

Sexual size dimorphism in the extreme SW breeding population of the European Storm Petrel *Hydrobates pelagicus* (Aves: Procellariiformes)

GUILLERMO DELGADO CASTRO^{1*}, JUAN D. DELGADO², JAVIER GONZÁLEZ³ & MICHAEL WINK³

¹ Museo de Ciencias Naturales, Organismo Autónomo de Museos y Centros, Antiguo Hospital Civil, Calle Fuente Morales s/n, 38003 Santa Cruz de Tenerife, Canary Islands, Spain — ² Área de Ecología, Dept. Sistemas Físicos, Químicos y Naturales, Univ. Pablo de Olavide, Ctra. de Utrera, km 1, 41013 Seville, Spain — ³ Institut für Pharmazie und Molekulare Biotechnologie, Ruprecht-Karls-Universität Heidelberg, Im Neuenheimer Feld 364, 69120 Heidelberg, Germany — * Corresponding author: Guillermo Delgado Castro; guille(at)museosdetenerife.org

Accepted 01.xi.2013.

Published online at www.senckenberg.de/vertebrate-zoology on 18.xii.2013.

Abstract

We used molecular sexing and morphological analysis to characterise sexual size dimorphism (SSD) in the European Storm Petrel *Hydrobates pelagicus* (LINNAEUS, 1758) in the southernmost and westernmost breeding colony (El Hierro, Canary Islands). We analysed SSD in 79 individuals (46 males, 33 females). We performed discriminant analysis on 9 morphometric variables and correctly classified 70.5% of the original cases. By binary logistic regression, the accuracy in sexing birds previously sexed by DNA analysis was 70.5%. Females were larger than males in wing and tarsus length but there were no differences for bill size and shape. There was a large biometric overlap between sexes in this Canarian breeding colony of *H. pelagicus*. However, a moderate female-biased dimorphism in flight-relevant variables such as tail and wing traits could be of interest in trophic segregation of sexes and optimization of resource use in these pelagic-feeding birds.

Key words

Canary Islands, character variation, discriminant analysis, female-biased SSD, geographical range, *Hydrobates pelagicus*.

Introduction

Sexual size dimorphism (SSD) is a widespread feature in many bird species (ANDERSON 1994, LEZANA *et al.* 2000, CAMPOS *et al.* 2005). In most bird taxa, males are of the same size, slightly larger, or significantly larger than females (PRICE 1984) and it is usually more pronounced in aggressive and predatory species (THIOLLAY 1994). In raptors, females are larger and in extreme cases may have twice the weight of the males (THIOLLAY 1994). By contrast, in some terrestrial, cursorial birds such as bustards, females are much lighter and smaller (COLLAR 1996). The evolution of SSD has probably occurred through

sexual selection together with promiscuous mating (LACK 1971), reflecting differences in sex-specific reproductive roles and responses to environmental factors (BLONDEL *et al.* 2002, GONZÁLEZ-SOLÍS 2004), feeding behaviour (DE JUANA *et al.* 2004), trophic niche segregation of sexes and sexual selection (GUTIÉRREZ-CORCHERO *et al.* 2007, NAVARRO *et al.* 2009).

Pelagic seabirds exhibit SSD. Among the Procellariidae, females are generally slightly smaller and more lightly built than males (RISTOW & WINK 1980, CARBONERAS 1992, GONZÁLEZ-SOLÍS *et al.* 2000). However, Storm pet-

rels (Hydrobatidae) show female-biased SSD (WARHAM 1990), along with Sulidae and Fregatidae, whereas Phaetontidae show no apparent SSD (SERRANO-MENESES & SZÉKELY 2006).

Several methods have been used to sex bird species which are monomorphic in plumage. Molecular sexing consists of a non-intrusive technique and the most popular, requiring only a small amount of DNA from blood or a single feather (TABERLET & BOUVET 1991, RISTOW & WINK 2004, SUTHERLAND *et al.* 2004, INFANTE & PERIS 2004). For many species of birds including Procellariiformes, sexing has been achieved by discriminant analysis using morphological measurements (RISTOW & WINK 1980, COULTER 1986, LO VALVO 2001, BUTLER & GOSLER 2004, ALARCOS *et al.* 2007).

The European Storm Petrel *Hydrobatas pelagicus* (LINNAEUS, 1758) is a small pelagic seabird species distributed in the NE Atlantic from S Iceland to NW France, N Spain, Salvages and Canary Islands; it is also present in the Mediterranean from N Spain to Greece (DEL HOYO *et al.* 1992). *Hydrobatas pelagicus* has been considered a monotypic species although some authors suggest that the Mediterranean population must be re-established as a valid subspecies (*H. p. melitensis* SCHEMBRI, 1843). This distinction has been supported not only by biometrics (HEMERY & D'ELBEE 1985, MASSA & CATALISANO 1986, CATALISANO *et al.* 1988, LALANNE *et al.* 2001) but also by genetics (CAGNON *et al.* 2004). The Canary Islands correspond to the southern limit of the European Storm Petrel distribution range and represent the only known nesting place among the Macaronesian archipelagos. On the Canary Islands, a minimum of 1,000 breeding pairs of *H. pelagicus* have been estimated occupying small islets and rocks such as Alegranza (N of Lanzarote) and Roque Grande de Salmor, a rocky islet off El Hierro (NOGALES *et al.* 1993). Like other pelagic seabird species, *H. pelagicus* is a colonial breeder nesting in caves or small crevices, under heaps of boulders or under collapses of coastal cliffs (DELGADO *et al.* 1985, NOGALES *et al.* 1993).

Geographical variation in morphology and ethology has great importance in species-specific recognition (BRETAGNOLLE 1989). Variation underlying speciation processes is methodologically difficult to delimit in these marine birds (BOLTON & THOMAS 2001, CAGNON *et al.* 2004 and references there in). Morphological differences have been found for *H. pelagicus* even within the Mediterranean Basin (LALANNE *et al.* 2001), where there is, however, low genetic variation. Genetic differentiation is higher among the Atlantic populations. In addition, there is strong evidence of a lack of genetic exchange between Mediterranean and Atlantic populations.

Among the Atlantic Ocean populations, movements of Canarian *H. pelagicus* are poorly known. There have been records of birds reaching South Africa, and the islands would receive European migrants (MARTÍN & LORENZO 2001). Despite the wide distribution of *H. pelagicus*, the large population size of this species (130,000–290,000 pairs after Carboneras 1992; 130,000–400,000 pairs after FRANCO *et al.* 2004), and the morphological vari-

ation throughout the distribution range, biometric data characterizing live specimens are scarce. There is to our knowledge no available biometric work from the breeding colonies of the Canary Islands. Moreover, because of *post mortem* shrinkage of museum skins (KUCZYNSKI *et al.* 2003) our data from live birds become especially relevant. Here we assess SSD through molecular sexing and biometry, and analysed external morphological variation between sexes of the European Storm Petrel from its southern- and westernmost known breeding colony.

Study site

During the summer of 2007, field work was performed in Roque Grande de Salmor, a rocky islet of 0.03 km² (NW El Hierro island; 27°49' N 17°59' W, Canary Islands; Fig. 1). This is one of the largest colonies in the Canaries and the largest of El Hierro, with up to 200–300 pairs (NOGALES *et al.* 1993, MARTÍN & LORENZO 2001).

Methods

Biometrics

A total of 79 individuals of *H. pelagicus* (46 males and 33 females) were caught by mist nets. Birds were ringed with conventional metal rings and no tape lures were used. All the biometric measurements were taken by G. DELGADO CASTRO.

Wing length (maximum chord) was taken using a stopped rule (SVENSSON 1992) to 1 mm. Tail length was measured using an unstopped rule to the nearest millimetre. Following HEMERY & D'ELBEE (1985), five morphological variables of bill were measured to 0.1 mm using digital callipers: bill length (along the dorsal mid-line from the edge of the feathers at the base of the culmen to the most distant part of the hook), bill depth 1 (BD1, mandibles + narines), bill depth 2 (BD2, mandibles at level of the opening of narines), narine width (BW1), bill width (BW2, mandibles at level of the opening of narines) and “maximum” tarsus length (strictly, tarsometatarsus length; SUTHERLAND *et al.* 2004). Body mass was estimated using a 50 g Pesola balance to 0.1 g.

Molecular sexing methods

Blood samples were collected by venipuncture of the brachial vein. Total DNA was isolated from 100 µl of

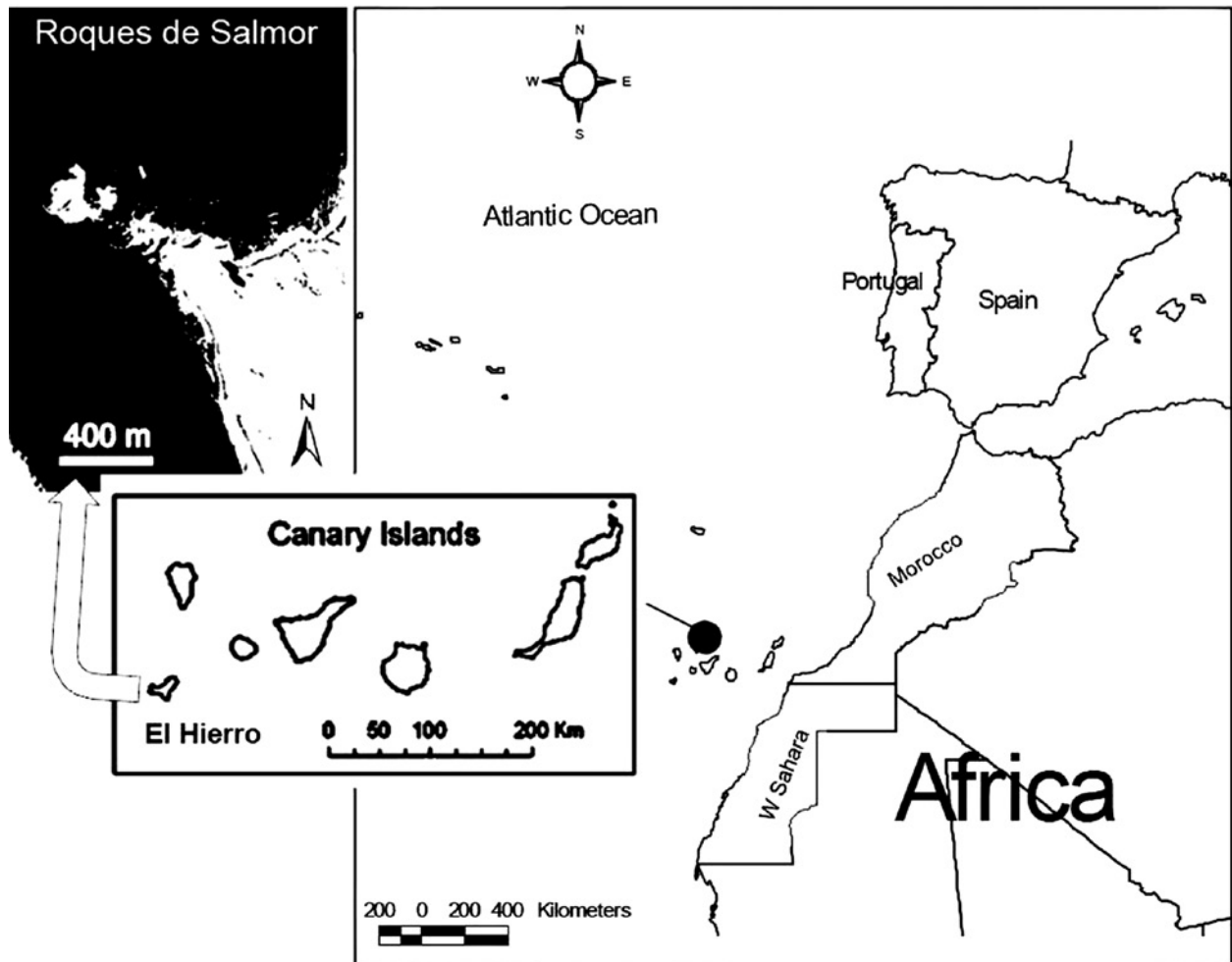


Fig 1. Study area. Location of the breeding colony of *Hydrobates pelagicus* studied in El Hierro, Canary Islands.

blood using standard Proteinase K (Merck, Darmstadt) and phenol/chloroform procedures (SAMBROOK *et al.* 1989). Sex identification was conducted following KAHN *et al.* (1998). PCR was performed with 30–60 ng of template DNA in 25 μ l reaction volume containing 8 pmol of the primer H1272 and 9 pmol of the primer L1237, 0.1 mM of dGTP, dCTP, and dTTP, 0.045 mM dATP, 1 μ Ci [α - 33 P]-dATP (AmershamBiosciences), 0.6 units of *Taq*-Polymerase (Pharmacia Biotech, Freiburg) and 2.5 μ l of 10 \times amplification buffer [10 mM Tris-HCl (pH 8.5), 50 mM KCl and 1.5 mM MgCl $_2$]. Each reaction was overlaid with two drops of mineral oil. Thermo cycling was performed with a Trio Thermo block TB1 (Biometra, Göttingen). Following the initial 5 min denaturation at 94°C, the program consisted of 31 cycles of 30 s at 94°C, 40 s at 56°C, 40 s at 72°C and 5 min at 72°C for final elongation. DNA fragments were separated by vertical PAGE (Polyacrylamide Gel Electrophoresis; length 40 cm) for 2 h at 65 W using a Base Acer Sequencer (Stratagene). After drying, the denaturing gels were exposed for 24 h to X-ray films (BioMax MR Film, Kodak). The bands were analyzed visually. The presence of two bands was scored as female and one band as male (KAHN *et al.* 1998).

Data analysis

A Kolmogorov-Smirnoff test was used to check for normality of variables. We used a Student t-test to analyse biometric differences between the sexes. For non-normal variables (wing length, tail length and mass) we used a non-parametric Mann-Whitney test. A Principal Component Analysis (PCA) was carried out in order to find a non-redundant pool of explanatory variables for biometrics of *Hydrobates pelagicus*. We applied the varimax rotation to achieve a better interpretation of the data pool. Discriminant analysis (SOKAL & ROLF 1995) was used to generate a function which is a linear combination of the morphometric variables measured from individuals of known sex, and to maximize the variance between groups (i.e. sexes). The method applied for calculating the discriminant function was Wilk's Lambda, and the probability for including a predictor variable in the model was set to 0.05 (0.1 for exclusion). Finally, a binary logistic regression function was calculated. All statistical analyses were performed using SPSS 12.0.

Table 1. Population biometrics. Biometrics for the European Storm Petrel at Roque Grande de Salmor (NE El Hierro). Data are: means \pm standard deviation (range), sample size: n. All data in mm except mass (g).

	Wing length	Bill length	Bill depth 1	Bill depth 2	Bill width 1	Bill width 2	Tail length	Tarsus length	Mass
Females	118.9 \pm 2.17 (113–123) n=33	11.43 \pm 0.54 (10.23–12.6) n=33	5.16 \pm 0.24 (4.76–5.73) n=33	3.70 \pm 0.18 (3.32–4.16) n=33	2.51 \pm 0.18 (2.06–2.78) n=33	2.25 \pm 0.19 (1.78–2.63) n=33	55.86 \pm 1.92 (53.5–61.5) n=33	24.19 \pm 0.76 (22.1–25.6) n=33	24.3 \pm 2.74 (20–30.5) n=33
Males	116.7 \pm 2.16 (110–120) n=46	11.26 \pm 0.56 (9.58–12.46) n=46	5.15 \pm 0.27 (4.57–5.78) n=46	3.76 \pm 0.18 (3.39–4.2) n=46	2.41 \pm 0.22 (2.0–2.97) n=46	2.26 \pm 0.21 (1.96–2.98) n=46	54.41 \pm 2.20 (49.5–58.5) n=45	24.09 \pm 0.81 (22.0–25.8) n=46	23.1 \pm 1.87 (20.0–28.5) n=46
All	117.6 \pm 2.77 (110–123) n=79	11.33 \pm 0.56 (9.58–12.60) n=79	5.16 \pm 0.26 (4.57–5.78) n=79	3.74 \pm 0.19 (3.32–4.2) n=79	2.47 \pm 0.22 (2.0–2.97) n=79	2.26 \pm 0.21 (1.78–2.98) n=79	55.03 \pm 2.20 (49.5–61.5) n=78	24.13 \pm 0.8 (22.0–25.8) n=79	23.6 \pm 2.35 (20.0–30.5) n=79

Table 2. Biometrics for live European Storm Petrels from different breeding colonies including Roque Grande de Salmor (Canary Islands). **a**=JAMES (1983); **b**=HEMERY & D'ELBEE (1985); **c**=LALANNE *et al.* (2001); **d**=present study. All data in mm except mass (g). **x**=mean; **s.d.**=standard deviation. Sample size: n. See Methods: Biometrics for explanation of codes.

	Great Britain (a)						Atlantic (b)			Biarritz (c)			Corse (c)			Salmor (d)		
	$\sigma\sigma$			$\varphi\varphi$			All sexes			All sexes			All sexes			All sexes		
	x	s.d.	n	x	s.d.	n	x	s.d.	n	x	s.d.	n	x	s.d.	n	x	s.d.	n
Wing	120.4	2.5	26	123.7	2.7	20	122.3		19	122.5	2.82	19	123.58	2.98		117.6	2.41	79
BL	12.0	0.4	26	12.1	0.7	20							11.99	0.71		11.33	0.56	79
BD1							5.28		19	5.29	0.29	19	5.56	0.22		5.16	0.26	79
BD2							3.86		19	3.87	0.42	19	4.07	0.17		3.74	0.19	79
BW							3.03		19							2.47	0.22	79
BW2							2.15		19							2.26	0.21	79
Tarsus	22.6	0.8	26	22.9	0.7	20										24.13	0.80	79
Tail	54.2	1.6	26	56.5	1.8	20	53.28		19							55.03	2.2	78
Mass							28.39		19	28.32	1.70	19	28.94	2.97		23.62	2.35	79

Results

Biometrics

The placement of males and females in the ordination space showed no clear separation of sexes (Fig. 2). Sex ratio in adults differed significantly from unity ($\chi^2_1=9.0$, $P<0.01$). Mean values for confirmed males and females are presented in Table 1. Between the sexes, significant differences were found for wing length ($U=340.5$, $P<0.001$) and tail length ($U=478.5$, $P=0.007$). No significant differences were observed for the other biometric measurements (i.e. bill length: $t=-1.44$, $df=77$, $P=0.166$; BD1: $t=0.055$, $df=77$, $P=0.956$; BD2 $t=1.224$, $df=77$, $P=0.225$; BW1: $t=-1.650$, $df=77$, $P=0.103$; BW2: $t=0.118$, $df=77$, $P=0.986$; tarsus length: $t=-0.661$, $df=77$, $P=0.511$; mass: $U=563.0$, $P=0.05$). Female wing was, on average, 2.2 mm longer than that of males, and females also showed longer tails, 1.45 mm on average, than did males. Further information is summarized in Table 2 where data of JAMES (1983), HEMERY & D'ELBEE

(1985), and LALANNE *et al.* (2001) from other Atlantic and Mediterranean (Corsican) colonies are included.

The PCA analysis shows that a set of three factors accounted for 48.82% of the total variation (Fig. 2; Table 3). The first factor (PC1) was related to bill width 1 (BW1), bill length and tarsus. The second factor (PC2) involved the wing length, tail length and weight, and the third (PC3) was related to bill depth 1 (BD1) and bill depth 2 (BD2). The placement of males and females in the ordination space showed no clear separation of sexes (Fig. 2).

Sexing

The discriminant linear model based on the original combination of 9 morphometric variables was highly significant (Wilk's Lambda=0.752; $F=12.361$, $P<0.001$) and included wing length and tail length as the best discriminators (Tables 4 and 5). The discriminant analysis correctly classified 70.5% of the original cases.

A binary logistic regression function was calculated to predict sex from a full set of variables ($n=79$ individu-

Table 3. PCA on biometric variables. Results of the Principal Component Analysis (PCA) on biometric measurements of *Hydrobates pelagicus* in Roque de Salmor, El Hierro (Canary Islands). Minor scores for each component and each variable have not been included. See Methods: Biometrics for explanation of codes.

	PC1	PC2	PC3
BW1	0.675		
Bill length	0.610		
Tarsus	0.575		
Winglength		0.722	
Tail		0.680	
BW2		-0.502	
Weight		0.410	
BD1			0.735
BD2			0.730
Eigenvalues	1.744	1.392	1.259
% variance	19.382	15.462	13.985
Cumulative % variance	19.382	34.844	48.829

Table 5. Correlations between variables and functions. Correlations between the discriminant variables and the canonical discriminant function.* Variables not included in the analysis; correlations of higher order are shown in bold.

Variable	Correlation coefficient
Wing length	0.879
Tail length	-0.606
Tarsus	0.134
Weight	0.132
An1*	0.122
An2*	-0.105
LTP*	-0.063
AL1*	-0.042
AL2*	0.021

Table 4. Discriminant-functions coefficients. Coefficients of the discriminant functions for both sexes and of the final canonical discriminant functions for the model (CDF).

	Sex		CDF
	Male	Female	
Wing length	34.61	35.04	0.37
Tail length	661.21	658.55	-2.31
Constant	-10751.40	-10732.02	17.26

als). Backwards deletion of the variables was used so that only those elements that were significant ($P < 0.05$) were retained. This generated a significant binary logistic regression function (Chi-square = 23.27, $df = 2$, Nagelkerke $R^2 = 0.347$, $P < 0.001$), including only wing and tail length as significant parameters:

$$X = 0.49 (\text{wing length}) - 2.87 (\text{tail length}) + 18.07.$$

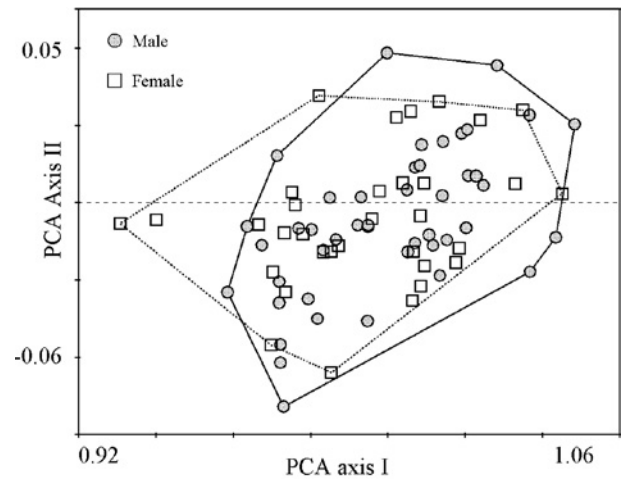


Fig. 2. PCA biometrics diagram. Biplot of a PCA run on the measured set of biometrical variables, with samples (individuals) separated by sex.

Discussion

We found that sexing *H. pelagicus* from the Canary Islands by using simple morphometric measurements renders a low degree of accuracy. There is a high degree of overlap between sexes in evolutionarily relevant biometrics such as bill size and shape. Also, as in the few studies published, we found a moderate female-biased sexual size dimorphism. Our results suggest the existence of large geographical variation in biometric parameters of *H. pelagicus*. Mean wing length of individuals caught at different North Atlantic breeding colonies have been reported to range between 116.7 and 123.7 mm (Table 2). Data regarding weight show similar trends (FURNESS & BAILLIE 1981, and references therein). Nevertheless, FURNESS & BAILLIE (1981) showed that storm petrel breeding on St. Kilda (Scotland) presented longer wings than wanderer birds, an observation which is also supported by FOWLER *et al.* (1986) at Yell (Shetland), where statistical differences were found between breeding and non-breeding birds. Although a proportion of birds at Roque Grande de Salmor may be non-breeding birds (at least 70% of the birds caught regurgitated food on nets), these were clearly lighter (Table 1). The Mediterranean birds are the larger ones with a wing length range of 129.25–123.58 mm. The tail length is quite similar between Atlantic and Mediterranean populations (Table 2).

Accuracies in sex assignment vary widely in the literature (Appendix 1). In studies using discriminant analysis for larger species, similar degrees of accuracy have been either enough (i.e. Tundra Swan, 74.1%; MILLER *et al.* 1988) or inadequate (i.e. Tengmalm's Owl, 70%; HIPKISS 2007) depending on season, bird development stage, age and the methods, among other factors (see Appendix 1).

Our values are also smaller than those above 92% found in *Larus argentatus* (FOX *et al.* 1981) and *Puffinus tenuirostris* (EINODER *et al.* 2008). LO VALVO (2001) found that bill depth and body mass were the best discriminants (92 and 84% of correctness) for *Calonectris diomedea* in Sicily. The values presented by our *Hydrobates* population were lower than those of JAMES (1983), where a discriminant function based on the same variables accounted for 88.5% and 80.0% for males and females, respectively. We achieved the best separation between sexes by using the wing and tail discriminant functions.

Our results on SSD in the European Storm Petrel support findings of previous works with biometrics from live birds (JAMES 1983). Tail and wing were larger in females than in males in this El Hierro breeding colony. Other biometric variables showed no statistically significant differences. For example, bill size and shape did not differ between sexes in this population of *H. pelagicus*. Bill size and shape is one of the most relevant sexually dimorphic variables with repercussions in foraging, sexual selection and speciation in seabirds (e.g. NAVARRO *et al.* 2009). In this population, however, *H. pelagicus* showed a slight departure from monomorphy (the degree to which sexes are of equal size) in tail and wing size, with probable effects in resource partition between sexes and intersexual competition (FIGUEROLA 1999). In *H. pelagicus*, the variables involved in dimorphism are directly related to flight performance, which might be relevant for foraging optimization in these pelagic-feeding birds. Furthermore, the observed intersexual variation may have repercussions in the optimization of intraspecific resource use (NAVARRO *et al.* 2009). The influence of low SSD in aspects of evolutionary biology and conservation of *H. pelagicus* such as assortative mating and mating success, phylogeny, foraging strategies, namely differential niche exploitation, especially when sea productivity fluctuates due to drastic environmental change, or sexual selection, deserves further research.

Acknowledgements

We are grateful to Gobierno de Canarias and the Cabildo Insular de El Hierro for authorising the collection of biological samples. We also thank Eduardo García del Rey for assisting with the field work. We thank Hedi Sauer-Gürth for technical assistance in the laboratory.

References

- ALARCOS, S., DE LA CRUZ, C., SOLÍS, E., VALENCIA, J. & GARCÍA-BAQUERO, M.J. (2007): Sex determination of Iberian Azure-winged Magpies *Cyanopica cyanuscooki* by discriminant analysis of external measurements. – *Ringling & Migration*, **23**: 211–216.
- ANDERSON, M. (1994): Sexual size dimorphism. In: KREBS, J.R., CLUTTON-BROCK, T. (eds.). *Sexual Selection. – Monographs in Behaviour and Ecology.* – Princeton University Press, Princeton, New Jersey. Pp. 246–293.
- BLONDEL, J., PERRET, P., ANSTETT, M.C. & THÉBAUD, C. (2002): Evolution of sexual size dimorphism in birds: test of hypotheses using blue tits in contrasted Mediterranean habitats. – *Journal of Evolutionary Biology*, **15**: 440–450.
- BOLTON, M. & THOMAS, R. (2001): Moulting and ageing of Storm Petrels *Hydrobates pelagicus*. – *Ringling & Migration*, **20**: 193–201.
- BOURGEAIS, K., CURE, C., LEGRAND, J., GÓMEZ-DÍAZ, E., VIDAL, E., AUBIN, T. & MATHEVON, N. (2007): Morphological versus acoustic analysis: what is the most efficient method for sexing yellow shearwaters *Puffinus yelkouan*? – *Journal of Ornithology*, **148**: 261–269.
- BRETAGNOLLE, V. (1989): Calls of the Storm Petrel *Hydrobates pelagicus*. – *Bioacoustics*, **2**(2): 166–167.
- BUTLER, C.J. & GOSLER, A. (2004): Sexing and ageing Rose-ringed Parakeets *Psittacula krameri* in Britain. – *Ringling & Migration*, **22**: 7–12.
- CAGNON, C., LAUGA, B., HÉMERY, G. & MOUCHÈS, C. (2004): Phylogeographic differentiation of storm petrels (*Hydrobates pelagicus*) based on cytochrome *b* mitochondrial DNA variation. – *Marine Biology*, **145**: 1257–1264.
- CAMPOS, F., HERNÁNDEZ, M., ARIZAGA, J., MIRANDA, R. & AMEZCUA, A. (2005): Sex differentiation of Corn Buntings *Miliaria calandra* in Northern Spain. – *Ringling & Migration*, **22**: 159–162.
- CARBONERAS, C. (1992): Family Procellariidae (Petrels and Shearwaters). – In: DEL HOYO, J., ELLIOTT, A., SARGATAL, J. (eds.): *Handbook of the Birds of the World. Vol. 1. Ostrich to Ducks.* – Lynx Editions, Barcelona. Pp. 216–257.
- CATALISANO, A., LO VALVO, F., LO VERDE, G. & MASSA, B. (1988): Dati biometrici dell'ucello delle tempeste (*Hydrobates pelagicus*). – *Naturalista siciliano*, **S IV, XII (suppl.)**, 261–265.
- COLLAR, N. (1996): Family Otidae (Bustards). – In: DEL HOYO, J., ELLIOTT, A. & SARGATAL, J. (eds.): *Handbook of the Birds of the World. Vol. 3. Hoatzin to Auks.* – Lynx Editions, Barcelona. Pp. 240–273.
- COULTER, M.C. (1986): Assortative mating and sexual dimorphism in the Common Tern. – *Wilson Bulletin*, **98**(1): 93–100.
- DEL HOYO, J., ELLIOTT, A. & SARGATAL, J. (1992): *Handbook of the Birds of the World. Vol. 1. Ostrich to Ducks.* – Lynx Edicions, Barcelona, 696 pp.
- DE JUANA, E., SUÁREZ, F. & RYAN, P.G. (2004): Family Alaudidae. – In: DEL HOYO, J., ELLIOTT, A., CHRISTIE, D.A. (eds.): *Handbook of the Birds of the World. Vol. 9. Cotingas to Pipits and Wagtails.* – Lynx Edicions, Barcelona. Pp. 496–541.
- DELGADO, G., HERNÁNDEZ, E., MARTÍN, A., TRUJILLO, O. & DÍAZ, G. (1985): Datos preliminares sobre la distribución y reproducción del Paño Común (*Hydrobates pelagicus*) en las Islas Canarias. – *Asturnatura*, **4**: 19–20.
- DONOHUE, K.C. & DUFTY, A.M. (2006): Sex determination of Red-tailed Hawks (*Buteo jamaicensis calurus*) using DNA analysis and morphometrics. – *Journal of Field Ornithology*, **77**(1): 74–79.
- EINODER, L.D., PAGE, B. & GOLDSWORTHY, S.D. (2008): sexual size dimorphism and assortative mating in the Short-tailed shearwater *Puffinus tenuirostris*. – *Marine Ornithology*, **36**: 167–173.

- FIGUEROLA, J. (1999): A comparative study on the evolution of reversed size dimorphism in monogamous waders. – *Biological Journal of the Linnean Society*, **67**: 1–18.
- FOWLER, J.A., HULBERT, M.E. & SMITH, G. (1986): Sex ratio in a sample of tape-lured Storm Petrel *Hydrobates pelagicus* from Shetland, Scotland. – *Seabird*, **9**: 15–19.
- FOX, G.A., COOPER, C.R. & RYDER, J.P. (1981): Predicting the sex of herring gulls by using external measurement. – *Journal of Field Ornithology*, **52**(1): 1–9.
- FRANCO, J., ETXEZARRETA, J., GALARZA, A., GOROSPE, G. & HIDALGO, J. (2004). Seabird populations. – In: BORJA, A. & COLLINS, M. (eds.): *Oceanography and marine environment of the Basque Country*. – Amsterdam: Elsevier Oceanography Series, 70. Pp. 515–529.
- FURNESS, R.W. & BAILLIE, S.R. (1981): Factors affecting capture rate and biometrics of storm petrels on St. Kilda. – *Ringling & Migration*, **3**: 137–148.
- GONZÁLEZ-SOLÍS, J., CROXALL, J.P. & WOOD, A.G. (2000): Sexual dimorphism and sexual segregation in foraging strategies of Northern giant petrel, *Macronectes halli*, during incubation. – *Oikos*, **90**: 390–398.
- GONZÁLEZ-SOLÍS, J. (2004): Sexual size dimorphism in Northern giant petrels: ecological correlates and scaling. – *Oikos*, **105**: 247–254.
- GUTIÉRREZ-CORCHERO, F., CAMPOS, M., HERNÁNDEZ, A., AMEZCUA, A. (2007): Biometrics of the Southern Grey Shrike (*Lanius meridionalis*) in relation to sex and age. – *Ringling & Migration*, **23**: 141–146.
- HEMERY, G. & D'ELBEE, E. (1985): Discrimination morphologique des populations atlantique et méditerranéenne de Pétrels Tempête *Hydrobates pelagicus*. Oiseaux Marins Nicheurs du Midi et de la Corse. – *Annales du C.R.O.P.*, **2**: 63–67.
- HERMOSELL, I.G., BALBONTÍN, J., MARZAL, A., REVIRIEGO, M. & DE LOPE, F. (2007): Sex determination in barn swallows *Hirundo rustica* by means of discriminant analysis in two European populations. – *Ardeola*, **54**(1): 93–100.
- HIPKISS, T. (2007): Can migrating Tengmalm's Owls *Aegolius funereus* be reliably sexed in autumn using simple morphometric measurements? – *Ringling & Migration*, **23**: 201–204.
- HULL, C.L. (1996): Morphometric indices for sexing adult Royal, *Eudyptes schlegeli*, and Rockhopper Penguins, *E. chrysocome*, at Macquarie Island. – *Marine Ornithology*, **24**: 23–27.
- INFANTE, O. & PERIS, S.J. (2004): Sexual dimorphism in the Southern Grey Shrike *Lanius meridionalis* en el centro-oeste de la península Ibérica. – *Ardeola*, **51**(2): 455–460.
- JAMES, P.C. (1983): Storm petrel tape lures: which sex is attracted? – *Ringling & Migration*, **4**: 249–253.
- KAHN, N.W., JOHN, J.S. & QUINN, T.W. (1998): Chromosome-specific intron-size differences in the avian CHD gene provide an efficient method for sex identification in birds. – *The Auk*, **115**: 1074–1078.
- KUCZYNSKI, L., TRYJANOWSKI, P., ANTZAK, M., SKORACKI, M. & HROMADA, M. (2003): Repeatability of measurements and shrinkage after skinning: the case of the Great Grey Shrike *Lanius excubitor*. – *Bonner Zoologische Beiträge*, **51**: 127–130.
- LACK, D. (1971): *Ecological isolation in birds*. – Blackwell Scientific Publications, Oxford and Edinburgh, 404 pp.
- LALANNE, Y., HÉMERY, G., CAGNON, C., D'AMICO, F., D'ELBÉE, J. & MOUCHÉS, C. (2001): Discrimination morphologique des sous-espèces d'océanite tempête: nouveaux résultats pour deux populations méditerranéennes. – *Alauda*, **69**(4): 475–482.
- LEZANA, L., MIRANDA, R., CAMPOS, F. & PERIS, S.J. (2000): Sex differentiation in the spotless starling (*Sturnus unicolor* TEMMINCK, 1820). – *Belgian Journal of Zoology*, **130**(2): 139–142.
- LIORDOS, V. & GOUTTNER, V. (2008): Sex Determination of Great Cormorants (*Phalacrocorax carbo sinensis*) using Morphometric Measurements. – *Waterbirds*, **31**(2): 203–210.
- LO VALVO, M. (2001): Sexing adult Cory's Shearwater by discriminant analysis of body measurements on Linosa Island (Sicilian Channel), Italy. – *Waterbirds*, **24**(2): 169–174.
- MALLORY, M.L. & FORBES, M.R. (2005): Sex discrimination and measurement bias in Northern Fulmars *Fulmarus glacialis* from the Canadian Arctic. – *Ardea*, **93**(1): 25–36.
- MARTÍN, A. & LORENZO, J.A. (2001): *Aves del archipiélago canario*. Francisco Lemus, La Laguna, 787 pp.
- MASSA, B. & CATALISANO, A. (1986) Observations on the Mediterranean Storm Petrel *Hydrobates pelagicus* at Marettimo isle. – *Avocetta*, **10**: 125–127.
- MILLER, S.L., GREGG, M.A., KURITSUBO, A.R., COMBS, S.M., MURDOCK, M.K., NILSSON, J.A., NOON, B.R. & BOTZLER, R.G. (1988): Morphometric variation in tundra swans: relationships among sex and age classes. – *The Condor*, **90**: 802–815.
- NAVARRO, J., KALIONTZOPOULOU, A. & GONZÁLEZ-SOLÍS, J. (2009): Sexual dimorphism in bill morphology and feeding ecology in Cory's shearwater (*Calonectris diomedea*). – *Zoology*, **112**: 128–138.
- NOGALES, M., MARTÍN, A., QUILIS, V., HERNÁNDEZ, E., DELGADO, G. & TRUJILLO, O. (1993): Estatus y distribución del Paiño Común (*Hydrobates pelagicus*) en las Islas Canarias. – In: AGUILAR, J.S., MONBAILLIU, X. & PATERSON, A.M. (eds.): *Estatus y Conservación de Aves Marinas*, SEO/BirdLife-Medmaravis, Madrid. Pp. 15–24.
- PRICE, T.D. (1984): The evolution of sexual size dimorphism in Darwin's finches. – *American Naturalist*, **123**: 500–518.
- RISTOW, D. & WINK, M. (1980): Sexual dimorphism of Cory's Shearwater. – *Il-Merill*, **21**: 9–12.
- RISTOW, D. & WINK, M. (2004): Seasonal variation in sex ratio of nestling Eleonora's Falcons. – *Journal of Raptor Research*, **38**(4): 320–325.
- SAMBROOK, J., FRITSCH, E.F. & MANIATIS, T. (1989): *Molecular cloning: a laboratory manual*. – Cold Spring Harbor Laboratory Press, New York, 1626 pp.
- SERRANO-MENESES, M.A. & SZÉKELY, T. (2006): Sexual size dimorphism in seabirds: sexual selection, fecundity selection and differential niche-utilisation. – *Oikos*, **113**: 385–394.
- SOKAL, R.R. & ROLF, F.J. (1995): *Biometry*. – W.H. Freeman and Company, New York, 887 pp.
- SUTHERLAND, W.J., NEWTON, I. & GREEN, R.E. (eds.) (2004): *Bird Ecology and Conservation: A Handbook of Techniques*. – Oxford University Press, Oxford, 386 pp.
- SVENSSON, L. (1992): *Identification guide to European Passerines*. Fourth edition. – Svensson, Stockholm, Sweden.
- TABERLET, P. & BOUVET, J. (1991): A single plucked feather as a source of DNA for bird genetic studies. – *The Auk*, **108**(4): 959–960.

THIOLLAY, J.M. (1994): Family Accipitridae (Hawks and Eagles). – In: DEL HOYO, J., ELLIOTT, A. & SARGATAL, J. (eds.) Handbook of the Birds of the World. Vol. 2. – New World Vultures to Guineafowl. – Lynx Editions, Barcelona. Pp. 52–205.

WARHAM, J. (1990): The Petrels. Their Ecology and Breeding Systems. – Academic Press, London, 440 pp.

APPENDIX

Examples of sex determination in birds and degrees of accuracy by Determinant Function Analysis (DFA) based on various measurements.

Bird species	% classification accuracy	Reference	Measures
<i>Aegolius funereus</i>	70	Hipkiss 2007	Wing length
<i>Buteo jamaicensis calurus (adults)</i>	98	Donohue & Dufty 2006	Wing chord and body mass
<i>Buteo jamaicensis calurus (yearlings)</i>	97	Donohue & Dufty 2006	Body mass, wing chord, hallux and culmen
<i>Cyanopica cyanus cooki (adults)</i>	90	Alarcos et al. 2007	Tail length, head plus bill length and wing length
<i>Cyanopica cyanus cooki (yearlings)</i>	90	Alarcos et al. 2007	Tail length and head plus bill length
<i>Cygnus columbianus (adult)</i>	74.1–91.3	Miller et al. 1988	Several internal and external traits
<i>Cygnus columbianus (immature)</i>	27.3–52.4	Miller et al. 1988	Several internal and external traits
<i>Eudiptes chrysocome</i>	93.2	Hull 1996	Bill depth and bill length
<i>Eudiptes schlegeli</i>	97.1	Hull 1996	Bill depth and bill length
<i>Fulmarus glacialis</i>	88–90	Mallory & Forbes 2005	Head, bill and tarsus
<i>Hirundo rustica</i>	90.1–91.9	Hermosell et al. 2007	Length of outermost tail feathers, inner tail feathers and keel
<i>Phalacrocorax carbo</i>	92.6–95.1	Liordos & Gouttner 2008	Wing length, culmen length and tarsus length
<i>Puffinus yelkouan</i>	87.2	Bourgeois et al. 2007	Bill depth at gonys, nalospi and tarsus height
<i>Puffinus yelkouan</i>	100	Bourgeois et al. 2007	Four acoustic parameters