonald T, Vega-Cendejas M (2014) *Lagodon rhomboides*. The IUCN Red List of threatened species 2014. e.T170250A1301642. <https://doi.org/10.2305/IUCN.UK.2014-3.RLTS.T170250A1301642.en>
- Shao K, Matsuura K, Leis JL, Hardy G, Larson H, Liu M (2014) *Sphoeroides testudineus*. The IUCN Red List of threatened species 2014: e.T193813A2281154. <https://doi.org/10.2305/IUCN.UK.2014-3.RLTS.T193813A2281154.en>
- Soeth M, Fávoro LF, Spach HL, Daros FA, Woltrich AE, Correia AT (2018) Age, growth, and reproductive biology of the Atlantic spadefish *Chaetodipterus faber* in southern Brazil. Ichthyological Research 66(1): 140–154. <https://doi.org/10.1007/s10228-018-0663-2>
- Sparre P (1992) Introduction to tropical fish stock assessment. Part I. Manual. FAO Fishery Technical Paper. 306/1. Rev. I, FAO, Rome.
- Teixeira-de Mello F, Eguren G (2008) Prevalence and intensity of black-spot disease in fish community from Cañada del Dragón stream (Montevideo, Uruguay). Limnetica 27: 251–258. <https://doi.org/10.23818/limn.27.20>
- Teixeira-de Mello F, Iglesias C, Borthagaray A, Mazzeo N, Vilches J, Larrea D, Ballabio R (2006) Ontogenetic allometric coefficient changes. Implication of diet shift and morphometric attributes in *Hoplias malabaricus* (Bloch) (Characiformes, Erythrinidae). Journal of Fish Biology 69(6): 1770–1778. <https://doi.org/10.1111/j.1095-8649.2006.01245.x>
- Turk-Boyer P, Morzaria-Luna H, Martínez-Tovar I, Downton-Hoffmann C, Munguia-Vega A (2014) Ecosystem-based fisheries management of a biological corridor along the northern Sonora coastline (NE Gulf of California). Pp. 125–154. In: Amezcua F, Bellgraph B (Eds.) Fisheries management of Mexican and Central American Estuaries. Estuaries of the World. Springer, Dordrecht, Netherlands. https://doi.org/10.1007/978-94-017-8917-2_9
- Vega-Cendejas ME, Peralta-Meixuero MA, Hernández de SM (2017) Length–weight relationships of fishes that inhabit a hyperhaline coastal lagoon: Ria Lagartos, Yucatan, Mexico. Acta Ichthyologica et Piscatoria 47(4): 411–415. <https://doi.org/10.3750/AIEP/02239>
- Wotton RJ (1990) Ecology of teleost fishes. Chapman and Hall, London, UK. <https://doi.org/10.1007/978-94-009-0829-1>
- Xie JY, Kang ZJ, Yang J, Yang DD (2015) Length–weight relationships for 15 fish species from the Hunan Hupingshan National Nature Reserve in central China. Journal of Applied Ichthyology 31(1): 221–222. <https://doi.org/10.1111/jai.12465>
- Yago-Bruno SN, Mariana Barros A, Jailza F, Jackellynne FF, Ladilson Rodrigues S, Marina BF (2020) Length at first sexual maturity of economically important fishes in the Brazilian Northeast Coast. Ocean and Coastal Research 68: e20311. <https://doi.org/10.1590/s2675-28242020068311>
- Yáñez-Arancibia A, Amezcua LF (1979) Ecología de *Urolophus jamaicensis* (Cuvier) en Laguna de Terminos un sistema estuarino del sur del Golfo de México (Pisces: Urolophidae). Anales del Centro de Ciencias del Mar y Limnología. Universidad Nacional Autónoma de México 6(2): 123–13.