

# Detection and development of a quantitation method for undeclared compounds in antidiabetic biologically active additives and its validation by high performance liquid chromatography

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## Abstract

An isocratic, high-performance liquid chromatography (HPLC) quantitation method was developed for the quantitative determination of metformin, glibenclamide, gliclazide, glimepiride in some antidiabetic biologically active additives. A Nucleosil C<sub>18</sub>, 5 μm, 4.6 mm × 150 mm, column with mobile phase containing buffer (10 mm Na<sub>2</sub>HPO<sub>4</sub>, 10 mm sodium dodecyl sulfate): acetonitrile = 68 : 32 (V/V), pH = 7.5 was used. The flow rate was 1.0 mL/min, and effluents were monitored at 226 nm. The retention times of gliclazide, glibenclamide, glimepiride and metformin, were 2.203, 4.587, 5.667 and 10.182 min, respectively. Linearity was studied by preparing standard solutions of gliclazide, glibenclamide, glimepiride and metformin at the concentration range of 50% to 150% of working concentration from a stock solution. The method was successfully applied to the estimation of gliclazide, glibenclamide, glimepiride and metformin in some antidiabetic biologically active additives. This method was validated to confirm its system suitability, selectivity, linearity, precision and accuracy according to international conference on harmonization (ICH) guidelines.

## Keywords

HPLC, biologically active additives, metformin, glibenclamide, gliclazide, glimepiride

## Introduction

Biologically active additives (BAA) contain bioactive elements - vitamins, minerals, proteins, are often supplemented with organ tissues, plants, etc., which in contrast to

drugs are in less quantities than therapeutic dosages and affect the body within physiological norms, are intended to strengthen the health, the body's resistance to pathogenic factors and to improve the quality of life (Gabrielyan et al. 2002; Zemtsova et al. 2020; Makhmudov et al. 2021).

There is no reasonable guarantee that BAA are absolutely safe, as there are many cases when undeclared substances such as hormones, analgesics, antidiabetic and other drugs are added to BAA to stimulate their effect, the long-term uncontrolled use of which is extremely hazardous (Peng et al. 2013; Mihaylova et al. 2020).

The correct selection of the method of analysis is of great importance for the detection of undeclared chemicals in BAA. The selected method should have high sensitivity, the ability to work with small quantities of samples, high selectivity, be distinguished by the rate of expertise, the simplicity of sample preparation, ease of maintenance of analytical equipment, reliability and reproducibility of the method, universality, automation of the analytical process. In practice, the most widely used method (95% of studies) is the high performance liquid chromatography (HPLC) with different detection techniques (Watson 2012; Kloos et al. 2014).

High performance liquid chromatography is a high precision physical method that meets the modern requirements of drug quality control and is used for the separation, quantitative and qualitative identification of compounds. In this study an available reversed-phase HPLC method for identification of metformin, glibenclamide, gliclazide, glimepiride with UV detection and isocratic elution mode was developed, evaluation of the method applicability and determination of validation indicators were carried out (Attimarad et al. 2011; Monzón et al. 2016; Mahmoud et al. 2019) It has been shown that the developed method meets the current international requirements. For the detection of metformin, glibenclamide, gliclazide, glimepiride, the developed method has been used in some BAA (“Dialevel”, “Sugar Balance”, “Blood Sugar”, “Karela Capsules”).

Taking into account that quite large quantities of BAA are consumed by the population in the Republic of Armenia and there is no permanent control over it, the following tasks have been set forward.

- Development of a detection method for undeclared chemicals in antidiabetic BAA and its validation,
- Confirmation of the applicability of the selected method for daily use in the laboratory,
- Implementation of research on some of the most common antidiabetic BAA in the Republic of Armenia (“Dialevel”, “Sugar Balance”, “Blood Sugar”, “Karela Capsules”) by the newly developed method.

## Materials and methods

All measurements were made with a “Shimadzu LC-20-MS” (Japan) equipped with an automatic injection system (SIL-20A), a detector (SPD-M20A), a chromatographic column (Nucleosil C18, 5 m, 250 × 4.6 mm), and a column thermostat (Shimadzu). Analytical balance (Shimadzu), deionized water system (Purelab, ELGA), “Vortex” core stirrer (Stuart, BioCote, UK), 0.45 m membrane filters (E-chrom Tech, Taiwan), glass volumetric flasks, measuring cylinders, cups, and pipettes of various capacities (Normax, Portugal) were used for the samples preparation.

For the mobile phase methanol (HPLC Grade, AppliChem), disodium hydrogen phosphate dihydrate (Na<sub>2</sub>HPO<sub>4</sub>·2H<sub>2</sub>O HPLC Grade, Sigma Aldrich), sodium dodecyl sulfate (HPLC Grade, ≥ 99%, Carl Roth), orthophosphoric acid (HPLC Grade, 85%, Carl Roth) were used.

Dialevel (Walmart, Czech Republic), Sugar Balance (Velt Farma, Germany), Blood Sugar (Nature’s Way, Australia), and Karela Capsules (Himalaya, India) were chosen as research samples.

As a standards metformin hydrochloride (BN-M0605000; 99.81%; E.Ph. RS), glibenclamide (BN-G0325000; 99.9%; E.Ph. RS), gliclazide (BN-M0605000, 99.81%, E.Ph. RS), glimepiride (BN-Y0000515, 99.46%, E.Ph. RS) were used and 10 mm Na<sub>2</sub>HPO<sub>4</sub>, 10 mm SDS (sodium dodecyl sulfate) have been used as buffer.

The chromatographic conditions are presented in Table 1.

Standard preparation: Methanol was used as a solvent, and for the preparation of the standard solutions (Table 2)

**Table 1.** Chromatographic conditions for the expertise of BAA method used in diabetes.

Chromatographic column	Nucleosil C <sub>18</sub> , 5 μm, 4.6 mm × 150 mm, (Macherey-Nagal, Germany) Nucleosil C <sub>18</sub> , 5 μm, 4.6 mm × 150 mm, (Macherey-Nagal, Germany)
Detector wavelength	226 nm
Flow rate	1 ml/min
Injection volume	20 μL
Column temperature	35 °C
Pump operating mode	Isocratic
Mobile phase	Buffer: acetonitrile = 68 : 32 (V/V), pH = 7.5

the corresponding weights of the standards were dissolved in 25 ml of solvent, after which they were placed in an ultrasonic bath for 10 minutes. The resulting solutions were then stirred with a VORTEX core stirrer for 5 minutes.

**Table 2.** Preparation of standard solutions.

Sample	STD-1	STD-2	STD-3	STD-4
	Metformin (99.81%)	Glibenclamide (99.90%)	Glimepiride (99.46%)	Gliclazide (99.80%)
Standard sample quantity (mg)	5.1	5.2	5.0	5.1
	5.0	5.1	5.0	5.1
	5.0	5.2	5.1	5.0

Mobile phase preparation: After mixing the organic and inorganic components of the mobile phase, the pH of the solution has been adjusted to 7.5 with orthophosphoric acid, filtered with a 0.45 μm membrane, again filtered and degasified.

Three injection were made from each solution.

**Preparation of standard mother solution (STD-5).** 1 ml of each of the above solutions was transferred to a 10 ml volumetric flask and adjusted to the mark in a mobile phase. The theoretical concentration of standards in this solution is presented in Table 3.

**Preparation of solutions for the construction of the calibration curve.** A mixture of 5 standards of different

**Table 3.** Concentration of standards in standard mother solution.

C (mg/ml)	Metformin	Glibenclamide	Glimepiride	Gliclazide
Concentration of the resulting solution (mg/ml)	0.020362	0.020779	0.019892	0.020359

concentrations was selected. For each preparation, the volumes shown in Table 4 were taken from STD-1, STD-2, STD-3, STD-3, STD-4 solutions and diluted in a mobile phase up to 20 ml.

**Preparation of solutions for interlaboratory accuracy and precision.** QCL (calib.1- 0.01 mg/ml), QCM (calib.3 - 0.02 mg/ml) and QCH (calib.5 - 0.03 mg/ml) solutions were prepared. Three injections were made from each solution.

**Table 4.** Preparation of calibration solutions.

Primary solutions Standard solutions (mg/ml)	Calib. 1 1 ml	Calib. 2 1.5 ml	Calib. 3 3 2 ml (STD-5)	Calib. 4 2.5 ml	Calib. 5 3 ml
STD-1 Metformin	0.010181	0.015272	0.020362	0.025453	0.030543
STD-2 Glibenclamide	0.0103895	0.015584	0.020779	0.025974	0.0311685
STD-3 Glimepiride	0.009946	0.014919	0.019892	0.024865	0.0298380
STD-4 Gliclazide	0.0101795	0.0152693	0.020359	0.025449	0.0305385

## Results and discussion

Validation of the method for quantitation of gliclazide, glibenclamide, glimepiride and metformin in antidiabetic biologically active additives was performed by evaluating the following indicators:

- Selectivity
- Accuracy
- Precision
- Linearity range

The most common representatives of chemical origin of this group of drugs are metformin from biguanides and glibenclamide, gliclazide, glimepiride derivatives of sulfonylurea.

The pharmacopoeia does not specify test methods for the associations of these substances. So, such a method should be selected that is universal for detecting substances from these two different chemical groups.

**System suitability test of the chromatographic system with the expertise BAA method used in diabetes.** It is to be implemented to perceive whether the selected method for the simultaneous determination of metformin and glibenclamide can be used for the simultaneous detection of other representatives of this series.

The retention times of the substances under the selected chromatographic conditions are shown in Figure 1.

### Acceptance criteria:

1. the tailing factor of the peak due to gliclazide, glibenclamide, glimepiride and metformin is not more than 2.0,
2. the number of theoretical plates of the peak due to gliclazide, glibenclamide, glimepiride and metformin is more than 2500,
3. resolution between peaks to gliclazide, glibenclamide, glimepiride and metformin is not less than 2.0. The results are presented in Table 5.

**Table 5.** System suitability test of the chromatographic system.

SST	Resolution (Distribution between peaks)	Tailing Factor	Theoretical Plate
Gliclazide	-	1.32	2500
Glibenclamide	10.4	1.00	4300
Glimepiride	3.6	0.97	4900
Metformin	8.9	1.67	3400

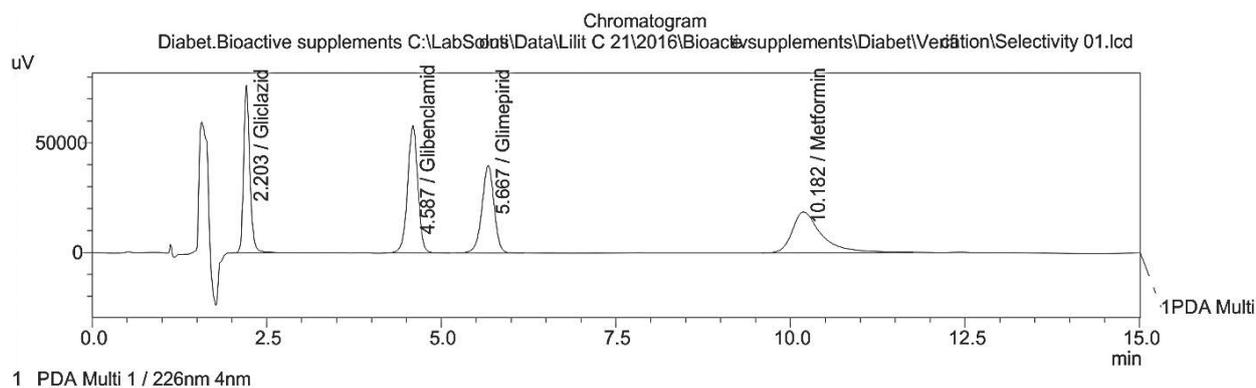
Selectivity is the ability to access unequivocally the analyte in presence of components, which may be expected to be present. To prove selectivity, the following experiment is carried out. Selectivity was tested on blank, active ingredient and finished product.

**Acceptance Criteria:** no interference of the blank at the retention time of gliclazide, glibenclamide, glimepiride and metformin. The results are presented in the Figure 2 and Table 6.

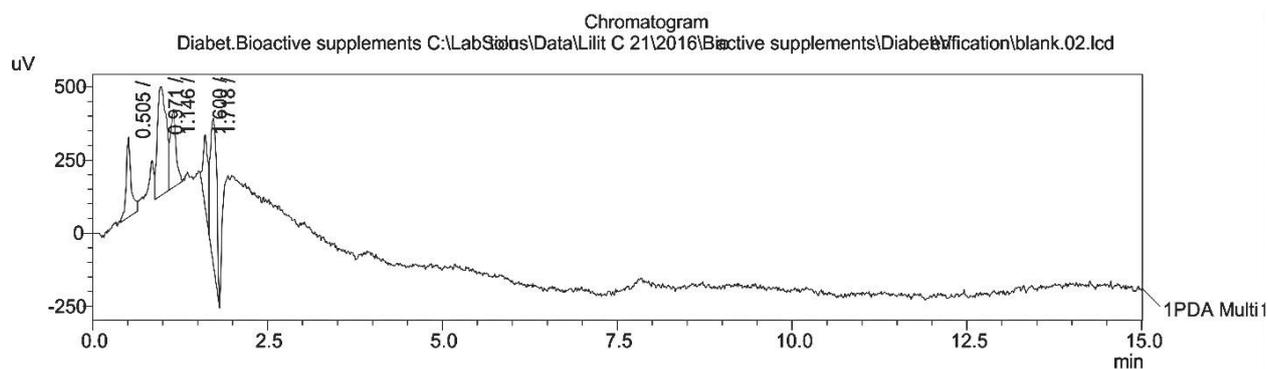
Linearity of an analytical procedure is its ability to obtain test results which are directly proportional to the concentration of analyte in the sample. Linearity was studied by preparing standard solutions of gliclazide, glibenclamide, glimepiride and metformin at the concentration range of 50% to 150% of working concentration from a stock solution and each concentration was injected in triplicate and chromatographed as per procedure. The results are presented in Tables 7 and 8 and Figure 3.

**Conclusion:** a method is linear in the range from 50 percent to 150 percent of gliclazide, glibenclamide, glimepiride and metformin concentration in standard solution chromatogram.

**Acceptance Criteria:** correlation coefficient is not less than 0.995.



**Figure 1.** Chromatogram of a standard mixture.



1 PDA Multi 1 / 226nm 4nm

## Quantitative Results

PDA								
ID#	Name	Ret. Time	Conc.	Area	Height	Channel	Theoretical Plate#	
1	Gliclazid	0.000	0.000000	0	0	Ch1	0.000	
2	Glibenclamid	0.000	0.000000	0	0	Ch1	0.000	
3	Glimepirid	0.000	0.000000	0	0	Ch1	0.000	
4	Metformin	0.000	0.000000	0	0	Ch1	0.000	

Figure 2. Method selectivity.

Table 6. Evaluation results of the indicator of "Method selectivity".

Sample code	Gliclazide		Glibenclamide		Glimepiride		Metformin	
	Retention time (min)	Surface of stress point	Retention time (min)	Surface of stress point	Retention time (min)	Surface of stress point	Retention time (min)	Surface of stress point
Sel.-1	2.203	466511	4.587	592230	5.667	481690	10.182	541124
Sel.-2	2.198	468454	4.575	591999	5.651	481894	10.190	542061
Sel.-3	2.206	468542	4.577	592152	5.648	482539	10.205	545546
Sel.-4	2.200	468974	4.563	592244	5.631	482160	10.206	543075
Sel.-5	2.194	468743	4.550	592677	5.614	482381	10.209	542536
Sel.-6	2.189	467345	4.538	592617	5.599	482241	10.213	544059
Average value	2.19833	468094.8	4.565	592319.8	5.635	482150.8	10.20083	543066.8
SD	0.00562	874.9086	0.01676	245.2206	0.023101	285.8306	0.011037	1425.881
RSD, %	0.25553	0.186908	0.36721	0.041400	0.40996	0.059282	0.108193	0.262561

Table 7. Linearity range: Evaluation results of the indicator of "Linearity range" for gliclazide and glibenclamide.

Calibration curve code	Calibration curve indicators			
	Gliclazide		Glibenclamide	
	Concentration (mg/ml)	Surface	Concentration (mg/ml)	Surface
Calib.1	0.0101795	240666	0.0103895	295663
Calib.2	0.0152693	359533	0.015584	445067
Calib.3	0.020359	468247	0.020779	592182
Calib.4	0.025449	575351	0.025974	741294
Calib.5	0.0305385	686518	0.0311685	889448
Correlation coefficient R (NLD 0.995)	0.9998101		0.9999979	

Table 8. "Linearity range" results for glimepiride and metformin.

Calibration curve code	Calibration curve indicators			
	Glimepiride		Metformin	
	Concentration (mg/ml)	Surface	Concentration (mg/ml)	Surface
Calib.1	0.009946	240467	0.010181	265471
Calib.2	0.014919	362123	0.015272	406362
Calib.3	0.019892	482007	0.020362	543058
Calib.4	0.024865	603238	0.025453	687018
Calib.5	0.0298380	723869	0.030543	823807
Correlation coefficient R (NLD 0.995)	0.9999982		0.9999730	

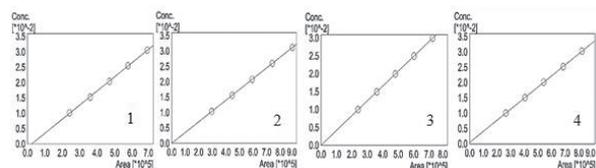


Figure 3. Calibration curves of 1-Gliclazide, 2-Glibenclamide, 3-Glimepiride and 4-Metformin.

**Acceptance Criteria:** the mean recovery should be in the range of 98.0 percent to 102.0 percent. RSD should be less than 2 percent. The results are presented in Table 9.

**Acceptance Criteria:** difference between the RSD results of Day 1 & Day 2 is not more than 2.0%. The results are presented in Tables 10 and 11.

Similar results were obtained with glibenclamide and glimepiride.

Table 9. Accuracy: Evaluation results of the indicator of "Accuracy".

Sample code	Gliclazide	Glibenclamide	Metformin
	0.020359 mg/ml	0.020779 mg/ml	0.020362 mg/ml
Sel.-1	0.020411	0.020782	0.020055
Sel.-2	0.020500	0.020774	0.020089
Sel.-3	0.020504	0.020780	0.020215
Sel.-4	0.020524	0.020783	0.020125
Sel.-5	0.020513	0.020798	0.020106
Sel.-6	0.020449	0.020796	0.020161
Average value	0.0204835	0.0207855	0.020125167
SD	4.0103E-05	8.63616E-06	5.15636E-05
RSD, %	0.195781934	0.041548985	0.256214558
Average recovery, %	100.61	100.03	98.84

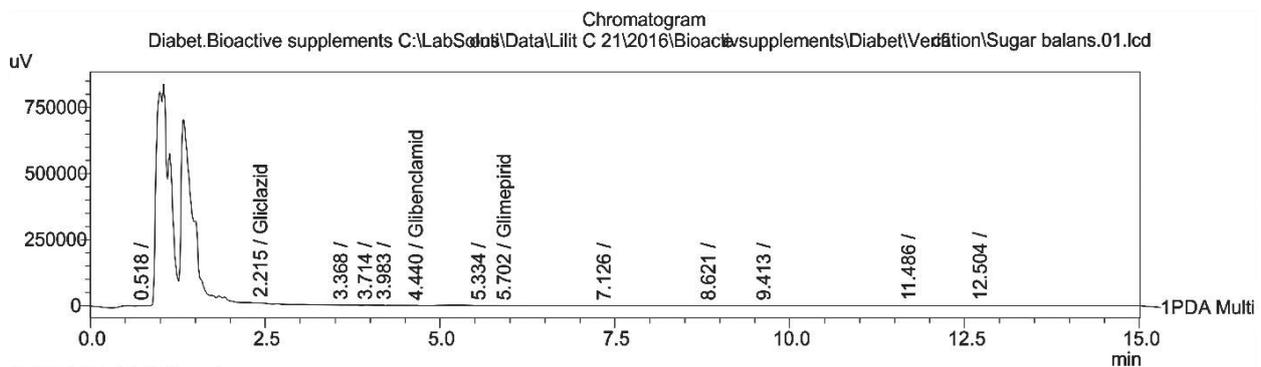
In large pharmacy chains surveys were conducted for the detection of the most common BAA in these groups, which resulted in the selection of "Dialevel", "Sugar Balance", "Kerala Capsules", "Blood Sugar" additives used in diabetes.

**Table 10.** Interlab Precision: Evaluation results of the indicator of “Interlab Precision” for gliclazide.

Gliclazide	Day 1			Day 2		
	QCL 0.01 mg/ml	QCM 0.02 mg/ml	QCH 0.03 mg/ml	QCL 0.01 mg/ml	QCM 0.02 mg/ml	QCH 0.03 mg/ml
1	0.010208	0.020646	0.028036	0.010222	0.020654	0.028052
2	0.010140	0.020592	0.028192	0.010151	0.020589	0.028181
3	0.010095	0.020614	0.028219	0.010099	0.020626	0.028223
Average value	0.0102347	0.02061733	0.028149	0.0101573	0.020623	0.028152
SD	0.0001263	2.2171E-05	8.0659E-05	5.0414E-05	2.66208E-05	7.27599E-05
RSD, % <sup>2</sup>	1.23442271	0.10753598	0.28654581	0.4963295	0.129083033	0.258453677

**Table 11.** Evaluation results of the indicator of “Interlab Precision” for metformin.

Metformin	Day 1			Day 2		
	QCL 0.01 mg/ml	QCM 0.02 mg/ml	QCH 0.03 mg/ml	QCL 0.01 mg/ml	QCM 0.02 mg/ml	QCH 0.03 mg/ml
1	0.009987	0.020314	0.030389	0.009959	0.020323	0.030376
2	0.010072	0.020218	0.030399	0.010061	0.020211	0.030382
3	0.010159	0.020193	0.030246	0.010145	0.020186	0.030239
Average value	0.0100727	0.02024167	0.03034467	0.010055	0.02024	0.0303323
SD	7.0221E-05	5.2156E-05	6.9887E-05	7.6053E-05	5.95707E-05	6.60421E-05
RSD, %	0.69713702	0.25766529	0.2303114	0.7563662	0.294321573	0.217728301

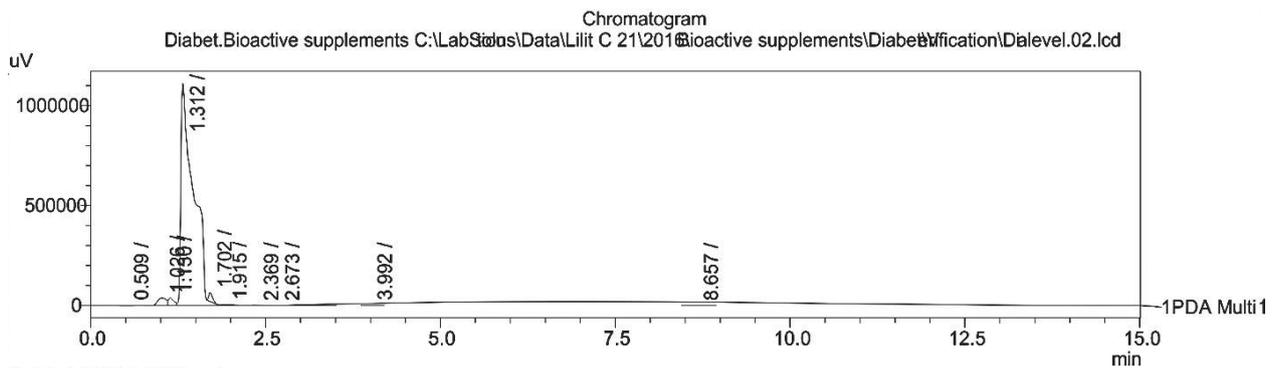


1 PDA Multi 1 / 226nm 4nm

Quantitative Results

PDA ID#	Name	Ret. Time	Conc.	Area	Height	Channel	Theoretical Plate#
1	Gliclazid	2.215	-0.000294	16725	2893	Ch1	2626.805
2	Glibenclamid	4.440	0.000142	3256	199	Ch1	2421.371
3	Glimepirid	5.702	0.000159	3001	254	Ch1	4627.354
4	Metformin	0.000	0.000000	0	0	Ch1	0.000

**Figure 4.** “Dialevel” sample injection chromatogram.



1 PDA Multi 1 / 226nm 4nm

Quantitative Results

PDA ID#	Name	Ret. Time	Conc.	Area	Height	Channel	Theoretical Plate#
1	Gliclazid	0.000	0.000000	0	0	Ch1	0.000
2	Glibenclamid	0.000	0.000000	0	0	Ch1	0.000
3	Glimepirid	0.000	0.000000	0	0	Ch1	0.000
4	Metformin	0.000	0.000000	0	0	Ch1	0.000

**Figure 5.** “Sugar Balance” sample injection chromatogram.

As shown in Figure 4 and 5, no undeclared substance in additives “Dialevel” and “Shugar Balance” were detected by automatic registration system as a result of expertise. Similar results were obtained during the expertise of BAA in “Karela Capsules” and “Blood Sugar”.

## Conclusion

A method for detecting undeclared chemicals in antidiabetic BAA has been developed and introduced by us. The

developed new method was used to study some of the most common antidiabetic BAA in the Republic of Armenia (Dialevel, Sugar Balance, Blood Sugar, Karela Capsules). The applicability of the selected method for daily use in the laboratory has been confirmed.

## Conflict of interest

The authors declare no conflict of interest.

## Author's contribution

H. Petrosyan participated in the preparation, writing and editing of the manuscript. V. Kirakosyan, E Minasyan performed quantitative determinations using Shimadzu

LC-20-MS, as well as processing and interpretation of the manuscript. L. Poghosyan took part in obtaining test samples, conducting experiments, as well as processing and interpreting data. L. Yu. Sahakyan, participated in the design and implementation of experiments, as well as in the processing and interpretation of data. A. Tsaturyan, T. Sargsyan participated in the design and implementation of experiments, as well as in the processing and interpretation of data.

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