

Angiosperms, Kuhdasht gypsum areas, Lorestan, Iran

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ABSTRACT: There is a lack of information on flora of gypsophilous plants in gypsum habitat in Lorestan province. In this paper, we report a species list of the gypsum flora of Kuhdasht, Iran. The study took place between 2009 and 2010. Plant species were identified and their chorology and life form determined through laboratory examinations and using reference books. We recorded about 1,000 specimens belonging to 39 families, 137 genera, and 190 taxa. An overall of 14 taxa (7.36%) are endemic to Iran. Asteraceae (29 taxa), Poaceae (24 taxa), and Fabaceae (19 taxa) were the richest taxa. The two largest genera are *Gypsophila* L. and *Astragalus* L., with six and five species, respectively. Irano-Turanian elements were the most dominant chorotypes (48.43%). Also, therophytes (51.58%) and hemicryptophytes (30.53%) were the most abundant life forms.

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INTRODUCTION

Gypsophilous plants are one of the most specific xerophilous plants. They are usually rare and endangered species. Gypsum affects plant growth and development. The removal of gypsum layers prevents vegetation from increasing in density (Pueyo and Alados 2007). Gypsum presumably decreases moisture stress during droughts of early summer, due either to decreased water competition because of low densities or inherent characteristics of the gypsum soil (Meyer 1986).

Areas with gypsum soil cover about 9.816 million ha of Iran, which represents about 16% of country area. Gypsum ($\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$) is broadly scattered on the soil surface in arid areas with less than 400 mm mean annual rainfall (Eftekhari and Assadi 2001; Eftekhari *et al.* 2002), where gypsum resources are one of the customary and rich minerals of soils (FAO 1990). Gypsiferous soils are those with at least 5% of gypsum (Soil Survey Staff 1999).

Studies of flora in gypsum soils are scarce. Johnston (1941) first reported that vascular plants are rare in gypsum soil in the Chihuahuan Desert of northern Mexico. This pattern has been seen in several arid and semi-arid areas of the world (Parsons 1976). Nevertheless, the selective forces for the evolution of gypsophile endemics have not been clearly identified (Powell and Turner 1977).

Floristic surveys are particularly important in the Lorestan province, for its ecological and weather conditions. Additionally, floristic lists are a first step for protecting endangered plants and planning a sustainable use of, for example, medicinal plants. Several studies have been conducted on the flora of non-gypsum areas in Kuhdasht, Lorestan province, Iran (Azadbakht *et al.* 2007; Dehshiri *et al.* 2011). Some previous studies have also been conducted in gypsum areas in Iran, such as Eftekhari and Assadi (2001) in Semnan province, and Eftekhari *et al.* (2002) in Khorasan province. Here, we identified Gypsophilous plants in Kuhdasht, Lorestan province,

Iran. Results of this study can also be useful for rangeland management and conservation.

MATERIALS AND METHODS

Study site

The Lorestan province is in the Irano-Turanian phytogeographic region in western Iran. It has hot and dry summers, wet and cold winters. The Kuhdasht region is in western Lorestan. The studied area covers about 3962.8 ha in southwestern Kuhdasht ($47^{\circ}21'$ and $47^{\circ}30'$ E; $33^{\circ}22'$ and $33^{\circ}25'$ N; Figure 1). The area has an arid and cold climate (Sabeti 1969) that is common in western Iran. The studied area has a rough footed topography and its altitude ranges from 1,500 to 1,800 m. Soil texture is sandy clay, silty clay or clay; pH is about 7.2–8.2; Electrical conductivity was 0.78–3.1 ds m^{-1} ; lime is 5–38.6%; soil fertility is low; gypsum is 0.5–63.4%; nitrogen is highly limited, whereas potassium is nearly sufficient (Dehshiri *et al.* 2011).

Data collection

Plant specimens were collected from areas where gypsum is dense and clearly seen on the surface. We collected specimens of vascular plants in the study area during growth periods between 2009 and 2010. They were dried by means of standard herbarium methods. Specimens were identified by using the *Flora Iranica* (Rechinger 1965–2010), *Flora of Iran* (Assadi *et al.* 1988–2010; Ghahreman 1975–2006), and additional guides (Boissier 1867–1888; Komarov and Shishkin 1934–1960; Tutin *et al.* 1964–1980; Davis 1965–1988; Zohary 1966–1986; Townsend and Guest 1966–1985; Ghahreman 1990–1994; Maassoumi 1986–2005; Mozaffarian 2008). Threatened statuses are proposed for endemic taxa following Jalili and Jamzad (1999), and Ghahreman and Attar (1999). Life forms followed the Raunkiaer's life form specifications (Mueller-Dombois and Ellenberg 1974; Asri 2003). Phytogeographical regions were also determined

by using several reference books (Rechinger 1965–2010; Assadi *et al.* 1988–2010; Akhani 2005; Davis 1965–1988; Komarov and Shishkin 1934–1960; Townsend and Guest 1966–1985; Tutin *et al.* 1964–1980; Zohary 1966–86; Zohary 1973; Takhtajan 1986). Specimens are deposited at the Central Herbarium of Islamic Azad University, Borujerd Branch.

RESULTS AND DISCUSSION

Floristic richness and taxonomic diversity

About 250 plant species have been recorded in gypsum areas in Iran (e.g., Rechinger 1965–2010; Assadi *et al.* 1988–2010; Eftekhari and Assadi 2001; Eftekhari *et al.* 2002). However, information on habitats of some plants are either absent or not given in detail. We collected about 1,000 angiosperm plant specimens in the study area, comprising 190 taxa, belonging to 137 genera of 39 families (Tables 1–2). Of these, 31 were Monocotyledons and 159 taxa were Dicotyledons. Of all taxa, 165 have not been reported yet for Semnan and Khorasan provinces (Eftekhari and Assadi 2001; Eftekhari *et al.* 2002).

The largest nine families are Asteraceae 29, Poaceae 24, Fabaceae 19, Lamiaceae 14, Brassicaceae 12, Apiaceae 9, Caryophyllaceae 9, Boraginaceae 7, and Rosaceae 6 (Table 1). Approximately 67% of the taxa belong to these nine families. These families are also the largest families in the Iran and Irano-Turanian floras (Ghahreman and Attar 1999), which explains their higher representation. Conversely, species from Asteraceae, Poaceae, and Fabaceae are the main bulk of alien plants in Iran, and also in agro-ecosystems of adjacent countries, such as Saudi Arabia and Kuwait (Abd El-Ghani and El-Sawaf 2004; Abd El-Ghani and Abd El-Khalik 2006). This result was also found by previous studies (e.g., Hoagland and Buthod

2005; Barber 2008; Akpulat and Celik 2005).

Asteraceae species have great ecological tolerance and break up their seeds easily (Akpulat and Celik 2005; Abd El-Ghani and Abd El-Khalik 2006). It is the second large family in the flora of Iran (Ghahreman and Attar 1999), and also the largest and most widespread family of flowering plants in the world (Good 1974). Additionally, the extensive occurrence of *Centaurea* and *Crepis* in gypsum soils contributed to this pattern.

Poaceae is third largest family in the flora of Iran (Ghahreman and Attar 1999). Species of this family tolerate soils with 40% of gypsum (Eftekhari and Assadi 2001; Eftekhari *et al.* 2002). In addition, *Hordeum* was the genus with highest occurrence. This provides evidence that monocotyledons are dominant groups in gypsum areas.

Fabaceae ranks as the third most speciose family in our study and is the largest family in the flora of Iran (Ghahreman and Attar 1999). Species of this family are calciphilous and grows easily in gypsum places (Eftekhari and Assadi 2001). In addition, the widespread presence of *Astragalus*, *Medicago*, *Onobrychis*, and *Trigonella* contributed to representation of the family.

Lamiaceae is the fourth most species rich family in our study and in the flora of Iran (Ghahreman and Attar 1999). Species of this family are calciphobic and excrete calcium (Eftekhari and Assadi, 2001; Eftekhari *et al.* 2002). The genera *Salvia*, *Phlomis*, and *Stachys* were the most common ones of this family.

A previous study in Turkey (Akpulat and Celik 2005) found that Liliaceae was the fourth most speciose family (25 taxa). No other study has recorded such high species richness for this family. They hypothesized that the commonness of *Allium* and *Iris* species in gypsum areas may be the main reason for that. Previous studies recorded only three Liliaceae species (*Allium bungei*, *Asparagus breslerianus*, and *Eremorus luteus*) in gypsum areas in Iran (Eftekhari and Assadi 2001). We found five Liliaceae species (3 species of *Allium*), while Azadbakht *et al.* (2007) has reported six Liliaceae species (2 species of *Allium*) in the adjacent non-gypsum area. Our literature review shows that 82 species of *Allium* exist in gypsum areas in Iran. This genus is one of the largest in the flora of Iran, which could explain its high representation in our study.

Eftekhari and Assadi (2001) and Eftekhari *et al.* (2002) found that Chenopodiaceae is the most abundant family (16 and 13 taxa, respectively). They suggested that Chenopodiaceae are successful in establishing and spreading in gypsum soils. We also found two Chenopodiaceae species, which disagrees with Eftekhari and Assadi (2001) and Eftekhari *et al.* (2002).

Astragalus has about 2,500 species in the world and is one of the largest genera of flowering plants. It is common in steppe areas, since excessive destruction provides new habitats for members of the genus. Therefore, these areas contains by far the greatest number of *Astragalus* species (Akpulat and Celik 2005). The estimated number of species of this genus for Iran is about 800 (Maassoumi 1986–2005). There are eight *Astragalus* species (*A. citrinus*, *A. fridae*, *A. semnanensis*, *A. iranicus*, *A. podolobus*, *A. kavirensis*, *A. verus*, and *A. reuterianus*) in gypsum areas

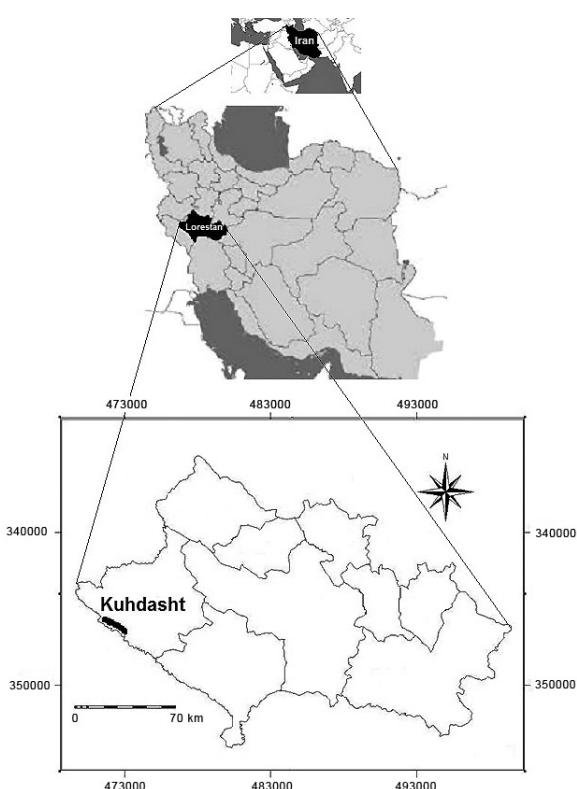


FIGURE 1. Location of the studied area (—) in Iran and Lorestan province.



TABLE 1. Taxa list in the studied area. IT: Irano-Turanian; M: Mediterranean; ES: Euro-Siberian; SS: Saharo-Sindian; Pl: polyregional; Cosm: Cosmopolitan; 1: Endemic; 2: Lower risk; 3: Vulnerable; Th: Therophyte; Hem: Hemicryptophyte; Ge: Geophyte; Ch: Chamaephyte; Ph: Phanerophyte.

TAXA		LIFE FORM	CHROTYPE	VOUCHER NO.
Magnoliophyta				
Magnoliopsida				
Amaranthaceae				
<i>Amaranthus blitoides</i> S. Watson var. <i>halophilus</i> Aell. ^{1,2}	Th	IT		2524
Aristolochiaceae				
<i>Aristolochia bottae</i> Jaub. & Spach	Hem	IT		2547
Boraginaceae				
<i>Anchusa italicica</i> Retz	Hem	IT-M-ES		2355
<i>Anchusa strigosa</i> Labill. subsp. <i>strigosa</i>	Hem	IT-M		2399
<i>Arnebia decumbens</i> (Vent.) Coss. & Karl subsp. <i>decumbens</i>	Th	IT-SS		2390
<i>Onosma bulbotrichum</i> DC.	Hem	IT		2385
<i>Onosma albo-roseum</i> Fisch. & C. A. Mey.	Ge	IT		2370
<i>Onosma trachytrichum</i> Boiss.	Hem	IT		2366
<i>Paracaryum rugulosum</i> (DC.) Boiss.	Hem	IT-SS		2377
Capparidaceae				
<i>Capparis spinosa</i> L.	Ch	IT-M-SS		2566
Caryophyllaceae				
<i>Buffonia oliveriana</i> Ser. in DC.	Th	IT		2310
<i>Gypsophila bicolor</i> (Freyn & Sint.) Grossh.	Hem	IT		2349
<i>Gypsophila elymaitica</i> Mozaff. ^{1,2}	Hem	IT		2345
<i>Gypsophila linearifolia</i> (Fisch. & C.A. Mey.) Boiss.	Th	IT		2333
<i>Gypsophila obconica</i> Barkoudah	Th	IT		2347
<i>Gypsophila pallida</i> Stapf var. <i>pallida</i>	Hem	IT		2326
<i>Gypsophila pilosa</i> Huds.	Th	IT		2330
<i>Silene conoidea</i> L.	Th	Pl		2339
<i>Vaccaria grandiflora</i> (Fisch. & DC.) Jaub. & Spach	Th	IT		2336
Chenopodiaceae				
<i>Noaea mucronata</i> (Forssk.) Aschers. et Schweinf.	Hem	IT-M		2599
<i>Salsola nitraria</i> Pall.	Th	IT-SS		2587
Cistaceae				
<i>Helianthemum salicifolium</i> (L.) Miller	Th	IT-M-SS		2623
Asteraceae				
<i>Achillea wilhelmsii</i> C. Koch	Hem	IT		2007
<i>Anthemis cotula</i> L.	Th	IT-ES-SS		2014
<i>Atractylis cancellata</i> L.	Th	IT-M		2025
<i>Carduus arabicus</i> Jacq. ex Murray	Th	IT-SS		2017
<i>Carthamus oxyacantha</i> M. B.	Th	IT		2020
<i>Centaurea bruguierana</i> (DC.) Hand.-Mzt.	Th	IT		2036
<i>Centaurea koeieana</i> Bornm.	Hem	IT		2049
<i>Centaurea solstitialis</i> L. subsp. <i>solstitialis</i>	Th	IT		2040
<i>Centaurea virgata</i> Lam. subsp. <i>squarossa</i> (Willd.) Gugler	Hem	IT		2024
<i>Chardinia orientalis</i> (L.) O. Kuntze	Th	IT		2047
<i>Crepis kotschyana</i> (Boiss.) Boiss.	Th	IT		2030
<i>Crepis sancta</i> (L.) Babcock	Th	IT		2022
<i>Crepis foetida</i> L. subsp. <i>foetida</i>	Th	IT-M-ES		2010
<i>Crupina crupinastrum</i> (Moris) Vis.	Th	IT-M		2016
<i>Cymbalaria griffithii</i> (A. Gray) Wagenitz	Th	IT		2021
<i>Echinops viscidulus</i> Mozaff.	Hem	IT		2003
<i>Filago vulgaris</i> Lam.	Th	IT-ES		2045
<i>Gundelia tournefortii</i> L.	Hem	IT		2029
<i>Koelpinia chrysoglochis</i> Rech. f.	Th	IT-SS		2038
<i>Onopordon leptolepis</i> DC.	Hem	IT		2043
<i>Outreya carduiformis</i> Jaub. & Spach	Hem	IT		2031
<i>Pentanema divaricatum</i> Cass.	Th	IT-SS		2001
<i>Picris strigosa</i> M. B. subsp. <i>kurdica</i> Lack	Hem	IT		2013
<i>Postia bombycina</i> Boiss. & Hausskn. ^{1,2}	Hem	SS		2018
<i>Postia puberula</i> Boiss. & Haussk. ^{1,2}	Hem	IT-SS		2034
<i>Scariola orientalis</i> (Boiss.) Sojak	Hem	IT		2019
<i>Scorzonera luristanica</i> Rech. f.	Hem	IT		2028

TABLE 1. CONTINUED.

TAXA	LIFE FORM	CHROTYPE	VOUCHER NO.
<i>Tanacetum polyccephalum</i> Schultz-Bip. subsp. <i>polyccephalum</i>	Hem	IT	2042
<i>Zoegea leptarea</i> L. subsp. <i>mianensis</i> (Boiss.) Rech. f. ^{1,2}	Th	IT	2048
Convolvulaceae			
<i>Convolvulus commutatus</i> Boiss.	Ch	IT	2645
<i>Convolvulus dorycnium</i> L.	Hem	IT	2637
<i>Convolvulus reticulatus</i> Choisy in DC.	Hem	IT	2649
Brassicaceae			
<i>Alyssum desertorum</i> Staph var. <i>desertorum</i>	Th	IT	2210
<i>Biscutella didyma</i> L.	Th	IT-M	2220
<i>Clypeola jonthlaspi</i> L.	Th	IT-M	2217
<i>Diplotaxis harra</i> (Forssk.) Boiss.	Th	IT-M-SS	2230
<i>Eruca sativa</i> Miller	Th	PI	2240
<i>Erucaria hispanica</i> (L.) Druce	Th	IT-M-SS	2249
<i>Isatis raphanifolia</i> Boiss. ^{1,2}	Th	IT	2223
<i>Malcolmia africana</i> (L.) R. Br.	Th	IT-M-SS	2246
<i>Matthiola longipetala</i> (Vent.) DC.	Th	IT-M-SS	2215
<i>Sinapis arvensis</i> L.	Th	PI	2219
<i>Thlaspi perfoliatum</i> L.	Th	IT-M-ES	2238
<i>Torularia torulosa</i> (Desf.) O.E. Schulz	Th	IT-SS	2244
Dipsacaceae			
<i>Cephalaria dichaetophora</i> Boiss.	Th	IT	2671
<i>Pterocephalus plumosus</i> (L.) Coult.	Th	IT-M	2665
<i>Scabiosa olivieri</i> Coulter	Th	IT	2669
<i>Scabiosa persica</i> Boiss.	Th	IT	2655
<i>Scabiosa sculpta</i> L.	Th	IT	2658
Euphorbiaceae			
<i>Andrachne telephiooides</i> L.	Hem	IT-M-SS	2686
<i>Chrozophora tinctoria</i> (L.) A. Juss.	Th	IT-M	2699
<i>Euphorbia crasspedia</i> Boiss. ^{1,2}	Hem	IT	2697
<i>Euphorbia helioscopia</i> L.	Th	PI	2677
<i>Euphorbia macroclada</i> Boiss.	Hem	IT	2687
<i>Euphorbia falcatia</i> L.	Th	IT-M	2688
Fagaceae			
<i>Quercus brantii</i> Lindl. var. <i>persica</i> (Jaub. & Spach) Zohary ^{1,2}	Ph	IT	2705
Geraniaceae			
<i>Erodium cicutarium</i> (L.) L'Her. ex Aiton	Th	IT-M-ES	2729
<i>Geranium rotundifolium</i> L.	Th	IT-M-ES	2748
Hypericaceae			
<i>Hypericum helianthemooides</i> (Spach) Boiss.	Hem	IT	2773
Lamiaceae			
<i>Phlomis bruguieri</i> Desf.	Hem	IT	2199
<i>Phlomis polioxantha</i> Rech. f.	Hem	IT	2157
<i>Phlomis olivieri</i> Benth.	Hem	IT	2190
<i>Salvia compressa</i> Vent.	Hem	IT	2173
<i>Salvia macrosiphon</i> Boiss.	Hem	IT	2179
<i>Salvia palaestina</i> Benth.	Hem	IT-M	2162
<i>Salvia reuterana</i> Boiss.	Hem	IT	2172
<i>Scutellaria pinnatifida</i> A. Hamilt. subsp. <i>pinnatifida</i>	Ch	IT	2152
<i>Stachys benthamiana</i> Boiss.	Hem	IT	2193
<i>Stachys inflata</i> Benth.	Hem	IT	2163
<i>Stachys pilifera</i> Benth. in DC. ^{1,2}	Hem	IT	2178
<i>Teucrium orientale</i> L.	Hem	IT	2166
<i>Teucrium polium</i> L.	Ch	IT-M	2187
<i>Ziziphora capitata</i> L. subsp. <i>orientalis</i> Samuelsson ex Rech. f.	Th	IT-M	2164
Linaceae			
<i>Linum mucronatum</i> Bertol. subsp. <i>mucronatum</i>	Ch	IT	2797
<i>Linum strictum</i> L. var. <i>spicatum</i> Pers.	Th	IT-M	2786
Malvaceae			
<i>Alcea kurdica</i> (Schlesht.) Alef	Hem	IT	2812

TABLE 1. CONTINUED.

TAXA	LIFE FORM	CHROTYPE	VOUCHER NO.
<i>Malva parviflora</i> L.	Th	IT-M-SS	2824
<i>Malva sylvestris</i> L. var. <i>sylvestris</i>	Hem	IT-M-ES	2820
<i>Malva verticillata</i> L.	Th	IT-SS	2815
Mimosaceae			
<i>Prosopis farcta</i> (Banks & Soland.) Macbr.	Ch	IT-M-SS	2846
Moraceae			
<i>Ficus carica</i> L. subsp. <i>rupestris</i> (Hausskn. ex Boiss.) Browincz	Ph	IT-M	2872
Orobanchaceae			
<i>Orobanche ramosa</i> L.	Th	Pl	2891
Papaveraceae			
<i>Chelidonium majus</i> L.	Th	Pl	2923
<i>Glaucium fimberilligerum</i> Boiss.	Th	IT	2914
<i>Glaucium grandiflorum</i> Boiss. & Huet.	Hem	IT	2910
<i>Papaver bracteatum</i> Lindl.	Hem	IT	2906
Fabaceae			
<i>Alhagi persarum</i> Boiss. & Buhse	Hem	IT	2110
<i>Astragalus crispocarpus</i> Nabelek ²	Th	IT	2102
<i>Astragalus ecbatanus</i> Bunge ^{1,2}	Ch	IT	2125
<i>Astragalus eriosphaerus</i> Boiss.	Ch	IT	2107
<i>Astragalus fasciculifolius</i> Boiss. subsp. <i>fasciculifolius</i> ^{1,2}	Ph	IT	2135
<i>Astragalus kerkukiensis</i> Bornm.	Th	IT	2145
<i>Glycyrrhiza glabra</i> L. ²	Hem	IT-M	2132
<i>Medicago orbicularis</i> (L.) Bartalini ²	Th	Pl	2139
<i>Medicago polymorpha</i> L.	Th	Cosm	2128
<i>Medicago radiata</i> L.	Th	IT-M	2146
<i>Medicago rigidula</i> (L.) All. ²	Th	IT-M-ES	2138
<i>Onobrychis caput-galli</i> (L.) Lam.	Th	IT-M	2111
<i>Onobrychis crista-galli</i> (L.) Lam.	Th	IT-M-SS	2119
<i>Onobrychis ptolemaica</i> (Del.) DC.	Hem	IT-SS	2130
<i>Trifolium campestre</i> Schreb.	Th	IT-M-ES	2122
<i>Trigonella elliptica</i> Boiss. ^{1,2}	Ch	IT	2149
<i>Trigonella monantha</i> C.A. Mey.	Th	IT	2108
<i>Trigonella persica</i> Boiss. ^{1,2}	Th	IT	2129
<i>Vicia peregrina</i> L. var. <i>peregrina</i>	Th	IT-M	2142
Plantaginaceae			
<i>Plantago lanceolata</i> L.	Hem	IT-ES-SS	2947
Polygalaceae			
<i>Polygala hehenackeriana</i> Fisch. & C.A. Mey.	Hem	IT	2965
Polygonaceae			
<i>Pteropyrum noenum</i> Boiss. & Meisner ³	Ph	IT-SS	2979
Primulaceae			
<i>Anagallis arvensis</i> L.	Th	IT-M-ES	2231
Ranunculaceae			
<i>Adonis aestivalis</i> L.	Th	IT-M-ES	2046
<i>Anemone biflora</i> DC.	Ge	IT	2067
<i>Delphinium cyphoplectrum</i> Boiss.	Hem	IT-SS	2089
<i>Thalictrum isopyroides</i> C.A. Mey.	Hem	IT	2094
Rosaceae			
<i>Amygdalus arabica</i> Olivier	Ph	IT	2401
<i>Amygdalus lycioides</i> Spach var. <i>lycioides</i>	Ph	IT	2410
<i>Crataegus meyeri</i> A. Pojark.	Ph	IT-ES	2417
<i>Crataegus monogyna</i> Jacq.	Ph	IT-ES	2412
<i>Rosa canina</i> Boiss.	Ph	IT	2432
<i>Sanguisorba minor</i> Scop.	Hem	IT	2422
Rubiaceae			
<i>Gaillonia bruguieri</i> A. Rich. ex DC. ^{1,2}	Hem	IT	2557
<i>Galium setaceum</i> L.	Th	IT-M	2977
<i>Callipeltis cucullaria</i> (L.) Stev.	Th	IT-SS	2435

TABLE 1. CONTINUED.

TAXA	LIFE FORM	CHROTYPE	VOUCHER NO.
Rutaceae			
<i>Haplophyllum tuberculatum</i> (Forssk.) Juss.	Hem	IT-SS	2298
Scrophulariaceae			
<i>Albraunia fugax</i> (Boiss. & Nöe) Speta	Th	IT	2174
<i>Scrophularia striata</i> Boiss.	Hem	IT	2195
Tamaricaceae			
<i>Tamarix tetragyna</i> Ehrenb. var. <i>meyeri</i> (Boiss.) Boiss.	Ph	IT-SS	2476
Thymelaeaceae			
<i>Thymelaea passerina</i> (L.) Cosson & Germ.	Th	IT-ES	2596
Apiaceae			
<i>Anisosciadium orientale</i> DC.	Th	IT-SS	2299
<i>Bifora testiculata</i> (L.) Spreng.	Th	IT-M	2295
<i>Bunium cylindricum</i> (Boiss. & Hohen.) Druce	Ge	IT	2261
<i>Ferulago macrocarpa</i> (Fenzl) Boiss.	Hem	IT	2290
<i>Lagoecia cuminoides</i> L. ²	Th	IT-SS	2273
<i>Pimpinella barbata</i> (DC.) Boiss.	Th	IT-SS	2280
<i>Pimpinella eriocarpa</i> Banks. & Soland.	Th	IT-SS	2055
<i>Torilis leptophylla</i> (L.) Reichenb.	Th	IT-M-ES	2085
<i>Turgenia latifolia</i> (L.) Hoffm.	Th	IT-M-ES	2070
Valerianaceae			
<i>Valerianella vesicaria</i> (L.) Moench	Th	IT-M	2794
Liliopsida			
Poaceae			
<i>Aegilops umbellulata</i> Zhuk.	Th	IT	2051
<i>Avena fatua</i> L. var. <i>fatua</i>	Th	Pl	2057
<i>Avena wiestii</i> Steud.	Th	IT-M-SS	2054
<i>Bromus danthoniae</i> Trin. var. <i>danthoniae</i>	Th	IT	2063
<i>Bromus tectorum</i> L. var. <i>tectorum</i>	Th	Pl	2053
<i>Crypsis schoenoides</i> (L.) Lam.	Th	IT-M-ES	2078
<i>Cynodon dactylon</i> (L.) Pers.	Ge	Cosm	2099
<i>Echinaria capitata</i> (L.) Desf.	Th	IT-M	2081
<i>Eremopoa persica</i> (Trin.) Roshev. subsp. <i>persica</i>	Th	IT-M	2088
<i>Eremopyrum bonaepartis</i> (Spreng.) Nevski var. <i>bonaepartis</i>	Th	IT	2065
<i>Heterantherium piliferum</i> (Banks & Soland.) Hochst	Th	IT	2087
<i>Hordeum bulbosum</i> L.	Ge	IT-M-SS	2077
<i>Hordeum leporinum</i> Link	Th	IT-M	2064
<i>Hordeum spontaneum</i> C. Koch	Th	IT-M	2091
<i>Lolium temulentum</i> L.	Th	Pl	2097
<i>Melica jacquemontii</i> Decne. ex Jacquem	Ge	IT	2069
<i>Phalaris minor</i> Retz.	Th	IT-M	2074
<i>Phragmites australis</i> (Cav.) Trin. ex Steud. subsp. <i>australis</i>	Ge	Cosm	2095
<i>Poa bulbosa</i> L.	Ge	IT-M-ES	2080
<i>Polypogon monspeliensis</i> (L.) Desf.	Th	Pl	2052
<i>Secale montanum</i> Guss.	Hem	IT-M	2050
<i>Setaria viridis</i> (L.) P. Beauv.	Th	Pl	2061
<i>Stipa barbata</i> Desf	Hem	IT	2092
<i>Taeniatherum crinitum</i> (Schreb.) Nevski	Th	IT-M	2086
Iridaceae			
<i>Gynandriris sisyrinchium</i> (L.) Parl.	Ge	IT-SS	2134
<i>Iris hymenophytha</i> Mathew & Wendelbo ²	Ge	IT	2592
Liliaceae			
<i>Allium longisepalum</i> Bertol.	Ge	IT	2499
<i>Allium jesdianum</i> Boiss. & Buhse	Ge	IT	2467
<i>Allium rotundum</i> L.	Ge	IT-ES	2475
<i>Muscaria tenuiflorum</i> Tausch	Ge	IT-ES	2477
<i>Ornithogalum cuspidatum</i> Bertol.	Ge	IT	2456

in Iran (Eftekhari and Assadi 2001; Eftekhari *et al.* 2002), but only two (*A. arpilobus* and *A. oxyglottis*) in gypsum areas in China (Langran and Podlech 2010). In addition, only three *Astragalus* species of the section *Hololeuce* Bunge (*A. bicolor* subsp. *karputanus*, *A. andrasovszkyi* and *A. stenosemius*) were reported in gypsum areas in Turkey (Eküçü and Eküm 2004). We found five *Astragalus* species, while there were 25 *Astragalus* species in adjacent non-gypsum areas. This genus is not halophytic and does not grow in salty areas (Akhani and Ghorbanli 1993). Our results show that *Astragalus*, with about 800 species in Iran, does not occur in gypsum areas. This result is in conflict with Akpulat and Celik (2005).

Phytogeographic regions

The proportions of taxa in each phytogeographic regions are as follows: 92 species from IT (48.43%), 1 species from SS (0.53%), 6 species from IT-ES (3.16%), 20 species from IT-SS (10.53%), 28 species from IT-M (14.74%), 14 species from IT-M-ES (7.37%), 12 species from IT-M-SS (6.31%), 2 species from IT-ES-SS (1.05%), 12 species from multi-regional (6.31%), 3 species from cosmopolitan (1.57%) (Tables 1–2). These results are in accordance with our expectation, since the study area is in the IT region (see also Eftekhari and Assadi, 2001; Eftekhari *et al.*, 2002). IT-M elements are present since the study area is open and steppe. Elements from two or more regions comprise more than half of the species, due to the low altitude (Noroozi *et al.* 2008).

We found 14 taxa (7.36%) endemic to Iran. However, *Gypsophila elymaitica* Mozaff. is obligatory gypsophyte. The other 13 taxa are preferential gypsophyte or indifferent plants. Endemism rate is lower than the average for Iran (23.91%), since the region is in lowland (Noroozi *et al.* 2008). This result is in disagreement with Akpulat and Celik (2005), who suggested that the reason for the high endemism in Turkey is the gypsum habitats. For example, *Astragalus aytatchii* only grows in deep soil, gypsum fields, and slopes in Turkey (Akan and Civelek 2001). In arid and semi-arid environments with less than 400 mm average annual rainfall, gypsum crystals sediment on upper surface of gypsiferous soils by evaporation (Pueyo and Alados 2007).

We found that 21 taxa were threatened (20 taxa LR and 1 taxon VU). We suggest that three taxa: *Euphorbia craspedia* Boiss., *Quercus brantii* Lindl. var. *persica* (Jaub. & Spach) Zohary, and *Gypsophila elymaitica* Mozaff., whose conservation status is unknown, must be assigned LR status in the study area, according to IUCN (2010). *Astragalus kerkukiensis*, previously a Data Deficient (DD) species (Jalili and Jamzad 1999) was very abundantly in the study area.

Life forms

Therophytes, hemicryptophytes, geophytes, phanerophytes, and chamaephytes included 51.58, 30.53, 7.89,

5.26, and 4.74% of the total species, respectively. Ninety-eight species were annuals and 73 perennials. From all species, 19 were woody plants.

The environment of the studied area, with arid and cold climate, favors therophytes and hemicryptophytes. The active growth periods of therophytes and hemicryptophytes are concurrent with the rainy season in late winter and early spring (Tavili *et al.* 2009). During most of the summer and all winter, hemicryptophytes loose their aboveground parts while therophytes remain as seed. Chamaephytes and phanerophytes only occur when gypsiferous soils are not open. Altitudinal variation can also be a determinant factor for the relative abundance and geographic distribution of different life forms, along with climate (Noroozi *et al.* 2008).

Hemicryptophytes were the most abundant life form in the studies of Eftekhari and Assadi (2001) and Eftekhari *et al.* (2002; 49.1% and 52.9%, respectively). They suggested that hemicryptophytes could adapt to harsh environmental conditions for growth and establishment, such as in gypsum habitats. Therophytes were the most abundant life form (51.58%) in the studied area. These plants avoid stresses from the summer drought and winter cold (Barbour *et al.* 1987). Moreover, the high proportion of therophytes could also attributed to human activities (Barbero *et al.*, 1990).

TABLE 2. Floristic properties of the studied area. IT: Irano-Turanian; M: Mediterranean; ES: Euro-Siberian; SS: Saharo-Sindian; Pl: polyregional; Cosm: Cosmopolitan; LR: Lower risk; VU: Vulnerable; Th: Therophyte; Hem: Hemicryptophyte; Ge: Geophyte; Ch: Chamaephyte; Ph: Phanerophyte.

	Dicotyledons	Monocotyledons	Total
Families	36	3	39
Genera	112	25	137
Species	159	31	190
Subspecies	14	4	18
Varieties	9	4	13
Endemic taxa	14	0	14
IT	82	10	92
SS	1	0	1
IT-ES	4	2	6
IT-SS	19	1	20
IT-M	21	7	28
IT-M-ES	12	2	14
IT-M-SS	10	2	12
IT-ES-SS	2	0	2
Pl	7	5	12
Cosm	1	2	3
LR	19	1	20
VU	1	0	1
Ph	10	0	10
Ch	9	0	9
Hem	56	2	58
Ge	3	12	15
Th	82	17	98

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