



# Development of novel effective agents from 1*H*-indolylammonium trifluoroacetates effective against conditionally pathogenic microorganisms

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## Abstract

**Introduction:** The problem of antibiotic resistance of microorganisms is becoming more urgent in the twenty-first century. More and more pathogenic microbes are becoming resistant to two or more antibiotics. This problem has become worse into the COVID-19 pandemic. The search for new compounds with antimicrobial activity is one of the principles for overcoming the antibiotic resistance of microorganisms.

**Materials and methods:** Methods for the preparation, isolation, and identification of salts of 2,3,5-trimethyl-, 1,2,3,5-tetramethyl-, 2,3-dimethyl-5-methoxy-, 5-methoxy-1,2,3-trimethyl-1*H*-indole-6-amines and trifluoroacetic acid were developed and laboratory microbiological studies of them for antimicrobial activity were carried out. Sensitivity of the test-strains of microorganisms to the new compounds was studied. A method of serial dilutions to determine the minimal inhibitory concentration (MIC) of the compounds under study was used in the study.

**Results and discussion:** The compounds 5–8 showed a pronounced antibacterial activity against the test strains of microorganisms *in vitro* with MIC from 0.98 µg/mL to 125.0 µg/mL. The prospects for targeted synthesis of biologically active compounds which are derivatives of 1*H*-indolylamines with a trifluoromethyl group in the molecule were determined, and after additional studies, the compounds 5–8 may find application as water-soluble synthetic antimicrobial agents.

**Conclusion:** The laboratory microbiological screening of showed that they have an antimicrobial effect that exceeds the activity of the reference drug, dioxidine. The presence of molecular mechanisms predicted *in silico* in the spectrum of biological activity of the studied compounds, such as Pseudolysin inhibitor, Omptin inhibitor, Undecaprenyldiphosphomuramoylpentapeptide beta-N-acetylglucosaminyltransferase inhibitor, UDP-epimerase inhibitor, Bacterial efflux pump inhibitor, suggests the presence of antimicrobial activity against gram-positive and gram-negative microorganisms. Trifluoroacetates 2,3,5-trimethyl-1*H*-indole-6-ammonium (5), 1,2,3,5-tetramethyl-1*H*-indole-6-ammonium (6), 2,3-dimethyl-5-methoxy-1*H*-indole-6-ammonium (7), 1,2,3-trimethyl-5-methoxy-1*H*-indole-6-ammonium (8), after additional studies, may find application as water-soluble synthetic antimicrobial agents.

## Keywords

new chemical compounds, 1*H*-indolylamines, conditionally-pathogenic microorganisms, antimicrobial activity.

## Introduction

Throughout the history of the existence of pathogenic microorganisms, the struggle against many of their representatives, both those already known and those recently identified, have been going on. The discovery of antimicrobial agents has led to the successful treatment and elimination of certain bacterial infections, but revealed the strains that are resistant to antimicrobials due to the numerous mechanisms of their antibiotic resistance (Kumarasamy et al. 2010; Parhizgari et al. 2017; Yokoyama et al. 2018).

The problem of antibiotic resistance is becoming more acute in the 21<sup>st</sup> century; a study of the mechanisms for acquiring resistance to antimicrobial agents underlies the development of new ways to combat this phenomenon (McKeegan et al. 2002; Savjani et al. 2009). Drug resistance is a growing global threat to public health that affects all major pathogens and antimicrobials (Brown and Wright 2016; Yadav et al. 2017; Obayiuwana et al. 2018; World Health Organization 2018). In the course of microbiological monitoring over recent years, the share of multiresistant strains has tended to grow, for example, methicillin-resistant *S.aureus* strains have a significantly higher frequency of resistance to gentamicin, clindamycin, rifampicin, tetracycline, chloramphenicol, ceftrofolin, ciprofloxacin, and erythromycin, when compared to methicillin-sensitive strains. *P.aeruginosa* is insensitive to antipseudomonal cephalosporins – cefepime and ceftazidime, as well as piperacillin-tazobactam, imipenem, meropenem. Representatives of the *Enterobacteriaceae* family are resistant to three and more traditionally used antibiotics, such as cefotaxime, ceftazidime, cefepime, aztreonam, etc. (Pop-Vicas et al. 2008; Lai et al. 2014; Natan and Banin 2017; Tacconelli et al. 2018).

The uncontrolled use of antimicrobial drugs in the treatment of COVID-19-associated pneumonia has led to an unprecedented proliferation of antibiotic-resistant nosocomial strains of microorganisms.

The search for and development of new antimicrobial agents is one of the fundamental principles of overcoming the resistance of microorganisms to antibiotics.

Substituted 1*H*-indolylamines with an amino group in the benzene ring are known as intact compounds for the production of trifluoromethyl-substituted indolylamides. Many of the products obtained show various kinds of

biological activity. So, in amides, based on substituted 1*H*-indol-4,7-ylamines and trifluoroacetate, based on 1*H*-indole-7-amines and ethyl trifluoroacetic acid, a rather high antimicrobial activity was found (Stepanenko et al. 2018; Stepanenko et al. 2019).

In this regard, it was of interest to obtain water-soluble derivatives of 1*H*-indolylamines with a trifluoromethyl group in the molecule. Such compounds could be salts formed by substituted 1*H*-indolylamines and trifluoroacetic acid. Our out-of-experimental computer bioscreening of the structures that are salts of 2,3,5-trimethyl-, 1,2,3,5-tetramethyl-, 2,3-dimethyl-5-methoxy-, 5-methoxy-1,2,3-trimethyl-1*H*-indole-6-amines and trifluoroacetic acid predicts antimicrobial activity for them.

Therefore, we have developed methods for the preparation, isolation, and identification of these salts and conducted laboratory microbiological studies on their antimicrobial effects. Depending on the results of the study, the direction of development – antimicrobial chemotherapy, antiseptic or disinfectology – will be determined.

## Materials and methods

### Chemistry

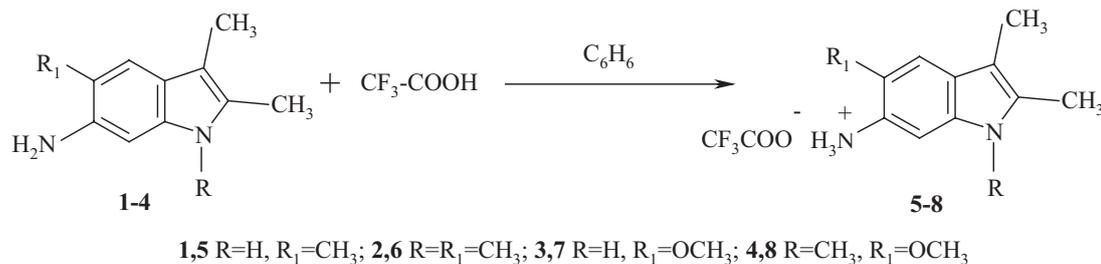
We found that equimolecular amounts of aminoindoles 1–4 and trifluoroacetic acid in a heated benzene solution react with the formation of indolylammonium trifluoroacetates 5–8, which precipitate upon cooling (Scheme 1).

The isolated compounds are light gray, light purple crystalline substances, soluble in water. The physicochemical, spectral characteristics of the obtained new compounds are shown in Tables 1, 2.

### Biological activity

#### *In silico* prediction of the spectrum of biological activity of new compounds

Computer system PASS (Prediction of Activity Spectra for Substances), version 9.1, registered in 2007, was used for predicting the biological activity of substances (Varnek 2008; Filimonov et al. 2014; Filimonov et al. 2018).



**Scheme 1.** Scheme for the synthesis of 1*H*-indol-6-ylammonium trifluoroacetates 5–8 from 1*H*-indol-6-ylamines 1–4 and trifluoroacetic acid.

**Table 1.** Physicochemical characteristics of compounds 5–8

*Compound	** Found, %		Gross formula	Calculated, %		*** $R_f$	$T_{\text{met}}$ (with decomposition), °C	Yield, %
	C	H		C	H			
Trifluoroacetate 2,3,5-trimethyl-1 <i>H</i> -indole-6-ammonium (5)	53.99	5.06	C <sub>13</sub> H <sub>15</sub> N <sub>2</sub> F <sub>3</sub> O <sub>2</sub>	54.16	5.24	0.20	>190	84
Trifluoroacetate 1,2,3,5-tetramethyl-1 <i>H</i> -indole-6-ammonium (6)	55.50	5.59	C <sub>14</sub> H <sub>17</sub> N <sub>2</sub> F <sub>3</sub> O <sub>2</sub>	55.63	5.67	0.38	>173	80
Trifluoroacetate 2,3-dimethyl-5-methoxy-1 <i>H</i> -indole-6-ammonium (7)	50.99	5.06	C <sub>13</sub> H <sub>15</sub> N <sub>2</sub> F <sub>3</sub> O <sub>3</sub>	51.32	4.97	0.24	>161	54
Trifluoroacetate 5-methoxy-1,2,3-trimethyl-1 <i>H</i> -indole-6-ammonium (8)	52.69	5.21	C <sub>13</sub> H <sub>15</sub> N <sub>2</sub> F <sub>3</sub> O <sub>3</sub>	52.83	5.38	0.46	>160	71

**Note:** \* – The amines and salts are named according to the rules of a computer program ACD/LABS IUPAC Name Generator; \*\* – Elemental analysis was performed on an elemental analyzer vario MICRO cube; \*\*\* – The purity of the obtained compounds was monitored,  $R_f$  was determined using TLC on Silufol UV-254 plates in the system: benzene-ethylacetate-methanol 1:1:0.1.

**Table 2.** Spectral characteristics of compounds 5–8

Compound	*Spectrum NMR (DMSO-d <sub>6</sub> ), ppm.	<sup>19</sup> F	** UV Spectrum	*** Mass spectrum: m/z
	<sup>1</sup> H		(ethanol): $\lambda_{\text{max}}$ (lg $\epsilon$ ), nm	(% to $J_{\text{max}}$ )
5	2.13 (3H, c, 3-CH <sub>3</sub> ), 2.29 (3H, c, 2-CH <sub>3</sub> ), 2.36 (3H, c, 5-CH <sub>3</sub> ), 7.26 (1H, c, H-4), 7.30 (1H, c, H-7), 9.69 (3H, c <sub>br</sub> , 6-NH <sub>3</sub> ), 10.81 (H, c, H-1)	-73.58	207 <sub>sh</sub> (4.26), 233(4.52), 295(3.76)	174(100), 173(86), 159(30), 69(53), 45(73)
6	2.16 (3H, c, 3-CH <sub>3</sub> ), 2.31 (3H, c, 2-CH <sub>3</sub> ), 2.38 (3H, c, 5-CH <sub>3</sub> ), 3.59 (3H, c, 1-CH <sub>3</sub> ), 7.31 (1H, c, H-4), 7.33 (1H, c, H-7), 9.69 (3H, c <sub>br</sub> , 6-NH <sub>3</sub> )	-73.66	210 <sub>sh</sub> (4.28), 235(4.53), 300(3.83)	188(100), 187(71), 173(39), 69(43), 45(78), 28(61), 17(12)
7	2.15 (3H, c, 3-CH <sub>3</sub> ), 2.29 (3H, c, 2-CH <sub>3</sub> ), 3.90 (3H, c, 5-OCH <sub>3</sub> ), 7.08 (1H, c, H-4), 7.26 (1H, c, H-7), 9.56 (3H, c <sub>br</sub> , 6-NH <sub>3</sub> ), 10.75 (1H, c, H-1)	-73.56	207 <sub>sh</sub> (4.42), 230(4.55), 303(4.03)	191(18), 190(100), 176(12), 175(96), 147(69), 145(14), 69(21), 45(24), 28(11)
8	2.18 (3H, c, 3-CH <sub>3</sub> ), 2.31 (3H, c, 2-CH <sub>3</sub> ), 3.59 (3H, c, 1-CH <sub>3</sub> ), 3.91 (3H, c, 5-OCH <sub>3</sub> ), 7.13 (1H, c, H-4), 7.30 (1H, c, H-7), 9.44 (3H, c <sub>br</sub> , 6-NH <sub>3</sub> )	-73.63	213 <sub>sh</sub> (4.63), 230(4.60), 300(4.06)	205(15), 204(100), 190(11), 189(74), 161(47), 160(10), 69(12), 45(12), 28(8)

**Note:** \* – NMR spectra were recorded on a Joel JNM-ECX400 multi-core nuclear magnetic resonance spectrometer (400 MHz) in DMSO-d<sub>6</sub>; \*\* – Electronic spectra were obtained on a LEKI SS2109UV device in ethanol; \*\*\* – Mass spectra were recorded on a Finnigan MAT INCOS-50 mass spectrometer with direct input of samples into an ion source at an ionization energy of 70 eV.

### Antimicrobial activity of new compounds (*in vitro*)

During the microbiological experiment, the studied compounds were used in the form of a solution (sterile water for injection was used as a solvent). The test compounds were the following: trifluoroacetate 2,3,5-trimethyl-1*H*-indole-6-ammonium (5), trifluoroacetate 1,2,3,5-tetramethyl-1*H*-indole-6-ammonium (6), trifluoroacetate 2,3-dimethyl-5-methoxy-1*H*-indole-6-ammonium (7), trifluoroacetate 5-methoxy-1,2,3-trimethyl-1*H*-indole-6-ammonium (8).

The following museum strains were used as the test microorganisms for studying the antimicrobial activity of the obtained compounds: *Staphylococcus aureus* 6538-P ATCC, *Staphylococcus aureus* 43300 ATCC (MRSA), *Escherichia coli* 25922 ATCC, *Pseudomonas aeruginosa* 27853 ATCC, and *Streptococcus pyogenes* 19615 ATCC. The museum strains used in this work were obtained from the collection of the Museum of Living Cultures of Federal State Budgetary Institution “Scientific Centre for Expert Evaluation of Medicinal Products (SCEEMP)” of the Ministry of Health of the Russian Federation and Becton Dickinson France S.A.S. The studied strains of microorganisms are the most frequent causative agents of infectious nonspecific human diseases, as well as the most common representatives of the nosocomial microbiota associated with diseases resulting from medical treatment. This is the reason for the choice of the

test and experimental strains. The antimicrobial activity of the obtained compounds was determined by the broth serial dilution method (macrotube method) (MUK 2004; ISO 2006; EUCAST 2019; EUCAST 2021).

The antimicrobial preparation **dioxidine** (a derivative of di-*N*-hydroxyquinoxaline) (Biosintez PJSC, Russia, a solution for topical application, endotracheal and intravenous administrations, 10 mg/mL), widely used in medical practice, was used as a comparison drug. This drug has a high *in vivo* chemotherapeutic activity on model infections similar in pathogenesis to human pathological processes (purulent meningitis, pyelonephritis, septicopyemia) and caused by strains of anaerobic bacteria that are resistant (including multiresistant) to drugs of other classes, including *Pseudomonas aeruginosa* strains and methicillin-resistant staphylococci. **Dioxidine** is characterized by a wide antibacterial spectrum with a bactericidal effect, and is also active against gram-positive and gram-negative aerobic conditionally pathogenic bacteria. The activity of **dioxidine** against *Mycobacterium tuberculosis* is shown (Padeiskaya 2011; Piopov et al. 2013).

To assess the sensitivity of microorganisms, Mueller-Hinton broth (MHB) (HiMedia Laboratories Pvt. Limited, India) was used. The concentration of the suspension of the studied microorganism was  $1.5 \times 10^8$  CFU/mL. The optical density of the bacterial suspension with a concentration

of  $1.5 \times 10^8$  CFU/mL upon visual inspection corresponded to the McFarland turbidity standard of 0.5. A commercial turbidity standard (Sensitre, UK) was used in the work. A bacterial suspension was prepared from agar cultures. A pure culture of microorganisms grown on solid nutrient media was used to prepare the inoculum. Several same-type clearly isolated colonies were selected, which had been grown on non-selective solid nutrient media. Using the loop, a small amount of material was transferred from the tops of the colonies into a test tube with sterile saline, adjusting the inoculum density to exactly 0.5 according to the McFarland standard. Inoculum was used within 15 minutes after preparation.

The broth serial dilution method (macro tube method). Testing was carried out in a volume of 1 mL of each dilution of the test compound with a final concentration of the studied microorganism of approximately  $5 \times 10^5$  CFU/mL. MHB was poured into 0.5 mL in each tube to determine sensitivity. The number of tubes was ten, plus one for a "negative" control, that is, eleven tubes in total. A working solution of the test compound was prepared from the main solution using a liquid nutrient medium – MHB. Then the working solution in an amount of 0.5 mL, using a micropipette with a sterile tip, was introduced into the first tube containing 0.5 mL of broth. The mixture was thoroughly mixed, and, by means of a new sterile tip, 0.5 mL of the broth solution of the test compound was transferred into a second tube containing initially 0.5 mL of broth. This procedure was repeated until all the necessary dilutions were prepared. From the last tube, 0.5 mL of broth was removed. Thus, a number of test tubes were obtained with solutions of the test compound, the concentrations of which are 2 times different in the neighboring tubes. For inoculation, a standard microbial suspension was used, equal to 0.5 according to the McFarland standard, diluted 100 times in MHB, after which the concentration of the microorganism in it was approximately  $10^6$  CFU/mL. Inoculum of 0.5 mL was added to each tube, containing 0.5 mL of the appropriate dilution of the test compound, and to one tube with 0.5 mL of MHB without antibiotic (negative control). The final concentration of the microorganism in each tube was approximately  $5 \times 10^5$  CFU/mL. The inoculum was introduced into the test tubes with dilutions of the test compound no later than 15–30 minutes after they had been prepared. The tubes were stoppered with sterile cotton-gauze plugs, and all the control tubes, except the negative control tube, were incubated in a normal atmosphere at a temperature of 37 °C for 16–20 or 20–24 h (depending on a type of the microorganism being tested). The negative control tube was placed in a refrigerator at 4 °C, where it was stored until the results were assessed. To determine whether there was a microorganism growth, the test tubes with inoculations were examined in transmitted light. The culture growth in the presence of the test compound was compared with the reference tube (negative control) containing the original inoculum and stored in the refrigerator. The minimum

inhibitory concentration (MIC) was determined by the lowest concentration of the test compound, which inhibits the visible growth of the microorganism. The experiment was carried out in four sequences.

## Results and discussion

The structure of the new compounds obtained for microbiological studies is unambiguously confirmed by an analysis of their spectral characteristics. The formation of indolylammonium salts **5–8** was confirmed by the obtained UV,  $^1\text{H}$  NMR,  $^{19}\text{F}$  NMR spectra and mass spectra (Table 2).

So, the UV spectra of the obtained compounds **5–8** are characterized by three absorption bands (207<sub>sh</sub>, 233, 295 nm for salt **5**; 210<sub>sh</sub>, 235, 300 nm for salt **6**, 207<sub>sh</sub>; 230, 303 nm for salt **7**; and 213<sub>sh</sub>, 230, 300 nm for salt **8**) in contrast to the spectra of the starting aminoindoles **1–4**, where there are four absorption bands. In the spectra of the compounds obtained by us, the two long-wavelength bands in the spectra of the starting amines are combined and appear as one broad long-wavelength absorption. Since absorption in the long-wave region is responsible for  $\pi$ - $\pi$  transitions in the benzene part of the molecule, a change in their nature indicates that a change has occurred in the nature of the substitution of this ring, i.e. an amino group has converted to an ammonium group.

The  $^1\text{H}$  NMR spectrum pattern also unambiguously confirms the formation of salts of structure **5–8**. The difference between the spectra of the obtained trifluoroacetates and the spectra of the starting amines is the absence of a proton signal with an integrated intensity of two 6-NH<sub>2</sub> protons in the region of 4–5 ppm and the presence of a downfield much broader peak of exchange hydrogens with an integrated intensity of three protons of the ammonium  $^+\text{NH}_3$  group (9.44–9.69 ppm). In the aliphatic part of the spectra of trifluoroacetates, there are also single singlets of hydrogens of methyl groups, and in the aromatic part of the spectrum, there are signals of two protons of the benzene ring and a 1-H pyrrole fragment (for compounds **5,7**). It should be noted that the values of chemical shifts of the signals of unambiguous protons towards the weak fields in the spectra of trifluoroacetates are compared with amines. Most of all, under this influence are the hydrogens of the benzol fragment, which are the closest to the positively charged ammonium group.

The presence of equivalent fluorine atoms in salt **5–8** molecules is evidenced by a single signal within the range -73.57(**5**), -73.66(**6**), -73.56(**7**), -73.63(**8**) ppm in the HMR  $^{19}\text{F}$  spectra.

Under mass-spectral conditions (high temperature), trifluoroacetates **5–8** decompose to form the corresponding amine and trifluoroacetic acid. Therefore, in the spectra, there are signals of molecular ions of 1*H*-indolylamines and signals of fragment ions obtained upon their splitting under conditions of electron ionization, as well as peaks of fragment particles with *m/z* 69, 45, 28, 17, which are formed during the decomposition of trifluoroacetic acid.

The direction of decay of molecular ions  $F_1$ - $F_4$  depends on the nature of the substituent in the aromatic ring. Thus,  $F_1$ ,  $F_2$  ions ( $R=H$ ,  $CH_3$ ;  $R_1=CH_3$ ) lose a hydrogen atom or methyl radical and rearrange themselves into positively charged particles  $F_5$ ,  $F_6$ ,  $F_7$ ,  $F_8$ , which, according to published data, have quinoline structures (Terent'ev 1979). In the case of molecular ions  $F_3$ ,  $F_4$  of aminoindoles **3**, **4**, the direction of decay is determined by the methoxy group. In this case, a  $CH_3$  radical is cleaved from molecular ions  $F_3$ ,  $F_4$  with the formation of fragment ions  $F_9$ ,  $F_{10}$ , which later, with the loss of the carbon monoxide molecule, produce ions  $F_{11}$ ,  $F_{12}$  having the structure of pyrrolopyridine. This decay is characteristic of *ortho*-anisidines (Khmel'nitskii 1974).

An out-of-experimental prediction of the antimicrobial activity of the synthesized substituted 1*H*-indol-6-ylammonium trifluoroacetates **5–8** was carried out. PASS predicts that a certain compound can manifest the biological activity, but makes impossible any conclusions regarding the magnitude of the activity and the conditions of the experimental testing (dose, route of administration, biological object, gender, age, etc.), under which this activity can occur. Thus, PASS makes it possible to narrow the scope of the experimental testing in relation to specific compounds; however, any prediction must be confirmed by an experiment. According to the PASS prediction, the new derivatives have the following molecular mechanisms: Pseudolysin inhibitor is predicted with a probability of  $Pa$  0.467 for compound **5**,  $Pa=0.433$  – for compound **6**,  $Pa=0.348$  – for compound **7**,  $Pa=0.428$  – for compound **8**; Omptin inhibitor with a probability of  $Pa=0.314$  for compound **5**,  $Pa=0.394$  – for compound **6**,  $Pa=0.364$  – for compound **7**,  $Pa=0.381$  – for compound **8**; UDP-*N*-acetylglucosamine 2-epimerase inhibitor with a probability of  $Pa=0.407$  for compound **5**; Bacterial efflux pump inhibitor with a probability of  $Pa = 0.320$  for compound **6**,  $Pa=0.410$  – for compound **7** (Tables 3–6). By interacting with the molecular targets, an antimicrobial effect can be achieved.

**Table 3.** The Predicted Spectrum of Biological Activity of Compound **5** (PASS) ( $Pa \geq 0.3$ )

<i>Pa</i>	<i>Pi</i>	Activity
0.533	0.008	Multiple sclerosis treatment
0.479	0.032	Autoimmune disorders treatment
0.456	0.038	HMGCS2 expression enhancer
0.438	0.052	Platelet derived growth factor receptor kinase inhibitor
0.467	0.168	<b>Pseudolysin inhibitor</b>
0.313	0.015	Antineoplastic (sarcoma)
0.388	0.110	Chloride peroxidase inhibitor
0.407	0.151	<b>UDP-<i>N</i>-acetylglucosamine 2-epimerase</b>
0.356	0.100	Antiarthritic
0.318	0.085	Plastoquinol-plastocyanin reductase inhibitor
0.321	0.155	Phosphatidylcholine-retinol O-acyltransferase inhibitor
0.320	0.157	Leukopoiesis stimulant
0.363	0.217	Aspulvinone dimethylallyltransferase inhibitor
0.311	0.167	Erythropoiesis stimulant
0.314	0.192	<b>Omptin inhibitor</b>

**Table 4.** The Predicted Spectrum of Biological Activity of Compound **6** (PASS) ( $Pa \geq 0.3$ )

<i>Pa</i>	<i>Pi</i>	Activity
0.427	0.021	Multiple sclerosis treatment
0.445	0.042	Autoimmune disorders treatment
0.433	0.043	<b>Pseudolysin inhibitor</b>
0.368	0.112	Antiinflammatory
0.394	0.206	<b>Omptin inhibitor</b>
0.331	0.146	CYP2D15 substrate
0.317	0.136	4-Nitrophenol 2-monoxygenase inhibitor
0.303	0.138	Dementia treatment
0.310	0.152	Platelet derived growth factor receptor kinase inhibitor
0.350	0.202	Glutamyl endopeptidase II inhibitor
0.320	0.187	<b>Bacterial efflux pump inhibitor</b>
0.302	0.177	Phosphatidylcholine-retinol O-acyltransferase inhibitor

**Table 5.** The Predicted Spectrum of Biological Activity of Compound **7** (PASS) ( $Pa \geq 0.3$ )

<i>Pa</i>	<i>Pi</i>	Activity
0.703	0.054	Gluconate 2-dehydrogenase (acceptor) inhibitor
0.480	0.013	Multiple sclerosis treatment
0.433	0.043	HMGCS2 expression enhancer
0.513	0.131	Aspulvinone dimethylallyltransferase inhibitor
0.461	0.093	Chlordecone reductase inhibitor
0.392	0.059	Autoimmune disorders treatment
0.348	0.035	<b>Pseudolysin inhibitor</b>
0.312	0.015	Antineoplastic (sarcoma)
0.431	0.140	Calcium channel (voltage-sensitive) activator
0.346	0.073	Plastoquinol-plastocyanin reductase inhibitor
0.364	0.096	<b>Omptin inhibitor</b>
0.362	0.099	Platelet derived growth factor receptor kinase inhibitor
0.410	0.199	<b>Bacterial efflux pump inhibitor</b>
0.333	0.142	CYP2D15 substrate

**Table 6.** The Predicted Spectrum of Biological Activity of Compound **8** (PASS) ( $Pa \geq 0.3$ )

<i>Pa</i>	<i>Pi</i>	Activity
0.674	0.071	Gluconate 2-dehydrogenase (acceptor) inhibitor
0.498	0.084	Chlordecone reductase inhibitor
0.354	0.035	Multiple sclerosis treatment
0.381	0.087	<b>Omptin inhibitor</b>
0.358	0.071	Autoimmune disorders treatment
0.372	0.097	CYP2D15 substrate
0.369	0.112	Antiinflammatory
0.428	0.174	<b>Pseudolysin inhibitor</b>
0.370	0.140	Ovulation inhibitor
0.391	0.227	CYP2H substrate
0.328	0.174	Octopamine antagonist
0.300	0.209	Thromboxane B2 antagonist

The antimicrobial activity of the new compounds was studied with respect to gram-positive *Staphylococcus aureus* 6538-P ATCC, *Staphylococcus aureus* 43300 ATCC (MRSA), *Streptococcus pyogenes* 19615 ATCC and gram-negative *Escherichia coli* 25922 ATCC, *Pseudomonas aeruginosa* 27853 ATCC test-strains of microorganisms.

Antimicrobial activity of trifluoroacetate 2,3,5-trimethyl-1*H*-indole-6-ammonium (**5**) (Table 7): against *S.aureus* 6538-P ATCC MIC=1.96 µg/mL; against *S.aureus* 43300 ATCC (MRSA) MIC=1.96 µg/

mL; against *E.coli* 25922 ATCC – 0.98 µg/mL; against *P.aeruginosa* 27853 ATCC – 0.98 µg/mL; against *S.pyogenes* 19615 ATCC – 0.98 µg/mL.

Antimicrobial activity of the trifluoroacetate 1,2,3,5-tetramethyl-1*H*-indole-6-ammonium (6) (Table 7): against *S.aureus* 6538-P ATCC MIC=7.9 µg/mL; against *S.aureus* 43300 ATCC (MRSA) MIC=7.9 µg/mL; against *E.coli* 25922 ATCC – 0.98 µg/mL; against *P.aeruginosa* 27853 ATCC – 3.9 µg/mL; against *S.pyogenes* 19615 ATCC – 31.3 µg/mL.

Antimicrobial activity of the trifluoroacetate 2,3-dimethyl-5-methoxy-1*H*-indole-6-ammonium (7) (Table 7): against *S.aureus* 6538-P ATCC MIC=31.2 µg/mL; against *S.aureus* 43300 ATCC (MRSA) MIC=31.2 µg/mL; against *E.coli* 25922 ATCC – 0.98 µg/mL; against *P.aeruginosa* 27853 ATCC – 7.9 µg/mL; against *S.pyogenes* 19615 ATCC – 125.0 µg/mL.

Antimicrobial activity of the trifluoroacetate 1,2,3-trimethyl-5-methoxy-1*H*-indole-6-ammonium (8) (Table 7): against *S.aureus* 6538-P ATCC MIC=125.0 µg/mL; against *S.aureus* 43300 ATCC (MRSA) MIC=125.0 µg/mL; against *E.coli* 25922 ATCC – 0.98 µg/mL; against *P.aeruginosa* 27853 ATCC – 1.96 µg/mL; against *S.pyogenes* 19615 ATCC – 0.98 µg/mL.

For **dioxidine** (comparison drug) against *Staphylococcus spp.* MIC=125.0–1000.0 µg/mL, against *Escherichia coli* 8.0–250.0 µg/mL, against *Pseudomonas spp.* 125.0–1000.0 µg/mL, *Streptococcus spp.* 64.0–1000.0 µg/mL (Padeiskaya 2011; Piopov et al. 2013).

## Conclusion

We continue studies to search for new compounds with an antimicrobial effect based on substituted 1*H*-indolylamines by targeted organic synthesis. Earlier, we synthesized the compounds of this series with a trifluoromethyl group in molecules, showing an effective antimicrobial activity. Previously investigated *N*-(indolyl) trifluoroacetamides 3 and 4 (Stepanenko et al. 2019) exhibited a high activity only against the gram-positive test strain of

*S.aureus*. Other derivatives of substituted 6-aminoindoles (Yamashkin et al. 2020) *N*-(indolyl) trifluoroacetamides C3, C4, X3 were highly effective against only gram-negative test strains of *E.coli* and *P.aeruginosa*. And only *N*-(indolyl) trifluoroacetamide based on substituted 6-aminoindoles X4 showed a high antimicrobial activity against gram-positive *S.aureus* and gram-negative *E.coli* and *P.aeruginosa* test strains of microorganisms.

Following the results of the out-of-experimental screening for biological activity, new water-soluble compounds based on substituted 1*H*-indol-6-ylamines with a predicted antimicrobial effect were obtained. The non-experimental PASS prediction of the biological activity correlates with the revealed antimicrobial activity of the compounds under study. The set of molecular mechanisms predicted *in silico* determines the ability of the studied indolyltrifluoroacetamides and trifluoroacetates based on substituted 1*H*-indol-5-ylamines to suppress the growth and reproduction of test strains of the studied microorganisms.

The structure of 1*H*-indolylammonium trifluoroacetates, not described in the literature earlier, has been reliably proved using modern methods of physicochemical analysis.

Laboratory microbiological screening showed that they have an antimicrobial effect that exceeds the activity of the reference drug, **dioxidine**. The presence of molecular mechanisms predicted *in silico* in the spectrum of biological activity of the studied compounds, such as Pseudolysin inhibitor, Omptin inhibitor, Undecaprenyldiphospho-muramoylpentapeptide beta-*N*-acetylglucosaminyltransferase inhibitor, UDP-epimerase inhibitor, Bacterial efflux pump inhibitor, suggests the presence of an antimicrobial activity against gram-positive and gram-negative microorganisms. But it must be remembered that the probability of *Pa* reflects, first of all, the similarity of the structure of molecules of a given organic compound with the structures of molecules that are most typical in the corresponding subset of “active” compounds in the training set (Varnek 2008; Filimonov et al. 2014). Therefore, as a rule, there is no direct correlation between the calculated

**Table 7.** The Sensitivity of the Test Strains of Microorganisms to New Compounds 5–8 (Mueller-Hinton broth serial dilution method)

Test-culture	Compound	<i>S.aureus</i> 6538-P ATCC				<i>S.aureus</i> 43300 ATCC (MRSA)				<i>E.coli</i> 25922 ATCC				<i>Paeruginosa</i> 27853 ATCC				<i>S.pyogenes</i> 19615 ATCC				
		5	6	7	8	5	6	7	8	5	6	7	8	5	6	7	8	5	6	7	8	
The concentration of a compound in the nutrient medium, µg/mL	125.0	0	0	0	+/- <sup>1</sup>	0	0	0	+/- <sup>1</sup>	0	0	0	0	0	0	0	0	0	0	0	+/- <sup>1</sup>	0
	62.5	0	0	+/-	+	0	0	+/-	+	0	0	0	0	0	0	0	0	0	0	+/-	+	0
	31.3	0	0	+/- <sup>1</sup>	+	0	0	+/- <sup>1</sup>	+	0	0	0	0	0	0	0	0	0	0	+/- <sup>1</sup>	+	0
	15.7	0	+/-	+	++	0	+/-	+	++	0	0	0	0	0	0	+/-	0	0	0	+	++	0
	7.9	0	+/- <sup>1</sup>	+	++	0	+/- <sup>1</sup>	+	++	0	0	0	0	0	+/-	+/- <sup>1</sup>	0	0	+	++	0	
	3.9	+/-	+	++	+++	+/-	+	++	+++	0	0	0	0	0	+/- <sup>1</sup>	+	+/-	0	++	+++	0	
	1.96	+/- <sup>1</sup>	+	++	+++	+/- <sup>1</sup>	+	++	+++	+/-	+/-	+/-	+/-	+/-	+	+	+/- <sup>1</sup>	+/-	++	+++	+/-	
	0.98	+	++	+++	+++	+	++	+++	+++	+/- <sup>1</sup>	+/- <sup>1</sup>	+/- <sup>1</sup>	+/- <sup>1</sup>	+/- <sup>1</sup>	++	+	+/- <sup>1</sup>	+++	+++	+++	+/- <sup>1</sup>	
	0.49	+	++	+++	+++	+	++	+++	+++	+	+	+	+	+	++	++	+	+	+++	+++	+	
	0.25	++	+++	+++	+++	++	+++	+++	+++	+	+	+	+	+	++	+++	++	+	+++	+++	+	
<b>Negative control</b>		+/-				+/-				+/-				+/-								

**Note:** <sup>1</sup> – activity titer, “+++” – diffuse cloud of MHB; “++” – moderate cloud of MHB; “+” – weak cloud of MHB; “+/-” – no cloud of MHB (as in the negative control); “0” – no growth when reseeded on MHA.

values of *Pa* and the quantitative characteristics of the activity. When analyzing the PASS-predicted spectra of biological activity, it is necessary to take into account the possibilities of experimental testing. Therefore, the general recommendation is to consistently study the various predicted biological activities, from the most likely to the least likely ones.

Trifluoroacetates 2,3,5-trimethyl-1*H*-indole-6-ammonium (5), 1,2,3,5-tetramethyl-1*H*-indole-6-ammonium

(6), 2,3-dimethyl-5-methoxy-1*H*-indole-6-ammonium (7), 1,2,3-trimethyl-5-methoxy-1*H*-indole-6-ammonium (8) may find application as water-soluble synthetic antimicrobial agents after additional studies.

## Conflict of interests

The authors declare no conflict of interests.

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