Re-description of *Xyliphius barbatus* (Siluriformes, Aspredinidae), with comments on osteology and distribution

Guillermo E. Terán1*, Alejandro Méndez-López†, Mauricio F. Benitez2, Wilson S. Serra3,4, Sergio Bogan5, Gastón Aguilera1

1 Fundación Miguel Lillo – Unidad Ejecutora Lillo FML-UEL-CONICET, (4000), Miguel Lillo 251, San Miguel de Tucumán PC 4000, Tucumán, Argentina
2 Instituto de Biología Subtropical (UNaM-CONICET), Félix de Azara 1552, Posadas, Misiones, Argentina
3 Sección Ictiología, Departamento de Zoología, Museo Nacional de Historia Natural, Montevideo, Uruguay
4 Centro Universitario Regional del Este (CURE) Sede Rocha, Rocha, Uruguay
5 División Ictiología, Museo Argentino de Ciencias Naturales “Bernardino Rivadavia”, Av. Ángel Gallardo 470, C1405DJR, Buenos Aires, Argentina

https://zoobank.org/64E8D409-0377-4629-90DC-946A2CB4BE61

Corresponding author: Gastón Aguilera (gaguilera@lillo.org.ar)

* These authors contributed equality.

Abstract

The banjo catfish, *Xyliphius barbatus*, belongs to the Aspredinidae family and typically inhabits the main channels of medium to large rivers in the La Plata River basin. The mimetic coloration with the substrate and the benthic lifestyle likely contribute to the challenge of sampling this species, resulting in its underrepresentation in museums and ichthyological collections. In fact, the original description of *X. barbatus* was based solely on two specimens. Consequently, little is known about its osteology, distribution, and phylogenetic relations. In this work, these information gaps are filled and the distributional range for *X. barbatus* is extended to northwestern Argentina.

Key Words

Banjo catfish, fossorial fishes, La Plata River basin, morphology, osteology

Introduction

The family Aspredinidae is composed of 13 genera and 49 valid species (Fricke et al. 2023a), among which the genus *Xyliphius* Eigenmann, 1912, include seven species (Fricke et al. 2023b). According to Friel (1994) the genus is distinguishable from all other aspredinids by the following combination of characters: eyes highly reduced, premaxillary toothless and displaced lateral to mesethmoid, a row of fleshy papillae on lower lip, *unculi* and *unculiferous* tubercles flattened, lamina of pterotic rounded, and lateral end of posterohyal expanded. Members of this genus are distributed in different basins in South America: *X. soiae* Sabaj Pérez, Carvalho & Reis, 2017 in the Amazon River basin in Peru, *X. melanopterus* Orcés, 1962 and *X. leptomurus* Orcés, 1962 in the upper Amazon and Orinoco River basins, *X. kryptos* Taphorn & Lilyestrom, 1983 in the Maracaibo basin, *X. anachoretes* Figueiredo & Britto, 2010 in the Tocantins River basin, *X. magdalenae* Eigenmann, 1912 in the Magdalena River basin, and *X. barbatus* Alonso de Arámburu & Arámburu, 1962 from La Plata River basin.
Except for Xyliophius lepturus, all the species of the genus were described based on one to three specimens (X. lombarderoi, X. melanopterus, X. magdalenae and X. soiæ based on a single specimen; X. barbatus and X. anachoretes on two specimens; and X. kryptos on three specimens). These fishes are rare in inventories of fish fauna and consequently in museum collections. This sparse representation is due to several factors, including their mimetic coloration with the substrate, their benthic lifestyle, the infanaul habits of many species, and, most importantly, their specific habitat preferences. Indeed, these species live in the main channels of medium to large rivers, where sampling is difficult (Carvalho et al. 2017). Xyliophius barbatus, originally described from the Paraná River in Rosario (Alonso de Aramburu and Aramburu 1962), has a few records along the La Plata River basin: Paraná River in Misiones, Santa Fe (García 1992; Calviño and Castello 2008) and Chaco provinces (type locality of Xyliophius lombarderoi Risso & Risso, 1964, a synonym of Xyliophius barbatus) (Risso and de Rosso, 1964); in the Uruguay River in Uruguay (Loureiro et al. 2013) and, more recently, in the Paraguay River basin, Pantanal, Brazil (Giménces Junior and Rech 2022).

In Argentina, other aspedinids have been recorded: Amuralia oviraptor Friel & Carvalho, 2016, Bunocephalus doriae Boulenger, 1902, Pseudobunocephalus iheringii (Boulenger, 1891), Pseudobunocephalus rugosus (Eigennan & Kennedy, 1903), and Pterobunocephalus depressus (Haseman, 1911). Although the upper section of the Bermejo River basin has many endemisms (e.g., Miranda et al. 2004a, b, 2006; Casciotta and Almirón 2004; Terán et al. 2016a, 2019; Alonso et al. 2018; Aguijera et al. 2019), it shares elements with the Paraguay-Paraná River basin (Alonso et al. 2016; Terán et al. 2016b; Vanegas-Ríos et al. 2019), and the Amazon River basins (Littmann et al. 2015; Aguijera et al. 2022). The only record of aspedinids in this area is Bunocephalus doriae Aguijera et al. (2016).

New inventories made in the Bermejo River basin have revealed the presence of Xyliophius barbatus, a species not previously recorded for the area. This record is the largest known batch regarding the species. The aim of this contribution, is to provide an accurate re-description of X. barbatus.

Materials and methods

Specimens were collected by electrofishing and hand nets, euthanized by immersion in tricaine methanesulfonate (MS222), fixed in 10% formalin solution, and transferred to individual batches in a 70% alcohol solution. The material was deposited in ichthyological collections. In some fresh individuals, a small tissue sample was taken and immediately preserved in absolute ethanol for genetic analysis. The tissue aliquots were deposited in the tissue collection of CIT-FML. The study complied with the animal welfare laws, guidelines, and policies of the Comité Nacional de Ética en la Ciencia y Tecnología, Argentina. Collection permits were granted by the Ministerio de Ambiente de Jujuy. (permits’ numbers: 1103-306-M/2016, Res. N° 137/2016-MA).

Point to point measurements were taken with a digital caliper to the nearest 0.1 mm and expressed as percentage of the standard length (SL), except for subunits of head, expressed as percent of the head length (HL) (see Table 1). Measurements follow Friel (1995) and Cardoso (2010), with the modifications proposed by Carvalho et al. (2017). Nomenclature of anatomical structures follows that used by the three mentioned authors and Dahdou et al. (2016), included in the Teleost Anatomy Ontology of Uberon Ontology Documentation (Mungall et al. 2012, http://oboontology.github.io/uberon/). Specimens were cleared and stained (C&S) following Taylor and Van Dyke (1985). An asterisk (*) indicates holotype counts. Vertebral counts included the Weberian Complex (5 vertebrae) plus all free vertebrae and the compound caudal centrum (PU1 +U1) counted as one element.

Total genomic DNA was extracted from ethanol-preserved muscle tissue of specimen CI-FML 7944 (tissue collection number: CIT-FML 00169), using a salt-based protocol (Aljanabi and Martinez 1997). A 651 bp fragment of mitochondrial gene Cytochrome oxidase subunit I (COI) was amplified by Polymerase Chain Reaction (PCR) using the cocktail primers proposed by Ivanova et al. (2007). The PCR protocol was the implemented by Ward et al. (2005). PCR-amplified product was purified with 20% PEG. The product was sequenced with automated sequencer (Macrogen, Korea) in both directions to check for potential errors. The Chromatogram was processed and edited using Geneious (Technelysis Pty Ltd) and deposited in GenBank under the accession number OQ539436. COI sequences of different Xyliophius species from GenBank (http://www.ncbi.nlm.nih.gov/Genbank) were used to assess the relationships between X. barbatus and the remaining species of the genus, providing a preliminary phylogenetic hypothesizes. GeneBank codes and references for the employed sequences are shown in Table 2. Pairwise genetic distances with K2P model and UPGMA analysis were performed for 651 -bp fragments using MEGA11 (Tamura et al. 2021). The Maximum likelihood tree was estimated employing RAxML (Randomized Accelerated Maximum Likelihood) under GTR+FO+G4 model and 100 bootstrap replicates.

Table 1. Morphometric data of *Xyliphius barbatus*. SD= standard deviation. Holotype (MLP 6798), Paratype (MLP 2799), Paraná River (MACN-Ict 6791. 5 ex.) Bermejo River specimens (CI-FML7944 and CFA-IC-12742 34 ex.).

<table>
<thead>
<tr>
<th>Species</th>
<th>GenBank ID</th>
<th>Catalog</th>
<th>Basin</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Xyliphius barbatus</em></td>
<td>OQ539436</td>
<td>CI-FML 7944</td>
<td>Bermejo River, Argentina</td>
</tr>
<tr>
<td><em>Xyliphius barbatus</em></td>
<td>KU288948</td>
<td>MG ZV P 355</td>
<td>Paraná River, Argentina</td>
</tr>
<tr>
<td><em>Xyliphius lepturus</em></td>
<td>MF489386</td>
<td>AUM46757</td>
<td>Amazon River, Peru</td>
</tr>
<tr>
<td><em>Xyliphius magdalenae</em></td>
<td>MF489382</td>
<td>ANSP192845</td>
<td>Magdalena River, Colombia</td>
</tr>
<tr>
<td><em>Xyliphius melanopterus</em></td>
<td>MF489383</td>
<td>MUSM36715, AP12</td>
<td>Amazon River, Peru</td>
</tr>
<tr>
<td><em>Xyliphius melanopterus</em></td>
<td>MF489384</td>
<td>MUSM36715, AP28</td>
<td>Amazon River, Peru</td>
</tr>
<tr>
<td><em>Xyliphius sofiae</em></td>
<td>MF489385</td>
<td>STRIO1784</td>
<td>Amazon River, Peru</td>
</tr>
<tr>
<td><em>Xyliphius sofiae</em></td>
<td>KU736764</td>
<td>ANSP 182322</td>
<td>Amazon River, Peru</td>
</tr>
</tbody>
</table>

Table 2. List of sequences, with accessions and vouchers employed for molecular analysis.

Results

*Xyliphius barbatus* Alonso de Arámburu & Arámburu, 1962

Figs 1–6

Material examined. All from Argentina: MLP 6798. Holotype. 90.3 mm SL. Paraná River at Rosario, Santa Fe province. Col. C. Vidal. MLP 2799. Paratype. 87.2 mm SL. Paraná River at Rosario, Santa Fe province. Col. R. Ringuet. MACN 6791. 5 ex. 44.6–93.6 mm SL. Paraná River near Curtiembre, border between Santa Fe and Entre Ríos provinces. 31°27’18.34”S, 60°10’11.95”W. 35–45 m depth 1961–1962. Col. N. Bellisio. MG-ZV-P 355 (LAR-254). 1 ex, 99.1 mm SL. Paraná River in front of Rosario, Entre Ríos province, Argentina. 32°55’58.8”S, 60°37’58.8”W. 6 m depth. 04/03/2013 Col. Julián Aguilar. C1-FML 7944. 29 ex (3 C&S), 79.6–111.3 mm SL. CFA-IC-12742. 5 ex. 79.7–99.2 mm SL. San Francisco River, Bermejo River basin, Jujuy province; 23°50’27.08”S, 64°37’24.70”W, ca. 370 m asl. 1–2 m depth. 30 Sep 2016. G.E. Terán, G. Aguilera and D. Delgado.

Diagnosis. *Xyliphius barbatus* is distinguishable from the remaining species of genus by the following combination of characters: (1) seven to 11 retrose denticles on posterior margin of pectoral-fin spine (vs. six in *X. anachoretes* and four or five in *X. magdalenae*); (2) 24 to 30 dendriform papillae on inferior lip (vs. 20–22 in *X. magdalenae*, 30 in *X. sofiae*, and 22 to 27 triangular papillae, with only the lateral ones branched in
X. kryptos); (3) I,3 or I,4 dorsal-fin rays (vs. I,5 in X. lepturus and X. melanopterus); (4) absence of dorsal pale band from snout tip to caudal-fin origin (vs. presence in X. anachoretes, X. magdalenae and X. melanopterus); (5) absence of a latero-dorsal band following the second row of tubercles on anterior part of body (vs. present in X. magdalenae and X. melanopterus); (6) eyes present and reduced (vs. absent in X. sofiae); (7) five to eight anal-fin rays (vs. nine in X. lepturus).

The additional characters that distinguish Xyliphius barbatus from the remaining species of the genus are: papillae dendriform on lower lip with large branches (vs. papillae with minute branches on X. anachoretes); three dorsal procurent rays (vs. two in X. anachoretes, four to five in X. lepturus, and four in X. magdalenae); pelvic fin not reaching anal-fin origin (vs. just reaching in X. magdalenae); maxillary barbel surpassing pectoral-fin spine insertion (vs. not quite reaching pectoral in X. magdalenae); four branchiostegal rays (vs. five in X. sofiae); and two ossified proximal radials on pectoral fin (vs. one in X. sofiae).

**Morphological description.** Morphometric data is summarized in Table 1. Head and anterior part of body depressed, compressed from dorsal-fin origin to caudal-fin insertion. Maximum depth at dorsal-fin origin. Dorsal profile straight from snout tip to dorsal-fin origin, relatively depressed along dorsal-fin base, and relatively straight from this point to caudal-fin origin. Ventral profile of body, straight from lower-jaw to pectoral-fin origin, convex to pelvic-fin origin with the lowest point at the origin of the pelvic bone, then slanted dorsally to anal-fin origin and relatively straight form this point to caudal-fin base. Greatest width just anterior to pectoral-fin origin.

Head triangular with a rounded snout. Eyes reduced (Fig. 3), located closer to snout tip than to supraoccipital protuberance, and covered with skin less pigmented than surroundings areas. Two nares, anterior nostril tubular, posterior one smaller and located closer to eyes than to anterior nostril. Maxillary barbel on side of snout, inserted just above rictus. Maxillary barbel reaching pectoral-fin base. Two pairs of mental barbels smaller than maxillary ones and located close to the mouth gape.
The external one just before the vertical line through eye, reaching mouth gape when adpressed; inner mental barbels smaller, reaching outer barbels origin, surpassing the lower lip and reaching end of papillae when adpressed. Mouth subterminal, much wider than snout, with 20 to 30 dendritic papillae. Anterior margin of snout with a groove at middle line in dorsal view. Gill slits small, located ventrally on head, before pectoral-fin origin; gill membranes united to isthmus. Small genital papillae just posterior to anus.

Head, trunk, and fins are all covered by thick skin, while the skin in the ventral area and at the fin insertions is thinner. Trunk covered by unculiferus tubercles, which are more concentrated in the head. Five lateral rows of large tubercles extending from post-cephalic region to caudal-fin base, and concentrated in mid-dorsal line.

Dorsal-fin I,2,i (1) or I,3,i (33*), spine feeble. Dorsal-fin insertion on anterior half of body, closer to snout tip than to caudal-fin insertion, anterior to pelvic-fin insertion; shape triangular, rays elongated beyond the membrane. Anal fin i,3,i(1), ii,3,i(6), iii-3,i(7), ii-4,i(16*), iii-4,i(1), ii-5,i(3), ovoid, first branched anal-fin ray longest. Anal-fin insertion on the posterior half of body, closer to caudal-fin insertion than to snout tip. Pectoral-fin I,4,i (34*), its origin almost at half way between snout tip and dorsal-fin origin, and its distal tip reaching pelvic-fin origin. Distal tips of branched rays elongated beyond membrane. Pectoral spines thick, with seven to 11 developed retorse serrae along posterior margin; spine capped by a fleshy elongation. Pelvic fin i,3,i (1) or i,4,i (33*), origin just posterior to vertical through dorsal-fin origin. The tip of pelvic-fin rays elongated throughout the membrane. Caudal fin 1,4/4,i (34*), The ventral-most three branched caudal-fin rays longer.

Osteology (Figs 4–6).

Mesethmoid deep, slightly longer (anteroposteriorly) than wide, with an anterior notch separating two anterior wings. Premaxillae articulating on a ventrolateral concavity of mesethmoid, not visible in dorsal view due to a dorsal lamina. Two diverging laminae articulating dorsally with frontals by interdigitations. Lateral ethmoids articulate with frontals through a dorsal interdigitated...

Figure 2. Lateral, dorsal and ventral views of Xyliphius barbatus. CI-FML 7944. San Francisco River, Bermejo River basin, Jujuy province. Scale bar: 10 mm.
process and medially with orbitosphenoid by cartilage; the latter covered ventrally by a laminar extension of the same bone articulating with parasphenoid; projecting lateral process joined to autopalatine at its middle length; ventrally flattened and extensively articulating with parasphenoid posteromedially. Frontals about 3.5 times longer than wide, their posterior wing articulating with supraoccipital, and lateral posterior margins enclosed by anterior extension of sphenotic. Anterior fontanel about 1.4 times larger than posterior one, synchondral articulation between lateral ethmoids and mesethmoid completely visible through fontanel. Supraoccipital enclosing almost half of posterior fontanel. Epiphyseal bar with a strong suture, its length equal to or greater than that of posterior fontanel. Supraoccipital a little longer than wide, extensively articulating with pterotic laterally, and with epioccipital posterolaterally; a notch in posterior region receiving the ascending process of posttemporal-supracleithrum; posterior process wide and short, in contact with dorsal portion of complex vertebrae. Sphenotic pitted; ventral surface with greater pores, sutured to pterotic laterally, leaving a posterior space with same bone in ventral view, with extensive anterior synchondral articulation with hyomandibula, sutured medially to parasphenoid and posteriorly to anterior lamina of prootic. Dorsal prootic covering lateral surface of frontals and with posterior articulation with supraoccipital. Pterotic with anterior lamina sutured to sphenotic, a concavity after its contact with suprapreopercle, and a lateral rounded expansion reaching opercle ventrally; contacting posttemporal-supracleithrum lateral arm at its posterior region. Epioccipital articulating anterolaterally with pterotic, medially with supraoccipital and dorsally with posttemporal-supracleithrum. The latter bone with dorsal process reaching lateral surface of supraoccipital over epioccipital; a ventral pointed process contacting posterior expansion of pterotic and posterior arm of the bone sutured to complex vertebrae. Wider portion of supraoccipital extending posteriorly with sphenotic, to middle basioccipital at its posterior end. Pterotic visible only in ventral view, anterior lamina over sphenotic cartilage, extensively contacting parasphenoid medially, and sutured to pterotic posterolaterally, leaving to an anterior space with the same bone. Exoccipital bearing two projections enclosing posterior parasphenoid, strongly sutured to postero medial edge of prootic, to epioccipital laterally and with complex vertebra by interdigitations. Basioccipital sub triangular, with a large foramen on its posterior half, a conspicuous pore anteromedially and additional ones laterally.

**Figure 3.** Head detail of *Xyliphius barbatus*. CI-FML 7944. San Francisco River, Bermejo River basin, Jujuy province. Scale bar: 5 mm.

**Figure 4.** Skull of *Xyliphius barbatus* CI-FML 7944. **A.** Ventral view; **B.** Dorsal view. ach: anterior ceratohyal; ang: anguloarticular; anp: anterior nuchal plate; br: branchiostegal rays; cl: cleithrum; co: scapulocoracoid; den: dentary; dfr: dorsal fin rays; epo: epioccipital; ex: extrascapular; fro: frontal; gch: gas bladder chamber; hyo: hyomandibula; io1: infraorbital 1; iop: interopercle; iot: infraorbital tubules; let: lateral ethmoid; mc: mandibular canal tubules; mes: mesethmoid; max: maxilla; na: nasal; op: opercle; pal: autopalatine; pch: posterior ceratohyal; pfr: pectoral-fin rays; pmx: premaxilla; pnp: posterior nuchal plate; po: preopercle; pso: pectoral-fin spine; pto: pterotic; pv5: parapophysis of vertebra five; qua: quadrate; rad: pectoral-fin radial; rb1: first rib; soc: supraoccipital; sc: posttemporal-supracleithrum; spo: sphenotic; sup: suprapreopercle; uh: urohyal; vh: ventral hypohyal. Scale bar: 1 mm.
Premaxillaries dorsomedially flattened and oval, in contact with ventrolateral notches of mesethmoid and separated by the latter. Premaxillary teeth absent. Maxillary tubular, with furrowlike opening on ventral surface; its condyle attached to anterior palatine cartilage. Dentary long and slender, laminar posterior region overlapping anterior face of angular; teeth conical and pointed inwards, arranged in two rows, the outer one with 3 to 5 teeth near the symphysis, and the inner row with 11–12 teeth; coronomeckelian absent. Meckel’s cartilage somewhat conical, wider laterally from its origin at angular, and slender medially at its joining with the dentary, which is located ventral to (or a little displaced medially) dentary dorsolateral notch.

Hyomandibula in dorsoventrally oblique position with respect to neurocranium; dorsal-most edge under anterior extension of sphenotic, followed by a synchondral articulation with the same bone; anterior cartilage contacting quadrate and extended to lateral portion of metapterygoid. Preopercle on lateral portion of hyomandibula, sutured to quadrate on its synchondral articulation with the same bone. Quadrate condyle anteroventrally oriented, to anguloarticular. Metapterygoid square, a little larger than endopterygoid, with a posterior concavity for the anterior lamina of hyomandibula. Endopterygoid ventral to posterior third of autopalatines, at posteroventral concavity of lateral ethmoids after its projecting lateral process, bearing a lateral pointed projection reaching autopalatine middle cartilage.

Autopalatines with expanded anterior and posterior edges, its narrower portion anterior to lateral ethmoids cartilage. Opercle medially articulated with lateral arm of hyomandibula, anteriorly expanded and sutured with interopercle; posterior pointed projection reaching ventral expansion of pterotic. Interopercle accompanying ventralmost edge of opercle, its anterior pointed projection over posterior ceratohyal, covering the interhyal articulation from lateral view.

**Figure 5.** Dorsal view of the neurocranium of _Xyliphius barbatus_ CI-FML 7944. mes: mesethmoid; io1: infraorbital 1; na: nasal; max: maxilla; fro: frontal; spo: sphenotic; soc: supraoccipital; pto: pterotic; epo: epiooccipital; sc: supracleithrum; cv: weber camara; gch: gas bladder chamber; pv5: parapophysis of vertebra five. Pores of the cephalic sensory lateral system s1 to s6 represent the pores from the supraorbital canal and i1 to i6 from the infraorbital canal. Scale bar: 2 mm.

**Figure 6.** Anal, caudal and pelvic fins of _Xyliphius barbatus_ CI-FML 7944. A. Anal-fin and pterygiophores; B. Caudal complex and fin; C. basipterygium. afr: anal-fin rays; apt: anal-fin pterygiophores; cfr: caudal-fin rays; dlw: dorsolateral wing of basipterygium; dpr: dorsal procurrent rays; nsp: neural spines of vertebrae 15 to 18; pcb: posterior cartilage of basipterygium; pfr: pelvic-fin rays; vpr: ventral procurrent rays. Scale bar: 1 mm.
Urohyal subtriangular, with a medial longitudinal cleft on anterior corner and posterolateral developed wings. Two basibranchials, the anterior one about twice larger, contacting first hypobranchial anteriorly and second hypobranchial cartilage posterolaterally; posterior one reached by third hypobranchial. Only the first hypobranchial ossified with wider distal portion. Hypohyal squarish, narrow in its proximal region, and articulated to ceratohyal synchondrally. Ceratohyls with posteroventrally expanded lamina at the articulation with banchiostegal rays; dorsal and ventral extensions over cartilage with posterior ceratohyal sutured to anterior extensions of same bone. Posterior ceratohyal rectangular, with a notch anterior to posterodorsal corner at the articulation with interhyal. Branchiostegal rays four, lateral ones thicker and with more developed lamina, first one (medial) between posterior urohyal and lateral to corner of ceratohyal expanded lamina, the remaining ones associated with anterior ceratohyal cartilage, lateral-most at the articulation cartilage of ceratohyls. Interhyal present, articulated to posterodorsal ceratohyal, with lateral portion of hyomandibular and quadrates.

Ceratobranchials five, first two and last one (fifth) with a single series of small gill rakers, third and fourth with two series; fifth ceratobranchial bearing a dorsal drop-shaped plate with conical teeth, posterior portion long and slender, with four or five gill rakers. Five gill rakers on the anterior border of first and second ceratobranchials, one in the cartilage with first epibranchial; only epibranchials one to three with gill rakers, first with single row and the remaining two with double row of one or two gill rakers restricted to proximal portion. Epibranchials four, an uncinated process on the third one. Third pharyngobranchial thicker at its articulation with cartilage of third epibranchial; fourth pharyngobranchial about half of the latter and located dorsal to an oval tooth plate.

Nasal separated in two tubular ossifications by supraorbital sensory pore s2, posterior tubular ossifications of the supraorbital canal enters frontal just lateral to its articulation with mesethmoid posterior projection. Antorbital present, small, its canal piercing base of dorsal projection of infraorbital 1 and exiting posteriorly. First infraorbital over anterior cartilage of autopalatine, with notch bordering maxillary condyle; anteromedial projection pointed and curved, limiting the anterolateral portion of nares. Posterior infraorbital as a small series of ossicles entering sphenotic canal laterally; ventral branches of i5 and i6 ossified.

Supraorbital sensory canal with pores s1-s2 and s2-s3 separating nasal in two tubular ossifications. Pore S4 opening at anterior frontal fontanel, s5 is missing and s6 opening at posterior frontal fontanel. Infraorbital sensory canal composed of six pores, the first and second ones opening at inner margin of infraorbital 1, and the third at outer margin. Pores i4 to i6 arranged in an arch reaching up close to anterior half of sphenotic.

Tubular series of preopercle mandibular canal initiating below posterior portion of dentary and separated by those joined to preopercle by a gap just lateral to quadratocondyle. Suprapreopercle as a small tubular canal between hyomandibula and pterotic, with dorsal and ventral laminae present. Extrascapular present. Lateral line complete, beginning at posterolateral exit of posttemporal-supracleithrum canal.

Dorsal lamina of Weber apparatus reaching dorsal surface, with flattened and slender process almost reaching nuchal plate posteriorly. The latter rhomboid in dorsal view, with a ventral lamina reaching neural processes of sixth vertebra. Posterior nuchal plate with two slender and pointed anterolateral projections joined to first rib by a ligament, ventrally contacting anterior plate. Gas bladder chamber evident from dorsal view, posteroventral portion partially enclosed by an anteriorly directed ventral lamina. Parapophyses of fifth vertebra reaching lateral wall of body, continuous with posterolateral edge of cleithrum; extensively joined to posterior region of complex vertebræ and covering its lateral border. Ribs six, first pair on sixth vertebra. Vertebrae 35, first 30 or 31 bearing transversal lateral processes.

Pectoral spine retrorse serrations larger distally; first branched ray reaching end of spine or slightly beyond. Two proximal radials associated with the three proximal most rays; first branched ray associated with scapulocoracoid cartilage. Cleithrum dorsomedial pointed projection entering cavity formed by ventral posttemporal-supracleithrum and lateral lamina of complex vertebrae; dorsolateral arm with a proximal pointed projection lateral to posttemporal-supracleithrum and rounded edge; medial arm anteriorly concave, bearing an extensive contact scapulocoracoid posteriorly, and sutured to contralateral cleithrum in larger C&S specimen (102.58 mm SL). Coracoids strongly interdigitated medially, their posterior processes passing base of last pectoral-fin rays and reaching vertical through dorsolateral arm of cleithrum. Pelvic fins not reaching anal-fin origin. Basipterygia with developed dorsolateral wings and lateral cartilage present; medial cartilage not reaching posterior medial margin of bone, which bear jagged borders; posterior cartilage short. First anal-fin pterygiophore reaching posterior portion of vertebra 15 and posterior anal-fin pterygiophore at posterior portion of vertebra 21. Caudal fin with five principal rays on both lobes, three dorsal and four or five ventral procurent rays.

**Coloration** (Fig. 7).

Ground of body dark brown to black, head light brown, pectoral region lighter than dorsal region. A barely evident light brown middorsal stripe on head from snout tip to the middle of caudal peduncle, interrupted at dorsal-fin base. Lateral rows of tubercles brown, lighter than the background. Maxillary barbels dark brown with lighter tips; the remaining barbels light brown. Pectoral fins black with whitish tips; anal fin black with distal half whitish; all the other fins black with the distal third whitish. After the fixation process, the color tends to become paler brown, and the white portions on the fins are less noticeable (Figs 1–3).
Molecular analysis. Molecular comparison employing the COI sequence (see Table 2) shows no difference between Xyliphius barbatus specimen from lot CI-FML 7944 and Xyliphius sp. reported by Díaz et al. (2016) from Paraná River Basin in Rosario, Argentina, herein identified as Xyliphius barbatus. The estimated evolutionary divergence (number of base substitutions per site from between sequences) is quite small (D ≥ 0.0031) between X. barbatus and X. melanopterus, but greater with specimens of X. magdalenae, X. sofiae and X. lepturus (D = 0.1092; 0.1414 and 0.1558). Comparisons with X. anachoretes and X. kryptos were not possible due the lack of available COI sequences for these species. Moreover, the tree topology for UPGMA and Maximum Likelihood analysis was similar (Fig. 8).

Distribution. Including the new record from the San Francisco River, upper Bermejo River basin, Jujuy province and the previous records from the Paraná River, in Rosario (Santa Fe province), and in Chaco province (locality of X. lombarderoi), together with the records from the Paraguay River basin in Paraguay reported by Carvalho et al. (2017) and from the Pantanal in Brazil (Gimênes Junior and Rech 2022), the species exhibit a broad distribution in the Parana-Paraguay system. The new record of specimens from the San Francisco River, Upper Bermejo River basin, Jujuy province, is approximately 750 km in a straight line from the closest record in Tragadero River, Paraná River basin in Chaco Province (Fig. 9).

Ecological notes. Most of the records of this species are from the main channel of big rivers and were collected by trawl nets from the bottom of Parana River at 35–40 m depth (MACN 6791). The specimens from the San Francisco River (Fig. 10) were collected (1 to 4 m depth) using cast nets, and hand nets. Other informal captures made by anglers, supported by photographic evidence (see Suppl. material 1), include one record from the Paraná River in Misiones province (about 4 m depth). In both cases, captures were made using earthworm as bait (Julio Endler and Roberto Toval), see appendix 1. Although speculative, based on available records, this species seems to exhibit fossorial habits, regardless of the substrate depth.

Conservation status. The global conservation status of Xyliphius barbatus was evaluated in 2021, being considered as Near Threatened under criteria B2a by the IUCN.
Discussion

In this contribution we report the northwesternmost record of *Xyliphius barbatus* in Argentina, more than 750 km from the previously distribution record of the species. According to Carvalho et al. (2017) species of *Xyliphius* are commonly found in the main channel of large rivers and most records are restricted to the upland portions of Andean piedmont rivers. Conversely *X. barbatus* and *X. sofiae* occur in relatively lowland stretches. Records of these species and their presence in museum collections are scarce, mainly due to the depth at which they occur, making them difficult to sample. Additionally, their fosorial habits and mimetic coloration contribute to this scarcity. *Xyliphius barbatus* was also recorded in the Uruguay River basin by Loureiro et al. (2013), but this record is indeed a miss-identification of *Bunocephalus doriae* (Loureiro Pers. Comm., 2022). García (1992) includes *X. barbatus* in a list of species from the Paraná River basin in Misiones, but the accuracy of this record cannot be confirmed because the reference specimen is missing; in addition, this record is part of the environmental impact report and is not a formal publication. Nevertheless, the information provided by García (1992) was subsequently repeated (Liotta 2006; Calviño and Castello 2008; Rosso and Liotta 2021).

The scarcity of material has led to biased descriptions, since variations in morphology and meristic counts have not been considered, making it difficult to establish comparisons among species. This situation was also highlighted by Figuereido and Britto (2010) in the description of *Xyliphius anchoretetes*; the authors considered that any inferences concerning the direction of change in the number of papillae from lower-lip is speculative and premature due to the absence of comparative material.

The specimens of *Xyliphius barbatus* collected in the upper Bermejo River represent the largest known batch for the species and one of the largest for the genus. Other numerous collections of aspredinids were made under special conditions. For example, when a river section was dried 70 exemplars of *Hoplomyzon sexpapilostoma* Taphorn and Marrero 1990 were collected, and when a dam was closed more than 100 exemplars of *Xyliphius cf. lepturus* were collected (Taphorn and Marrero 1990). This suggests the low efficacy of conventional fishing methods due to the distinct habitat use by members of the genus (Carvalho et al. 2017). The new *Xyliphius barbatus* material obtained has allowed us to make an accurate re-description for a species which, despite its wide distribution in the Paraná-Paraguay River basins, is poorly represented in fish collections in museums. The inclusion of new material for the re-description of *Xyliphius barbatus* has led to an objective assessment of inter-specific variation of characters useful for distinguishing the species from the remaining members of the genus. Among these characters, we found that the lower limit of papillae on lower lip ranges from 24 to 30 instead of the previously reported 28 to 30; the anal-fin rays now range from 5 to 8 instead of 6 to 8; the number of retrorse serrae along the posterior border of the pectoral-fin spine now ranges from 6 to 11 instead of 6 to 8; and the extension of lower limit range of vertebral count to 35.

From a genetic perspective, maximum likelihood analysis identified two main groups: one comprising *Xyliphius lepturus* and *X. sofiae*, and the other including *X. magdalenae*, *X. barbatus*, and *X. melanopterus*. This tree topology aligns with the morphological inferences made by Carvalho et al. (2017). They suggested that *X. sofiae* and *X. lepturus* shared enough features to support a sister-group relationship. These characters include snout morphology, lack of median notch, fifth ceratobranchial morphology, branchial apertures separation, anal-fin rays’ modal number; and lateral line extension. Carvalho et al. (2017) also proposed a close relationship between *X. anchoretetes*, *X. barbatus*, and *X. melanopterus* who share a relatively long and narrow fourth parapophysis and fifth parapophysis deflected anteriorly (vs. short and broad fourth parapophysis and fifth directed laterally in the remaining species of *Xyliphius*). Despite the lack of
X. anachoretes sequences, our ML tree recovers a very close relationship between X. barbatus and X. melanopterus. The low genetic distance between Xyliophius barbatus and X. melanopterus is noteworthy, since the distance to other species and between the other species is almost 50 times that value. In fact, one of the sequences of X. melanopterus (MF489383) differs only in 1 nucleotide from the X. barbatus sequence. It is evident that this comparison (based in one mitochondrial marker and a few sequences) is not enough to make inferences about the taxonomy of these two species (which have morphological differences, such as ray counts and coloration, and inhabit different basins), but it highlights the need for a complete morphological and molecular revision of the group in order to complement its definition and species diagnosis.

Additional similarities in the complex vertebra between Xyliophius melanopterus and X. barbatus were previously indicated by Carvalho et al. (2017); a narrow and long fourth procurrent vertebrae (observed in examined specimens), and anteriorly deflected fifth parapophyses, perpendicular in the examined X. barbatus. The same authors discuss the possible independent acquisition of gas bladder encapsulation in Xyliophius due to its absence in X. magaldaenae and X. kryptos (see also Carvalho et al. 2018, Fig. 3), and the presence in the remaining congeners (with the polymorphism in X. lepturus). Our results also suggest the reduction of this feature in X. magaldaenae instead of two independent events of gas bladder encapsulation (Fig. 6A). Nonetheless, these characters must be tested and optimized in a morphological analysis to support any conclusive hypothesis.

Neither Friel (1994) nor Cardoso (2008) included specimens of Xyliophius barbatus in their works. Xyliophius barbatus bears the eight synapomorphies proposed by the former author and the additional six synapomorphies for the subfamily Xyliophiinae proposed by the latter author: (1) lateral surface of frontal lacking orbital concavity; (2) antero-dorsal process of lacrimal-anterioral (first infrarostral) developed; (3) supra-preopercle present Friel (1994: Ch.28); (4) expanded proximal margin of posterior ceratohyal (Friel 1994: Ch.34); (5) more than 30 gill filaments on first epibranchial and ceratobranchial (Britto 2002: Ch. 126); (6) four to eight dorsal plus ventral procurrent rays on caudal-fin. We also observed the bony bulge on ventral surface lateral ethmoid possibly housing the olfactory bulb as suggested by Carvalho et al. (2017), as an additional synapomorphy of the genus.

Conclusions

In this work, an accurate re-description of Xyliophius barbatus based on osteological observations, morphometry, meristic counts and molecular data is provided. The distributional range of this species is widened to the upper Bermejo River basin in northwestern Argentina, more than 750 km in straight line from the previously known record of the species in the country. A provisional phylogenetic molecular hypothesis is provided in which the close relation with X. melanopterus is observed.

Acknowledgements

We extend our gratitude to Tiago Carvalho for reviewing the manuscript and making invaluable comments on Xyliophius species, which significantly enhanced the quality and accuracy of this paper. Pablo Pereyra (FML) made figures 4 and 6. We thank Gustavo Chiararamonte (MACN), James Anyelo Vanegas-Rios (MLP), Diego Nadaín (MLP), Germán Saigo (MG), Eugenia Montani (MG) and Adrián Giacchino (CFA, UMAI) for the support provided and for making available the ichthyological collections of the respective institutions. GET and GA thank Diego Delgado for help in sampling trips. Julio Endler and Roberto Tovai, for the photograph of specimens (Suppl. material 1). We thank Jorgelina Brasca for English review and Florencia Brancoolini, Pablo Calviño, Miguel Angel Cortés Hernández for valuable comments. Felipe Alonso and Marcos Mirande for permanent support. This manuscript benefited from the comments and revisions of Nicolas Hubert, Lucas Medeiros, and two anonymous reviewers.

References

Brito MR (2002) Análise filogenética da ordem Siluriformes com ênfase nas relações da superfamília Loricarioidea (Teleostei: Osteo-
physis). Tese de doutorado, Universidade de São Paulo, São Paulo.
512 pp.
Calviño PA, Castello HP (2008) Sobre um bagre ciego do rio Paraná medio, *Xyliphius barbatus* Arámburu and Arámburu, 1962 (Siluri-
Cardoso AR (2010) *Bunocephalus erondinae*, a new species of banjo catfish from southern brazil (siluriformes: Aspredinidae). Neotrop-
Carvalho TP, Arez MH, Reis RE, Sabaj Pérez MH (2018) Molecular phylogeny of banjo catfishes (Ostariophysi: Siluriformes: Aspredi-
org/10.1016/j.ympev.2018.04.039
Casciotta JR, Almirón AE (2004) *Astyanax chico* sp. n. a new species from the rio San Francisco basin, northwest of Argentina (Teleose-
tics Museums fur Tierkunde in Dresden 54: 11–17.
Dahdul WM, Lundberg JG, Midford PE, Balhoff JP, Lapp H, Vision TJ, Montevede, Urugu-
ay, 91–112
Mirande JM, Aguilera G, Azpelicueta MDLM (2006) *Astyanax endy* (Characiformes: Characidae) from the upper rio Bermejo basin, Salta, Argentina. Revue Suisse de Zoologi-
tional information on *Hypopthalmus edentatus* Spix & Agas-
org/10.1635/053.164.0115
Supplementary material 1
Informal records by anglers, with photograph evidence. Specimens collected with earthworm bait

Authors: Guillermo E. Terán, Alejandro Méndez-López, Mauricio F. Benítez, Wilson S. Serra, Sergio Bogan, Gastón Aguilera
Data type: docx
Explanation note: The information provided here represents records of *Xyliphius barbatus* capture by anglers in two localities along the Paraná river, also including a map of these localities.

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zse.100.121396.suppl1

Supplementary material 2
Comparative material examined

Authors: Guillermo E. Terán, Alejandro Méndez-López, Mauricio F. Benítez, Wilson S. Serra, Sergio Bogan, Gastón Aguilera
Data type: docx
Explanation note: In this file all reference material consulted in this work is listed.

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zse.100.121396.suppl2