

Revision of *Agathacrista* new genus (Hymenoptera, Braconidae, Agathidinae, Agathidini)

Michael J. Sharkey^{1,†}, Stephanie A.C. Stoelb^{2,‡}

1 Department of Entomology, University of Kentucky, S225 Agricultural Science Center North, Lexington, KY 40546-0091, USA **2** Bluegrass Community & Technical College, Lexington, KY 40546s

† <http://zoobank.org/77B8EC3A-442C-4A7A-AF85-A31C27E257F2>

‡ <http://zoobank.org/9C4BCEB9-A6C7-4E7B-B9ED-334F7C8CE709>

Corresponding author: Michael J. Sharkey (msharkey@uky.edu)

Academic editor: G. Broad | Received 21 November 2012 | Accepted 14 March 2012 | Published 1 August 2013

<http://zoobank.org/2B514381-7262-4609-8441-EEDE2AF235E9>

Citation: Sharkey MJ, Stoelb SAC (2013) Revision of *Agathacrista* new genus (Hymenoptera, Braconidae, Agathidinae, Agathidini). Journal of Hymenoptera Research 33: 99–112. doi: 10.3897/JHR.33.4373

Abstract

Based on a cladistic analysis, a new genus of Agathidini, *Agathacrista* Sharkey, is proposed and its phylogenetic position hypothesized. Two previously described (*Agathacrista cancellata*, *Agathacrista depressifera*) and three new species (*Agathacrista sailomi*, *Agathacrista winloni*, *Agathacrista krataeii*) are included. The distribution of *Agathacrista* is limited to the Oriental region and southern portion of the eastern Palearctic.

Keywords

Insecta, identification key, taxonomy, systematics

Introduction

Agathacrista, as proposed here, includes two previously described species, both of which were included in the paraphyletic concepts of *Bassus s.l.* and *Therophilus s.l.* This paper is part of a series that investigates these non-monophyletic taxa, while describing taxa from Thailand, Costa Rica, or more inclusive areas of the world.

Methods

Morphological terms: The length of the first metasomal tergite is measured from the apex of the tendon emanating from the propodeum to the posterior border of the tergite. Other terms are from Sharkey and Wharton (1997).

Museum acronyms

- HIC** Hymenoptera Institute Collection, University of Kentucky, Department of Entomology, Lexington, Kentucky, USA.
- QSBG** Queen Sirikit Botanic Gardens, Chiang Mai, Thailand.
- TARI** Taiwan Agricultural Research Institute, Entomology Collection, Taichung, Taiwan.

Species description format: Descriptions are of the holotype; variation is given in parentheses. In the species descriptions color is not extensively described because the images serve this purpose; however color characters that are variable or of diagnostic significance are detailed.

Species delimitation: The five species from Thailand treated here are very similar morphologically and are represented by only seven specimens. The seven specimens do not differ more than might be expected within one species of agathidine. In part, we relied on 28S sequence data to separate species. We did not employ any particular cut-off point in differences in 28S but rather calibrated the amount of sequence difference using species that exhibit significant morphological and host preference differences. This calibration comes from a large dataset of about 1,000 agathidine specimens for which we have 28S data. The three species of *Agathacrista* for which we have 28S sequences were identified according to this criterion. Morphological characters also played an important role in species delimitation. We examined ten specimens of *Agathacrista cancellata* (Enderlein, 1920), all from Taiwan. Body color varies considerably in this species, but wing color and the sculpture and dimensions of the first metasomal median tergite are uniform, suggesting that these are good species indicators within the genus. Therefore, these characters played an important role in our species concepts. *Agathacrista depressifera* (van Achterberg & Long, 2010) displays considerable variation in wing color-pattern. Despite the fact that we have some evidence to suggest that this species concept may include more than one species, our evidence is weak and not convincing enough to suggest changes. This revision of *Agathacrista* is very preliminary; many more specimens will be needed to fully understand species limits and species diversity.

Generic concepts: This is the fourth of a series of papers (Sharkey and Clutts 2011; Sharkey and Stoelb 2012; Sharkey et al. 2009) that reclassify what was once considered the genus *Bassus* (or *Microdus*, or *Agathis s.l.*). I (MJS) thought it best to give some details here on the criteria that were used to delineate generic limits. Most of these are stated in one manner or another in Backlund and Bremer (1998). The first considera-

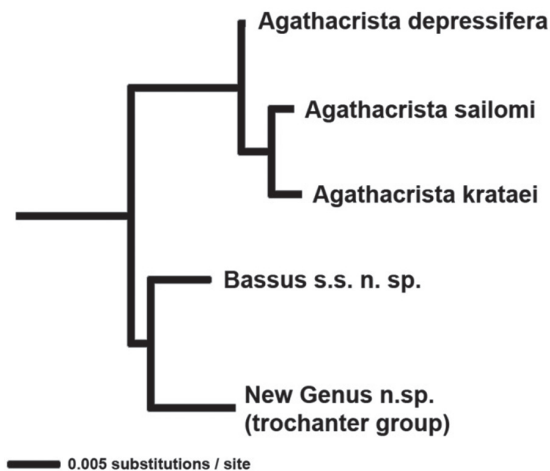


Figure 1. NJ tree showing 28S distances between species of *Agathacrista*.

tion is monophyly and this is why the polyphyletic concept of *Bassus s.l.* was rejected (Sharkey et al. 2009). The following criteria are in no particular order and may be more or less important depending on the genus.

Identifiability: *Agathacrista* is a good example of the utility of this criterion. *Agathacrista*, *Bassus s.s.* and a new genus to be described in a forthcoming paper, form a monophyletic group (Fig. 2). The only morphological synapomorphy to aid in the recognition of the group is the presence of small spines or pegs on the fore tibia; convergent in some members of *Earinus* and many other non-agathidines. If there was an easily observed character it might be best to expand the concept of *Bassus* to include *Agathacrista* and the new genus. Conversely, all three genera as here proposed have distinguishing characteristics. *Bassus s.s.* has long, simple claws that lack a basal lobe; *Agathacrista* has a crest between the antennae and the undescribed new genus has a distinct longitudinal carina on the hind trochanter, a character formerly thought to be restricted to some members of the Cremnoptini and Disophrini.

Biological information content: Typically the more that is known about organisms the more finely split the higher ranks are. This criterion has little application to any agathidines since we know so little about their natural history. Undoubtedly the morphological autapomorphies of the three genera referred to above will be shown to relate to unique behaviours.

Stability: This is somewhat tied to most other criteria and *Bassus s.l.* is a good example of an unstable concept. Over the years it has been included in the genus *Agathis s.l.* and numerous genera like *Camptothlipsis* and *Therophilus* have been included, or not. This instability has made information retrieval on any particular species rather cumbersome, as the number of different combinations they have been referred to is a minimum of three, i.e., *Microdus*, *Bassus*, *Agathis*, the foremost being a junior objective synonym of *Bassus*. Thus, generic concepts should be accompanied by solid phylogenetic evidence when such is available.

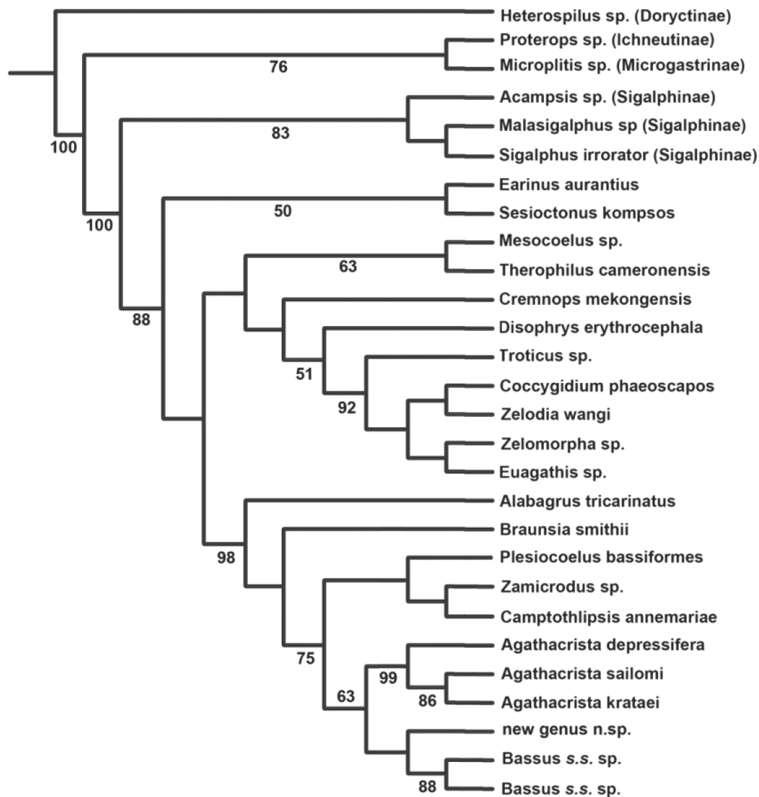


Figure 2. Maximum parsimony tree of some agathidine genera with a concentration near *Agathacrista* based on D2-D3 regions of 28S. Numbers below branches are bootstrap values; only values ≥ 50 are shown.

Phylogenetic evidence: This is directly related to stability. If new, but weak evidence suggests the non-monophyly of a well-recognized genus, it is usually best to wait for more data before making formal changes. In the case presented here, there is strong molecular phylogenetic evidence suggesting that the broad concept of *Bassus* (*Bassus s.l.*) is polyphyletic and there is significant morphological and molecular evidence for the monophyly of *Agathacrista*.

Specimen collection: As part of the inventory of Thai insects, 3 Malaise traps at each of 30 different localities throughout Thailand were operated from 2007-2010, comprising approximately 90 trap-years. The specimens dealt with here are primarily from these traps.

Phylogenetic methods: Regions D2-D3 of 28S rDNA (roughly 560 base pairs) were sequenced using the following primers: 28SD2hymF 5' - AGAGAGAGTTCAA-GAGTACGTG - 3' and 28SD3hymR 5' - TAGTTCACCATCTTTTCGGGTC - 3'. Sequences were edited using Geneious Pro v4.7.5 (Drummond et al. 2009) and aligned based on a secondary structure model for Ichneumonoidea developed by Yoder and Gillespie (2004) and Gillespie et al. (2005). Regions of expansion and contraction

(RECs), regions of slipped-strand compensation (RSCs), and short regions of alignment ambiguity were further aligned/corrected by eye. Three of these regions (~30 base pairs) were deleted because they could not be aligned with confidence, i.e., there were multiple equally supported alignment options.

The NJ analysis (Fig. 1) was conducted with PAUP* (Swofford 2003) using default settings. The parsimony analysis (Fig. 2) was performed using TNT (Goloboff et al. 2008) [traditional search with 100 random addition sequences followed by branch-swapping, saving 100 trees per replication; 1000 bootstrap replications were used to estimate branch reliability].

Morphological terms used in this revision were matched to the Hymenoptera Anatomy Ontology (HAO; Yoder et al. 2010) (Appendix 4). Identifiers (URIs) in the format http://purl.obolibrary.org/obo/HAO_XXXXXXX represent anatomical concepts in HAO version <http://purl.obolibrary.org/obo/hao/2011-05-18/hao.owl>. They are provided to enable readers to confirm their understanding of the anatomical structures being referenced. To find out more about a given structure, including images, references and other metadata, use the identifier as a web-link, or use the HAO:XXXXXXX (note colon replaces underscore) as a search term at <http://glossary.hymao.org>.

All species are treated with a diagnosis and distributional data. They are illustrated with color photos using a JVC digital camera mounted on a Leica MZ16 microscope and Automontage® stacking software. Distributional data are listed for all species and a link to a Google map is included for all species. The descriptions are of holotypes and variation is given in parentheses.

Phylogenetic considerations

Sharkey et al. (2006) showed that the entities formerly referred to as *Bassus s.l.* fall into two widely disparate clades. One associated with the genus *Mesocoelus*, *Therophilus* and allies and the other with *Bassus s.s.*, *Agathis*, *Alabagrus*, *Braunsia* and allies. *Agathacrista* falls into the latter lineage. It is sister to *Bassus s.s.* + a new genus to be described in another paper. All three genera are restricted to the Old World, and except for one species of *Bassus s.s.*, *B. calculator* Fabricius, 1798, the type species, which is widely distributed throughout the Palearctic, all are restricted to the Oriental region, the northern, tropical parts of the Australian region, and the far eastern Palearctic.

Members of all three genera have distinct pegs on the anterior surface of the fore tibia (Fig. 3a), a synapomorphy. This character state is convergent in some members of the distantly related genus *Earinus* (Agathidinae), some Braconinae, most Doryctinae, and has rare and scattered appearances in other braconid subfamilies. Members of *Agathacrista* and *Bassus* have similar and rather unique color patterns (see Figs 4–8). The thin, acute, interantennal crest (Fig. 3b) is an autapomorphy for *Agathacrista* that is found convergently though rarely in a few other agathidine genera, e.g., a few members of *Therophilus*.

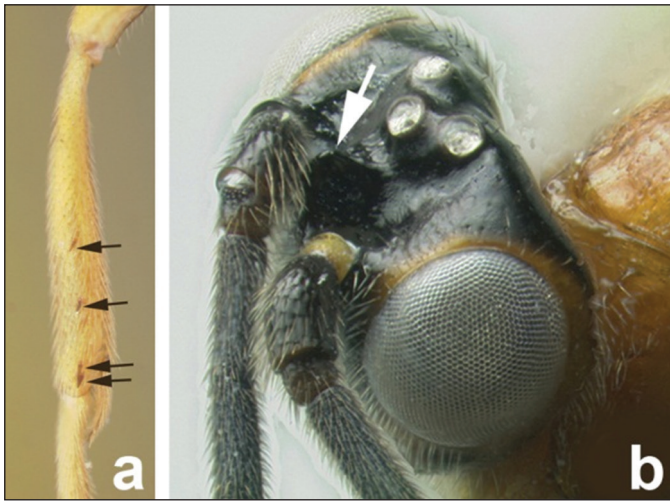


Figure 3. **a** fore leg of *Agathacrista* sp. showing pegs or spines **b** dorsolateral head of *Agathacrista* sp. showing interantennal crest.

Taxonomy

Agathacrista Sharkey, gen. n.

<http://zoobank.org/0E5C3C07-22D0-45B8-8B9A-4BA142EE1731>

<http://species-id.net/wiki/Agathacrista>

Type species. *Agathacrista winloni* Sharkey, sp. n.

Diagnosis. Interantennal space with longitudinal keel that is sharply declivous posteriorly (Figs 3b, 7d); ocelli elevated above surface of vertex; fore tibia with pegs (Fig. 3a); third metasomal median tergite lacking sculpture; basal lobe of tarsal claws large and right angled or slightly acute (Fig. 7a); fore wing partly or completely pigmented; hind wing vein CUB absent. Some members of a few other genera of Agathidini have a sharp keel between the antennae, but none of these have pegs on the fore tibia.

Description. Head. Lateral carina on frons (as in members of *Alabagrus*) absent; interantennal space with longitudinal keel that is sharply declivous posteriorly (Figs 3b, 7d); ocelli elevated above surface of vertex; gena not extended ventroposteriorly into sharp prominence (Fig. 6a); gena lacking sharp angle posteriad of eye; labial palpus with 4 segments, third segment less than $\frac{1}{2}$ as long as apical segment; apical antennomere acute but lacking nipple-like process. **Mesosoma.** Propleuron convex ventrally but lacking a sharp bump; notauli impressed and pitted, at least in part (Fig. 6c); posteroscutellar depression absent but sculpture usually present in this area; propodeum rugose to areolate-rugose (Fig. 7g); sclerite between hind coxal

cavities and metasomal foramen complete, its ventral margin situated ventral to dorsal margin of hind coxal cavities. **Legs.** Fore tibia with pegs (Fig. 3a); all tarsal claws with strong basal lobe and with right-angled or slightly acute angle (Fig. 7a). **Wings.** (Figs 6d, 8b). Fore wing RS+M vein mostly absent; second submarginal cell triangular; fore wing 3RSb straight and strong throughout; hind wing r and r-m cross veins absent; hind wing vein CUB absent; fore wing partially or completely pigmented, with yellow or melanic color. **Metasoma.** First median tergite longitudinally striate, with a slightly stronger pair of lateral striae and a medial stria (Fig. 7h, 8e), rarely striae reduced and sculpture mostly weakly rugose (see couplet 1 in the identification key); second median tergite with an elevated semi-circular area separated from the remainder of the tergite by a shallow groove (Fig. 8e); second median tergite from smooth to striate, usually smooth anteriorly with a few longitudinal striae in posterior half; median tergite 3 smooth; length of ovipositor rather uniform, as long as (Fig. 5) or slightly longer than body (Fig. 8c).

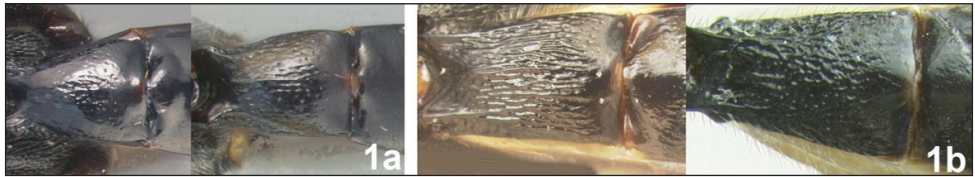
Biology. Unknown

Distribution. Restricted to the eastern Palearctic (Taiwan) and the Oriental region. For a distribution map, click here.

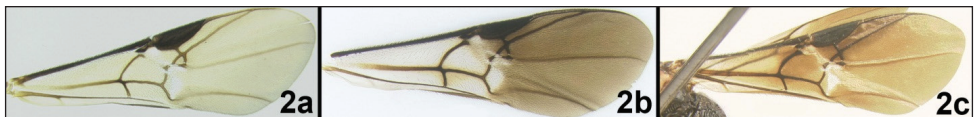
Etymology: From the Greek *agathis*, meaning “ball of thread”, and the Latin *crista*, meaning “crest”. Crest is in reference to the ridge between the antennal insertions. The gender is feminine.

Key

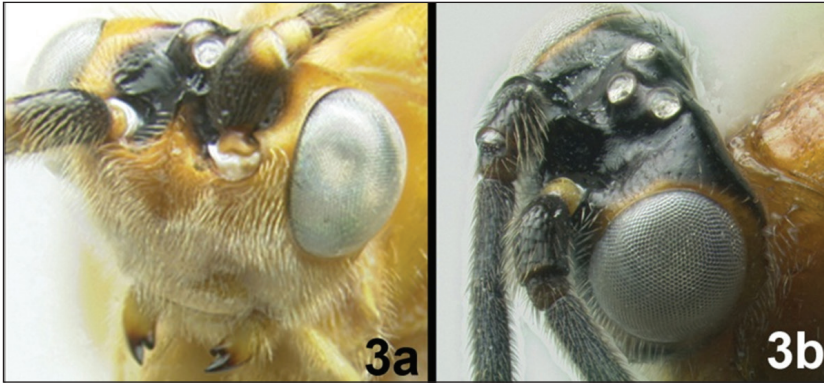
- 1a Median tergite 1 mostly smooth or weakly sculptured...*Agathacrista cancellata*
- 1b Median tergite 1 mostly striate or rugosostriate..... 2



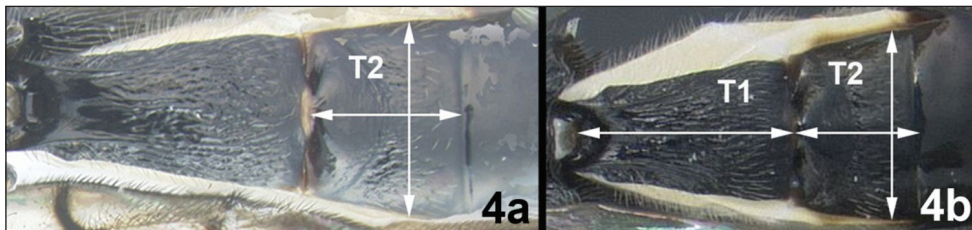
- 2a Fore wing uniformly yellow, at most slightly darker apically 3
- 2b Fore wing clear basally, infusate distally 4
- 2c Fore wing mostly evenly infusate, very slightly clearer at extreme base.....
..... *Agathacrista depressifera*



- 3a Head mostly yellow, black color restricted to medial dorsal surface; larger specimens, ~ 9.6 mm *Agathacrista sailomi*
- 3b Head mostly black, yellow color restricted to part of face and orbits of eyes; smaller specimens, ~ 6.2 mm *Agathacrista depressifera*



- 4a Median tergite 1 (T1) weakly converging anteriorly; T2 quadrate and ~1.3× wider than long..... *Agathacrista winloni*
- 4b Median tergite 1 strongly converging anteriorly; T2 diverging posteriorly and ~1.5× wider than long..... *Agathacrista krataeii*



Species descriptions

Agathacrista cancellata (Enderlein), comb. n.
http://species-id.net/wiki/Agathacrista_cancellata

Fig. 4

Agathis cancellata Enderlein, 1920

Diagnosis. This species has the northern-most distribution and appears to be restricted to Taiwan where it is the only species present. Members come in two color morphs; one with the mesosoma bicolored and the other with the mesosoma entirely melanic. *Agathacrista cancellata* differs from all other members of the genus by the relatively smooth surface of metasomal median tergite 1 (Fig. 1a in the identification key above).

Material examined. Taiwan: Female: “Kanshirei, 20 xi 1928, col. J Sonan” (HIC). “Kanshirei, 10 xi 1926, col. J Sonan” (TARI). Female: “Taichung, Wufeng,

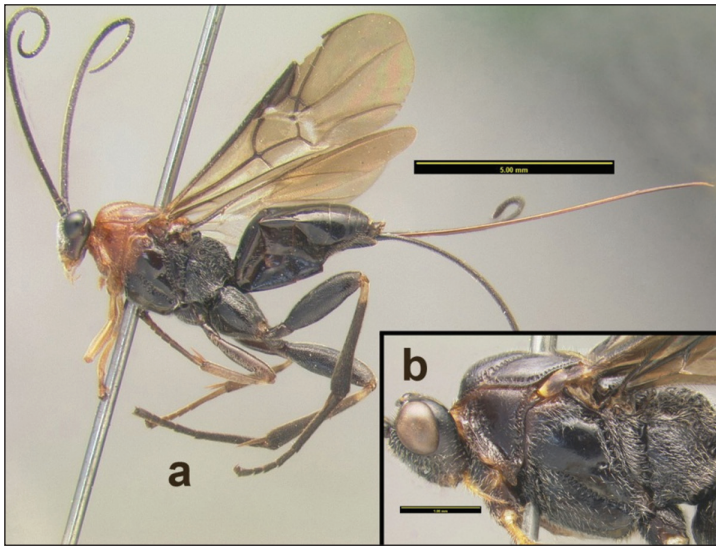


Figure 4. *Agathacrista cancellata*. **a** lateral habitus **b** lateral head and mesosoma.

Wanfeng, 2-7-v-1979, K.C. Chou" (HIC). Female: "Puli, Taichung. 16-19 v. 1956, K.S. Lin." (HIC). Female: "Taipei, 15.v.1964 K.S. Lin" (TARI). Male: "Shanti-men, Pingtung Hsien, 28.III.-1.IV.1981, C.C. Chen & C.C. Pang" (TARI). Female: "12.x.1952, Coll. S. C. Cheu" (TARI). Female: "Arisan, 5-VIII-1981, Col. T. Shiraki" (TARI). Female: "Chungho, 28.V.1963, Col. H.H. Tseng" (TARI). Female: "Suo-Suigen 24.VIII.1930, Col. J. Sonan" (HIC). For a distribution map, click here.

***Agathacrista depressifera* (Achterberg & Long), comb. n.**

http://species-id.net/wiki/Agathacrista_depressifera

Fig. 5

Therophilus depressiferus Achterberg & Long, 2010

Description. Length: 6.5 (7.8) mm. Antenna with 38 (42) flagellomeres. Number of spines on fore, mid, and hind tibiae 6(5), 8(11), 15(16). First median tergite 1.6 times longer than wide; second median tergite 1.2 times wider than long; median tergite 1 rugosostriate, except smooth near posterior margin, sculpture more reticulated than congeners; median tergite 2 smooth (weakly rugosostriate near midlength). Color: (melanic color of mesopleuron more extensive in the sole paratype.)

Material examined. **Vietnam:** Female: "Dak Lak Prov. Krong Bong, Chu Yan Sin NP, N12°31.303', E108°28.853', 28.vii.2008 MT" H7991 (QSBG). **Thailand:** Female: "Phetchabun, Khao Kho NP, 520m, 16°52.568'N, 101°08.104'E, 12-19.x.2006 Sintong leg. T807" H002 (HIC). For a distribution map, click here.

Molecular data. Genbank Accession KC556782 (H002).



Figure 5. *Agathacrista depressifera* lateral habitus.

***Agathacrista krataei* Sharkey, sp. n.**

<http://zoobank.org/FCDD9F11-0FF1-4B90-A7AB-5A75BE08A235>

http://species-id.net/wiki/Agathacrista_krataei

Fig. 6

Description. Length: 8.4 mm. Antenna with 41 flagellomeres. Number of spines on fore, mid, and hind tibiae 6, 10, 13. First median tergite 1.5 times longer than wide; second median tergite 1.5 times wider than long; median tergite 1 entirely rugosostriate; median tergite 2 mostly weakly sculptured with striae, smooth posterolaterally.

Etymology. Named after Kratae Sanok a collector for the TIGER project at Pa Hin Ngam National Park.

Material examined. Holotype: Female: *Thailand*, “Sakon Nakhon, Phu Phan NP, Dry evergreen near house no. 1567, 16°48.628'N, 103°53.591'E, Malaise trap, vi.2007, Winlon Kongnara leg., T2502” H268 (QSBG). For a distribution map, click here.

Molecular data. Genbank Accession KC556781 (H268).

***Agathacrista sailomi* Sharkey sp. n.**

<http://zoobank.org/61CEE178-5F5E-4302-8A10-E4AE769CAF12>

http://species-id.net/wiki/Agathacrista_sailomi

Fig. 7

Description. Length: 9.6(9.3) mm. Antenna with 44 flagellomeres. Number of spines on fore, mid, and hind tibiae 4(3), 6, 6(13). First median tergite 1.6 times longer than wide; second median tergite 1.4 times wider than long; median tergite 1 rugosostriate, except smooth near posterior margin; median tergite 2 mostly smooth with a weak indication of longitudinal sculpture at midlength. Color: (Melanic color of head restricted to ocellar triangle in the sole paratype.)

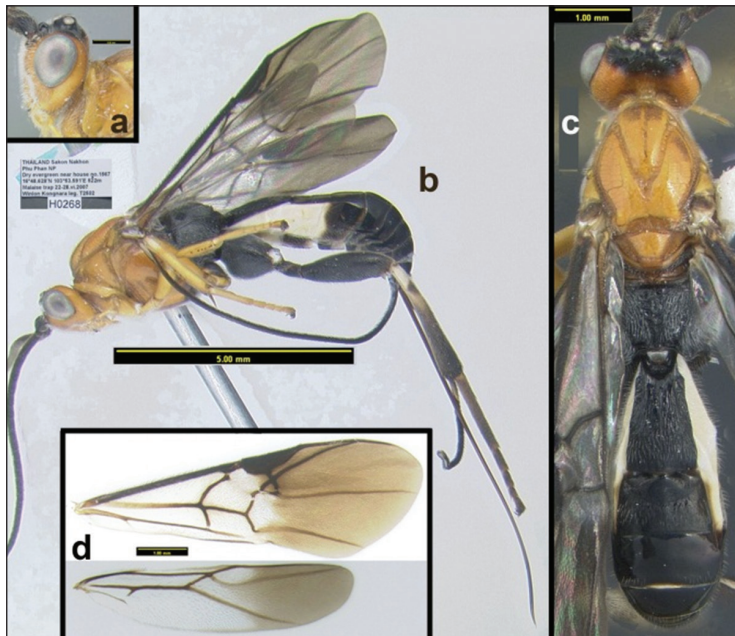


Figure 6. *Agathacrista krataei*. **a** lateral head **b** lateral habitus **c** dorsal habitus **d** wings.

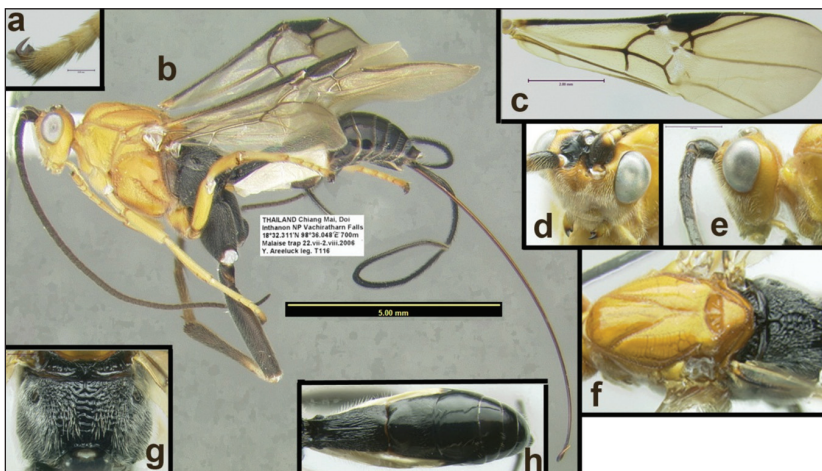


Figure 7. *Agathacrista sailomi*. **a** tarsal claw **b** lateral habitus **c** fore wing **d** anterolateral head **e** lateral head **f** dorsal mesosoma **g** dorsal propodeum **h** dorsal metasoma.

Etymology. Named after Sailom Tongbunchai a collector for the TIGER project at Phuphan National Park.

Material examined. Holotype: Female: *Thailand:* “Chiang Mai, Doi Inthanon NP, Vachiratharn Falls, 18°32.311'N. 98°36.048'E, 700m Malaise trap, 6–13.ix.2006 Y. Areeluck leg. T242” H013 (QSBG).

Molecular data. Genbank Accession KC556780 (H013).

Paratypes. Female: **Thailand:** “Chiang Mai, Doi Inthanon NP, Vachiratharn Falls, 18°32.311'N, 98°36.048'E, 700m Malaise trap, 22.vii–2.viii.2006 Y. Aree-luck leg. T116” H284 (HIC). Female: “Chiang Mai, 20.ix–20.x.(19)84, D. Jackson” H5904 (QSBG). For a distribution map, click here.

***Agathacrista winloni* Sharkey, sp. n.**

<http://zoobank.org/3556862D-AC1D-428F-818D-B8CA315212D1>

http://species-id.net/wiki/Agathacrista_winloni

Fig. 8

Description. Length: 8.4 (9.8) mm. Antenna with 43(45) flagellomeres. Number of spines on fore, mid, and hind tibiae 6, 8, 12(15). First median tergite 1.8 times longer than wide; second median tergite 1.2 times wider than long; median tergite 1

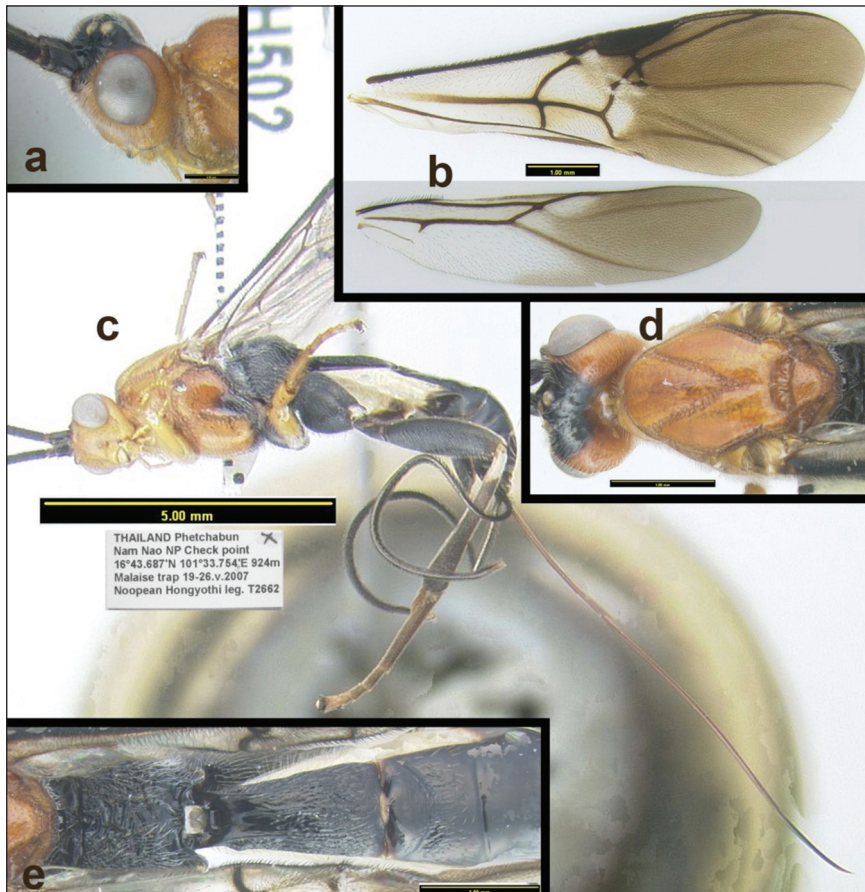


Figure 8. *Agathacrista winloni*. **a** lateral head **b** wings **c** lateral habitus **d** dorsal head and mesosoma **e** dorsal propodeum and metasomal median tergites 1–3.

rugosostriate, except smooth near posterior margin; median tergite 2 mostly smooth with a weak indication of longitudinal sculpture at midlength (almost entirely smooth). Color: (melanic color of mesopleuron more extensive in one paratype; mid femur melanic in basal half).

Etymology. Named after Winlon Kongnaka a collector for the TIGER project at Phuphan National Park.

Material examined. Holotype: Female: *Thailand*: “Phetchabun, Nam Nao National Park Check Point, 16°43.687'N, 101°33.754'E, 924m 19–26.v.2007 Noopean Hongyothi leg. T2662” H502 (QSBG).

Molecular data. Genbank Accession KC771135 (H502).

Paratypes. Female: *Thailand*: “Phetchabun, Nam Nao National Park Check Point, 16°43.687'N, 101°33.754'E, 924m 19–26.v.2007, Noopean Hongyothi leg. T2662” H501 (HIC). Female: “Phetchabun, Nam Nao NP Sam Makao forest, 16°41.067'N, 101°40.425'E, 540m, Malaise trap 4–11.ix.2006 Higgyothi & Janteab leg.” H466 (QSBG). Female: “800m, 180 km NE Bangkok Khao Yai Nat. Park 10-16.iv.1990 B.V. Brown, Moist evergreen for(est). MT.” (HIC). For a distribution map, click here.

Acknowledgements

We thank the staff at Queen Sirikit Botanic Garden in Chaing Mai for sorting the many hundreds of samples and for the Thai park staff for operating Malaise traps and other collection devices. A special thanks to Chaweewan Hutacharern for managing the Thai end of the TIGER project. Also many thanks to the curators around the world (London, Oxford, Warsaw, Leiden, Gainesville, Budapest, Beijing, Paris, Stockholm, Washington) for lending specimens and hosting MJS on visits. Special thanks to Kees van Achterberg for lending specimens and types from Vietnam. Funding was provided by NSF grants DEB-0542864 and EF-0337220.

References

- Backlund A, Bremer K (1998) To be or not to be - principles of classification and monotypic plant families. *Taxon* 47: 391–401. doi: 10.2307/1223768
- Bhat S, Gupta VK (1977) The subfamily Agathidinae (Hymenoptera, Braconidae) *Ichneumonologia Orientalis*. *Oriental Insects Monograph* 6: 1–353.
- Drummond AJ, Ashton B, Cheung M, Heled J, Kearse M, Moir R, Stones-Havas S, Thierer T, Wilson A (2009) Geneious v4.7, <http://www.geneious.com/>
- Gillespie JJ, Munro JB, Heraty JM, Yoder MJ, Owen, AK, Carmichael AE (2005) Secondary structural model of the 28S rRNA expansion segments D2 and D3 for chalcidoid wasps (Hymenoptera: Chalcidoidea). *Molecular Biology and Evolution* 22, 1593-1608. doi: 10.1093/molbev/msi152

- Goloboff PA, Farris JS, Nixon KC (2008) TNT, a free program for phylogenetic analysis. *Cladistics* 24: 774–786. doi: 10.1111/j.1096-0031.2008.00217.x
- Sharkey MJ (1992) Cladistics and tribal classification of the Agathidinae (Hymenoptera: Braconidae). *Journal of Natural History* 26: 425–447. doi: 10.1080/00222939200770251
- Sharkey MJ, Clutts SA (2011) A revision of Thai Agathidinae (Hymenoptera: Braconidae), with descriptions of six new species. *Journal of Hymenoptera Research* 22: 69–132. doi: 10.3897/jhr.22.1299
- Sharkey MJ, Clutts SA (2012) A revision of *Zelodia* (Hymenoptera: Braconidae: Agathidinae) from Thailand. *Journal of Hymenoptera Research* 26: 31–71. doi: 10.3897/jhr.26.2527
- Sharkey MJ, Laurenne NM, Sharanowski B, Quicke DLJ, Murray D (2006) Revision of the Agathidinae (Hymenoptera: Braconidae) with comparisons of static and dynamic alignments. *Cladistics* 22: 546–567. doi: 10.1111/j.1096-0031.2006.00121.x
- Sharkey MJ, Stoelb SAC (2012) Revision of *Therophilus s.s.* (Hymenoptera, Braconidae, Agathidinae) from Thailand. *Journal of Hymenoptera Research* 27: 1–36. doi: 10.3897/jhr.27.2832
- Sharkey MJ, Wharton RA (1997) Morphology and terminology. In: Wharton RA, Marsh PM, Sharkey MJ (Eds) *Manual of the New World genera of Braconidae (Hymenoptera)*. Special Publication of the International Society of Hymenopterists. Vol. 1: 19–38.
- Sharkey MJ, Yu DS, van Noort S, Seltmann K, Penev L (2009) Revision of the Oriental genera of Agathidinae (Hymenoptera, Braconidae) with an emphasis on Thailand including interactive keys to genera published in three different formats. *ZooKeys* 21: 19–54. doi: 10.3897/zookeys.21.271
- Swofford DL (2003) PAUP*. *Phylogenetic Analysis Using Parsimony (*and Other Methods)*. Version 4. Sinauer Associates, Sunderland, Massachusetts.
- van Achterberg C, Long KD (2010) Revision of the Agathidinae (Hymenoptera, Braconidae) of Vietnam, with the description of forty-two new species and three new genera. *ZooKeys* 54: 1–184. doi: 10.3897/zookeys.54.475
- Yoder M, Gillespie J (2004) jRNA. Exploring insect phylogeny using RNA secondary structure. <http://hymenoptera.tamu.edu/rna>
- Yoder MJ, Mikó I, Seltmann KC, Bertone MA, Deans AR (2010) A Gross Anatomy Ontology for Hymenoptera. *PLoS ONE* 5(12): e15991. doi: 10.1371/journal.pone.0015991
- Yu DS, van Achterberg C, Horstmann K (2005) *Taxapad 2004: World Ichneumonoidea, Taxonomy, biology, morphology and distribution*. Vancouver, Canada. www.taxapad.com