







## Research Article

# Ants in the clouds: A preliminary checklist of the ant (Hymenoptera, Formicidae) fauna of a Honduran cloud forest ecosystem, featuring a key to country genera

Frederik C. De Wint<sup>1,2,3</sup>, Dominik Oorts<sup>1</sup>, Michael G. Branstetter<sup>4</sup>, Dario De Graaf<sup>1</sup>, Wouter Dekoninck<sup>5</sup>, Merlijn Jocque<sup>1</sup>, Thomas E. Martin<sup>6,7</sup>, Jennifer Sudworth<sup>1</sup>, Ronja Van Osselaer<sup>1</sup>, Matthew T. Hamer<sup>1,8</sup>

<sup>1</sup> Biodiversity Inventory for Conservation npo (BINCO), Walmersumstraat 44, 3380 Glabbeek, Belgium

<sup>2</sup> Department of Zoology, Faculty of Science, University of South Bohemia, Branisovska 1160/31, 37005 České Budějovice, Czech Republic

<sup>3</sup> Biology Centre of the Czech Academy of Sciences, Institute of Entomology, Department of Ecology, Branisovska 1160/31, 37005 Ceske Budejovice, Czech Republic

<sup>4</sup> U.S. Department of Agriculture, Agricultural Research Service, Pollinating Insects Research Unit, Utah State University, Logan, UT, USA

<sup>5</sup> Entomology Department, Royal Belgian Institute of Natural Sciences, Vautierstraat 29, B-1000 Brussels, Belgium

<sup>6</sup> School of Natural Sciences, College of Environmental Sciences and Engineering, Bangor University, Bangor, Gwynedd, LL57 2DG, UK

<sup>7</sup> Operation Wallacea Ltd, Wallace House, Old Bolingbroke, Lincolnshire PE23 4EX, UK

<sup>8</sup> School of Biological Sciences, The University of Hong Kong, Kadoorie Biological Sciences Building, Pok Fu Lam Road, Hong Kong SAR, China

Corresponding author: Frederik C. De Wint ([frederikdwint@hotmail.com](mailto:frederikdwint@hotmail.com))



Academic editor: José Monzón Sierra

Received: 29 January 2024

Accepted: 30 April 2024

Published: 28 June 2024

ZooBank: <https://zoobank.org/3507972A-4260-44E9-BE7A-935D3826F524>

**Citation:** De Wint FC, Oorts D, Branstetter MG, de Graaf D, Dekoninck W, Jocque M, Martin TE, Sudworth J, Van Osselaer R, Hamer MT (2024) Ants in the clouds: A preliminary checklist of the ant (Hymenoptera, Formicidae) fauna of a Honduran cloud forest ecosystem, featuring a key to country genera. In: Lipińska M, Lopez-Selva MM, Sierra JM (Eds) Biodiversity research in Central America. Neotropical Biology and Conservation 19(2): 157–185. <https://doi.org/10.3897/neotropical.19.e119775>

## Abstract

Ant diversity in tropical montane rainforests is globally understudied. This is true for Cusuco National Park (CNP), a cloud forest ecosystem in northwestern Honduras that supports geographically isolated and threatened habitats. The current study presents the first comprehensive ant species checklist for CNP, which is also the first ant checklist for Honduras in over a century. Species records from several projects are also combined and presented. Sampling occurred along an elevational range (mainly between 1170 and 2030 m a.s.l.), with methodologies and intensities varying among projects and dates. Overall, 162 ant species belonging to nine subfamilies and 60 genera are reported from the CNP. Five species are recorded for the first time in Honduras (*Pheidole natalie* Longino, 2019; *Strumigenys* cf. *calamita*; *Solenopsis invicta* Buren, 1972; *Solenopsis texana/carolinensis*; *Pseudomyrmex pallens* Mayr, 1870). For the first time, male individuals are reported in *Pheidole balatro* Longino, 2019. For each species, we provide information on observed habitat preference, elevational range, and sampling technique. Species accumulation curves are provided for each sample technique, representing sampling intensity and community sample coverage. We also provide a key to the ant genera of Honduras to aid future taxonomic efforts in the country. Our research demonstrates that CNP harbours a surprisingly rich diversity of ant species, despite its small area, similar to many other taxa in the park. The information provided here represents baseline information for future work on ants in CNP and other Honduran cloud forests and will help guide research in these otherwise poorly explored yet highly threatened ecosystems.

**Key words:** Biodiversity hotspot, Cusuco National Park, insects, Mesoamerica, species diversity, tropical montane forest

Copyright: © Frederik C. De Wint et al.

This is an open access article distributed under terms of the Creative Commons Attribution License (Attribution 4.0 International – CC BY 4.0).

## Introduction

Tropical montane cloud forests are located in the humid tropics within the maximum cloud condensation zone (Ellenberg 1959). These forests are markedly different from those found at lower elevations, creating biogeographical isolation and harbouring abundant endemic flora and fauna as a result (Long 1995; Anderson and Ashe 2000; Bubb et al. 2004; Martin et al. 2021). For instance, because of their precipitation patterns, many cloud forests show high abundances of epiphytic plant growth, which provides unique niches and microhabitats for other species (Stadtmüller 1986). Cloud forests are understudied globally, particularly in terms of insect fauna, with baseline inventories lacking for many sites (Jones et al. 2008; Sabu et al. 2011). Habitat loss and climate change significantly threaten these unique ecosystems and the species they support (Freeman et al. 2018; Hansen et al. 2020).

Cusuco National Park (CNP hereafter), situated within the Merendon mountain range in northwest Honduras, is one such tropical montane cloud forest. Located in the Mesoamerican biological hotspot (Myers et al. 2000), CNP has been designated as one of the 137 most irreplaceable protected areas in the world (Le Saout et al. 2013). Despite this, the park is under severe threat from deforestation and subsequent land conversion for subsistence agriculture (Martin et al. 2021). Honduras as a whole is one of the most severely impacted countries in terms of deforestation within protected areas (Hansen et al. 2020). Biodiversity in the park is therefore under significant anthropogenic pressure, particularly for regionally endemic species, of which the park harbours a high number across many floral and faunal groups (Martin et al. 2021). This forest is known for harbouring understudied taxa, with ongoing discoveries of multiple novel species, particularly within the arthropod class. (Mendes et al. 2011; Pinto and Jocque 2013; Damron et al. 2018; Santos-Silva et al. 2018, 2021; Longino 2019; Jocque and Garrison 2022).

The ecological impact of ants on most communities is hard to overstate. Ants (Formicidae) are ecologically dominant and ubiquitous in nearly all habitats across the globe. They are key components of many ecosystems, influencing communities as predators, seed dispersers (myrmecochory), and direct and indirect herbivory (Hölldobler and Wilson 1990; Del Toro et al. 2012). These socially organised insects are often closely associated with a variety of organisms, ranging from plants to arthropods (Hölldobler and Wilson 1990). Arthropod community patterns are significantly shaped by ants across montane landscapes (Rudgers et al. 2010) and even increase plant growth (Moreira et al. 2012). The diversity of ants is typically higher in lowland tropical regions (Dunn et al. 2009; Economo et al. 2018), with abundance and diversity decreasing at high elevations (Longino et al. 2014). In Mesoamerica and other regions, a mid-elevation peak in ant diversity is generally observed, with montane specialist species from multiple subclades dominating the highest elevations (Longino and Branstetter 2019). In addition to natural diversity patterns, many species have also been anthropogenically redistributed across the globe, colonising areas that were previously inaccessible (Bertelsmeier 2021; Wong et al. 2023). Some of these species have had devastating ecological impacts in the ecosystems they have been introduced into (Tercel et al. 2023).

Although progress has been made towards understanding ant macrodiversity across biogeographical realms and continental and climatic scales (Janicki et al. 2016; Guénard et al. 2017), fundamental knowledge is still lacking on a local

scale, especially in tropical regions (Kass et al. 2022). This is particularly true for higher elevations that have historically been difficult to access and survey (Guénard et al. 2017; Liu et al. 2020). One clear example is Honduras, located in Mesoamerica. Historical country records originate from Wheeler (1907) and Mann (1922), who both compiled short species lists from their brief visits to the country over a century ago. Taxonomic literature and the implementation of database infrastructures have substantially changed myrmecology (e.g., Bolton 1995, 2003; [www.AntWeb.org](http://www.AntWeb.org); Janicki et al. 2016; Guénard et al. 2017); however, the Honduran ant fauna has not been reassessed since the development of such resources. Recent collections of ants for both Honduras and CNP have been made, resulting in the description of multiple novel species: *Octostruma lepticeps* Longino, 2013; *Stenamma cusuco* Branstetter, 2013; *Stenamma atribellum* Branstetter, 2013; *Temnothorax altinodus* Prebus, 2021; and *Pheidole cusuco* Longino, 2019. All but the latter species are considered restricted to CNP. However, most species records from these recent collections have not been published in comprehensive lists. One project in particular, the Leaf Litter Arthropods of MesoAmerica (LLAMA) project, collected leaf litter arthropods, including ants, across various elevational gradients throughout Mesoamerican mountain ranges (Longino et al. 2014). Alongside other sites in Honduras, the LLAMA project visited CNP in 2010. Compiling accurate species checklists is of vital importance, not only for taxonomic studies but also for providing fundamental biogeographical knowledge (Kass et al. 2022) and thus essential data for conservation efforts (Guénard et al. 2017; Liu et al. 2020). This is of particular importance considering the broad consensus of the heightened threats to biodiversity in CNP specifically and in cloud forest ecosystems generally (Bubb et al. 2004; Martin et al. 2021).

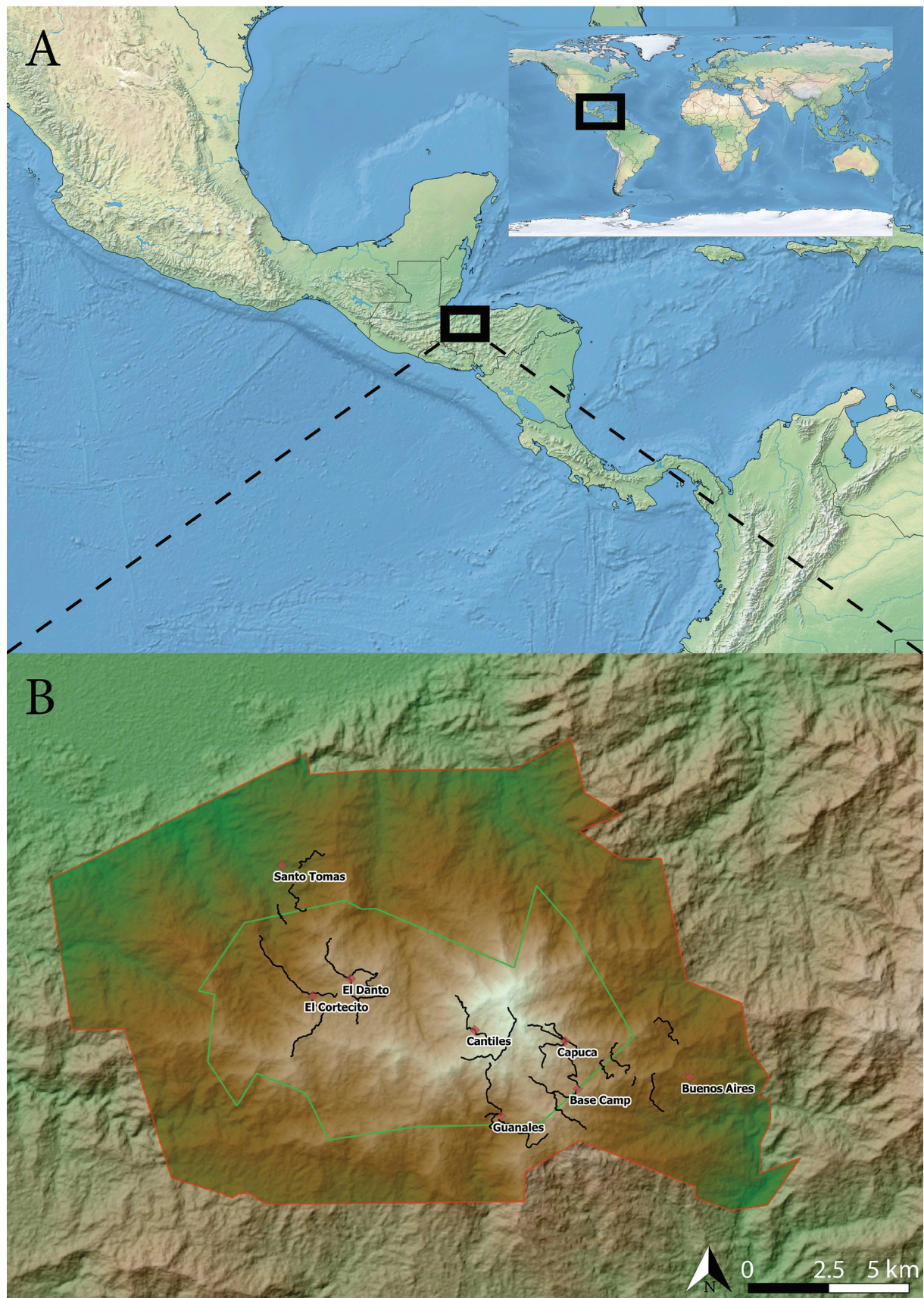
Here, we produce the first ant checklist of CNP by combining new sampling efforts with existing data records from the LLAMA project (2010). This list, as far as we are aware, represents the first checklist of the ant fauna from any Honduran site in over a century. We include information on species known to be of restricted distribution to CNP and species considered to be exotic (non-native) in the park. Knowledge gaps are highlighted in terms of considered elevation range, sampling techniques, and sampling intensity. Finally, we include an identification key for all ant genera in Honduras.

## Methods

### Study region

The CNP (15°32'31"N, 88°15'49"W) encompasses approximately 23,440 ha, with an elevational range of 500–2242 m a.s.l. (ICF 2015) (Fig. 1). The park is divided between an inner core zone (7690 ha) where settlement and resource extraction are prohibited and an encircling buffer zone (15,750 ha) where some of these practices are allowed. A closed canopy forest dominates the core zone, with a diverse community of broadleaf evergreen tree species present in the mid- to upper elevational ranges (1300–1800 m a.s.l.), interspersed with pine forest occurring mostly on the drier, eastern slopes of the park. Secondary forest, at various levels of succession, is also present below 1300 m a.s.l. as a result of commercial logging during the mid-20<sup>th</sup> century (Martin et al. 2021). At higher elevations (>1800 m a.s.l.), the upper montane rainforest, characterised





**Figure 1.** Map of Cusuco National Park. Buffer and core zone boundaries are shown (red and green, respectively) with camps (red dots) and corresponding transects with sampling subsites (black lines). Cusuco elevation data are derived from Burdgis.org (accessed 16 September 2021). Continental relief map derived from SimpleMappr, Shorthouse, 2010.

by dense growth of mosses, liverworts, ferns, and a high abundance of bromeliads, is present as a result of cool temperatures and comparatively higher rainfall. At the uppermost peak elevations (>2000 m a.s.l.), a combination of soil erosion and a lack of decomposition results in stunted but densely interwoven vegetation known as elfin forest (Martin et al. 2021).

Sampling and specimen processing

Ant-species observations from two different projects were pooled into a single dataset. Respective projects and methodologies are described below. The sampling techniques used for each project are summarised in Table 1 and Fig. 2. Sampling was completed mainly in the core zone of the park, between the elevations of 1170 and 2030 m a.s.l.

Table 1. Summary of the methods used in two ant collection projects in Cusuco National Park: LLAMA (Leaf Litter Arthropods of Mesoamerica) and MED (MyrmEcoDex: BINCO).

Sampling method	LLAMA	MED
Baiting	✓	
Berlese extraction	✓	
Hand collection	✓	✓
Malaise	✓	
Pitfall		✓
Vegetation beating	✓	
Winkler extraction	✓	✓

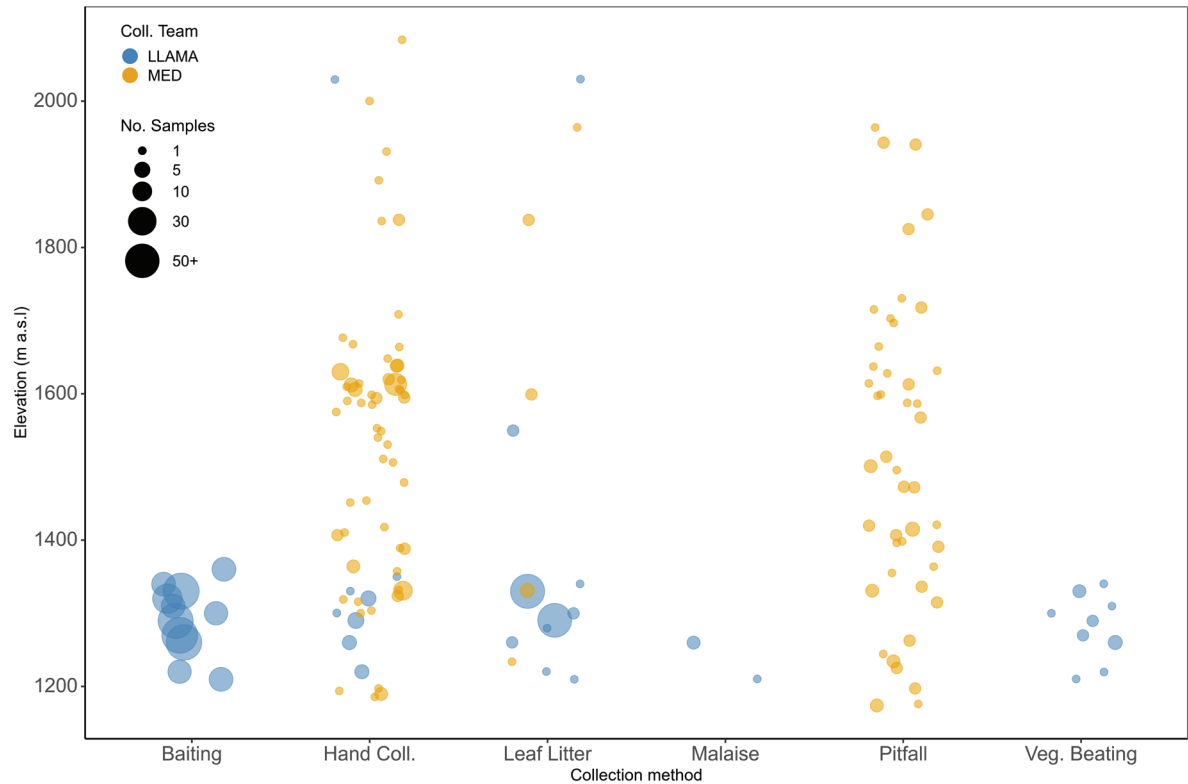


Figure 2. Distribution of sampling effort across the elevation gradient of Cusuco National Park. For the leaf litter arthropods of Mesoamerica (LLAMA) and MyrmEcoDex (MED) collections. The size of the circle corresponds to the number of samples at a specific elevation. Leaf litter sampling includes Mini-Winkler, Maxi-Winkler, and Berlese traps.



## 1. Leaf-litter arthropods of Mesoamerica (LLAMA)

Project LLAMA, funded by the U.S. National Science Foundation, sampled leaf litter-dwelling arthropods across Mesoamerica, from southern Mexico to Nicaragua, with a focus on ants and weevils (Curculionidae) (Longino et al. 2014). Specimens were collected in CNP from 29 May to 3 June 2010, during the transition from the dry season to the short wet season. Most collections were made between 1210 and 1360 m a.s.l., but additional non-quantitative samples were collected between 1580 and 2030 m a.s.l. (Fig. 2). A few additional leaf litter samples were collected earlier by R. Anderson on 24 August 1994, and included in this dataset.

### Sampling methods

Sampling was completed according to a standardised transect-based framework in mesophyll cloud forest. Arthropods were extracted from two transects of 50, 1 m<sup>2</sup> forest floor quadrats using MiniWinklers, following the methods used in Fisher (1999). Other sampling techniques used included general collection by hand, cookie baiting, vegetation beating, MaxiWinkler extraction, Berlese extraction, and Malaise trapping. The samples collected in 1994 were those obtained by the Berlese extractions.

### Specimen processing

Ants were sorted from the samples by project staff. For several reasons, only a subset of the ants present in samples were identified to species-level, with unidentified species designated a morphospecies code. Several taxa were only identified at the genus level due to taxonomic impediments, particularly within genera presenting challenges in species classification: *Azteca* Forel, 1878, *Brachymyrmex* Mayr, 1868, *Nylanderia* Emery, 1906, *Solenopsis* Westwood, 1840, and *Tapinoma* Foerster, 1850. *Hypoponera* Santschi, 1938 workers were classified at the genus level, except for two species that were readily distinguishable. *Pheidole* Westwood, 1839 workers were predominantly identified at the species level, though the rarely isolated minor workers were identified only at the genus level.

Voucher specimens were stored in regional collections in Honduras and temporarily in the Longino research collection at the University of Utah, as well as the Branstetter research collection. Comprehensive specimen data can be accessed on AntWeb ([www.AntWeb.org](http://www.AntWeb.org)). A full description of the LLAMA collection and processing methodology can be found in Longino et al. (2014).

## 2. BINCO: MyrmEcoDex (MED)

The Biodiversity Inventory for Conservation NPO (BINCO) project studies biodiversity in understudied regions globally. MyrmEcoDex (MED) is BINCO's ant workgroup. Samples were collected during Operation Wallacea (henceforth 'OpWall') biodiversity monitoring expeditions. OpWall has been conducting volunteer-funded biodiversity surveying and monitoring in CNP from June to August since 2006, operating from satellite camps distributed in the East and Western regions of CNP at different elevations. MyrmEcoDex members partic-

ipated in OpWall expeditions during the 2018 and 2019 field seasons. A total of six camps were operational: one in the buffer zone (Buenos Aires) and five in the core zone of the park (Base Camp, Guanales, Cantiles, El Danto, and El Cortecito) (Fig. 1). Each camp established three to four transects that extended into the park, which were used for surveying. Ant collections were made between 1170 and 2030 m a.s.l. Some opportunistic sampling was also completed at Santo Tomás in the lower elevational ranges of the park, a former camp that is no longer used for formal surveys.

## Sampling methods

Surveys were carried out every 3–5 days at up to eight subsites distributed at least 200 m apart along transects (Hinchcliffe et al. 2017). Four baited (horse dung) pitfall traps were deployed at each subsite. Pitfall traps were placed in a 20 × 20 m grid, 10 m from one another and 5 m from grid edges, to ensure compatibility with other plot sampling as well as to reduce interference between individual pitfalls. Ants were sorted from pitfalls in the field by MED members. During the 2019 field season, 61 out of 198 total pitfalls were screened for ants (31%). Ants extracted in 2018 only include six pitfall samples; other specimens were lost due to deterioration.

MyrmEcoDex members carried out additional sampling techniques. Ants were searched for and collected opportunistically by hand or aspirator from a variety of substrates: nests, soil, deadwood, leaves, tree bark, inside epiphytes, and others. Additionally, MaxiWinkler extractors were used to sample ants in leaf litter, with extraction times varying between 3–5 days (depending on time availability). Forty bromeliads of different sizes were also dissected leaf by leaf, and ants were collected when a colony was present.

Additional specimens originating from previous OpWall expeditions were provided by the Oxford University Museum of Natural History (OUMNH) and examined by the authors. The majority of such specimens originated from hand collection and pitfall trapping regimes, as described above.

## Specimen processing

The collected ants were stored in ethanol (70%). These specimens were sorted into morphospecies, point-mounted, and identified to the lowest taxonomic rank possible. Specimens that could not be assigned to species were given morphospecies codes. Identifications at the species level were verified by experts to ensure accuracy. The latter was facilitated via specimen pictures, taken using a quick and easy-to-use photographic setup detailed within (Mertens et al. 2017). Due to their taxonomic complexity, collected specimens of the genera *Nylanderia* as well as part of the hyperdiverse *Pheidole* were not considered for species-level identification. Specimens were deposited at the Royal Belgian Institute for Natural Sciences (RBINS) collections after identification.

## Images

A subset of species (43) were photographed, and pictures are provided in Suppl. material 3. These photographs were taken using the following setups: (1) Canon

80D with a Venus Optics Laowa 25mm f/2.8 2.5-5X Ultra Macro Lens, or EF 100mm f/2.8L Macro IS USM with Raynox 250DCR macro attachment. Images were taken using a homemade diffuser system and a manual rail system. Images were stacked in Adobe Photoshop (Adobe Inc.). (2) Canon-Cognisys set-up (Brecko et al. 2014).

### Unconfirmed identifications

Some identifications could not be confirmed and are marked as cf. (Latin: *confer*) or by a summation in the species epithet. These specimens appear similar to the named species, but verification was not possible. Verification requires more specimens and comparison with morphologically similar species.

### Spatial distribution status

An assessment of biogeographic distribution status was made for all recorded species in this study using Antmaps (Janicki et al. 2016; Guénard et al. 2017). The following categories were applied: regionally restricted (to Honduras), exotic (to Honduras, i.e., non-native), and globally invasive species (showing wide global occurrence patterns). Species not previously reported in Honduras were also noted.

### Species accumulation

Species accumulation curves were made to provide insight into sample completion and method efficiency. By assessing species richness cumulatively per additional sample, we show the intensity of individual sampling techniques, respectively, and the potential for collecting additional species with additional sampling. Accumulated species richness was also compared with sampling coverage of the community, which is the probability that an individual of the entire ant community belongs to a species that has been sampled before. As sampling techniques each address a different subset of the total ant community, respective subset communities are considered. Species presence-absence matrices using unique sample codes as individual sampling units were built per collection methodology. Non-species-level identified specimens and OUMNH material were excluded as a result of low taxonomic resolution and a lack of collection codes, respectively. Final accumulation curves and summary statistics were generated with the iNEXT R package using the first Hill number (species richness) (Chao et al. 2014; Hsieh et al. 2022).

### Identification Key: Ant genera of Honduras

To improve the accessibility of this work and the Honduran ant fauna in general, a dichotomous identification key was constructed for ant genera for the whole of Honduras (Suppl. material 2). Genera occurring in Honduras were determined using records from Antweb ([www.AntWeb.org](http://www.AntWeb.org)). The identification key was constructed manually by combining multiple works and identification keys on relevant taxa (Ward 1985; Hölldobler and Wilson 1990; Shattuck 1992; Bolton 1995; Longino 2007; Wild 2007; Donoso 2012; Fayle et al. 2014; Schmidt and Shattuck 2014; Baccaro et al. 2015; Borowiec 2016; Ward and Fisher 2016;



Ward et al. 2016; Williams and Lapolla 2016; Solomon et al. 2019; Prebus 2021b; Camacho et al. 2022; [www.Antwiki.org](http://www.Antwiki.org)), with respective citations provided (provided as Suppl. material 2).

## Results

### Ant fauna of the CNP

Across all sampling projects, a total of 5690 ants were collected in CNP, resulting in nine subfamilies comprising 60 genera and 162 species (Table 2). Appendix 1 provides a complete list of all ant species found in CNP, with the respective sampling method, recorded elevational distribution ranges, and habitat (data also in Suppl. material 1). The complete specimen data is made available as well (Suppl. material 4). Characteristic specimens from our collections are shown in Fig. 3.

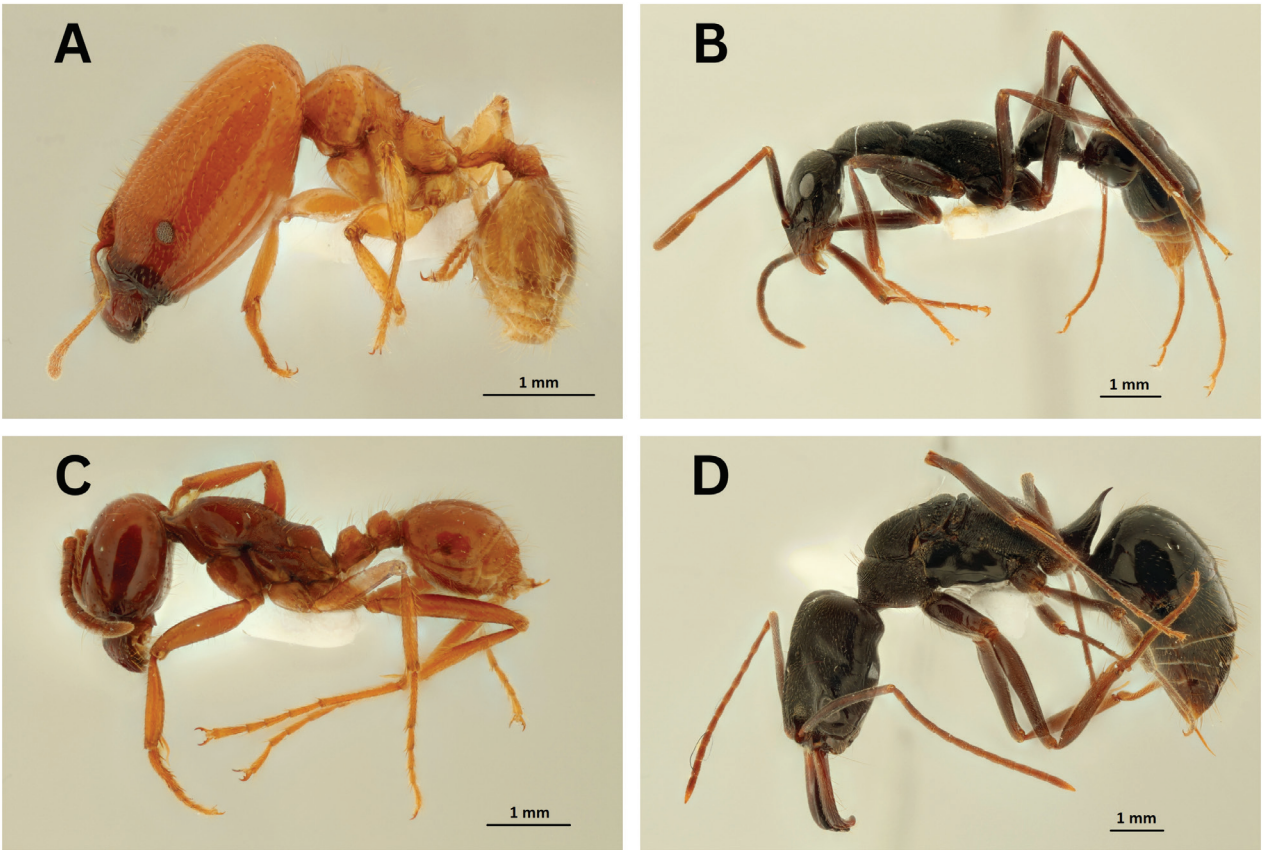
In addition to the first 127 species collected by LLAMA (3445 specimens), the MED collections resulted in an additional 41 species for CNP. The latter yielded a total of 78 species and 2155 specimens: 894 in 2018 and 1261 in 2019. Of these, 286 and 419 were mounted, respectively, and added to the RBINS collections (together with the remaining specimens in ethanol). The checklist also includes 90 mounted specimens from the OUMNH, collected during earlier field surveys in CNP and identified by MED.

Some ant species in CNP have notable distributions (Table 3). Seven species are regionally restricted to Honduras, and two species are exotic. Three species have global distributions (including the two exotic species), and five species were recorded for the first time in Honduras.

We report the first collection of male *Pheidole balatro* individuals (Fig. 4). Males of this species were previously unknown. Six male individuals were collected alongside minor and major workers of this species from a nest residing inside a bromeliad plant (Base Camp, 26 June 2019, 1613 m a.s.l.). The nest was basally located in the bromeliad between the leaves and was discovered by removing leaves from their basal attachment. All six specimens of *P. balatro* males were stored in the RBINS collections (three mounted, three preserved in ethanol; sample code: CNP-222).

**Table 2.** Composition of genera and species per subfamily contributing to total species richness detected in Cusuco National Park.

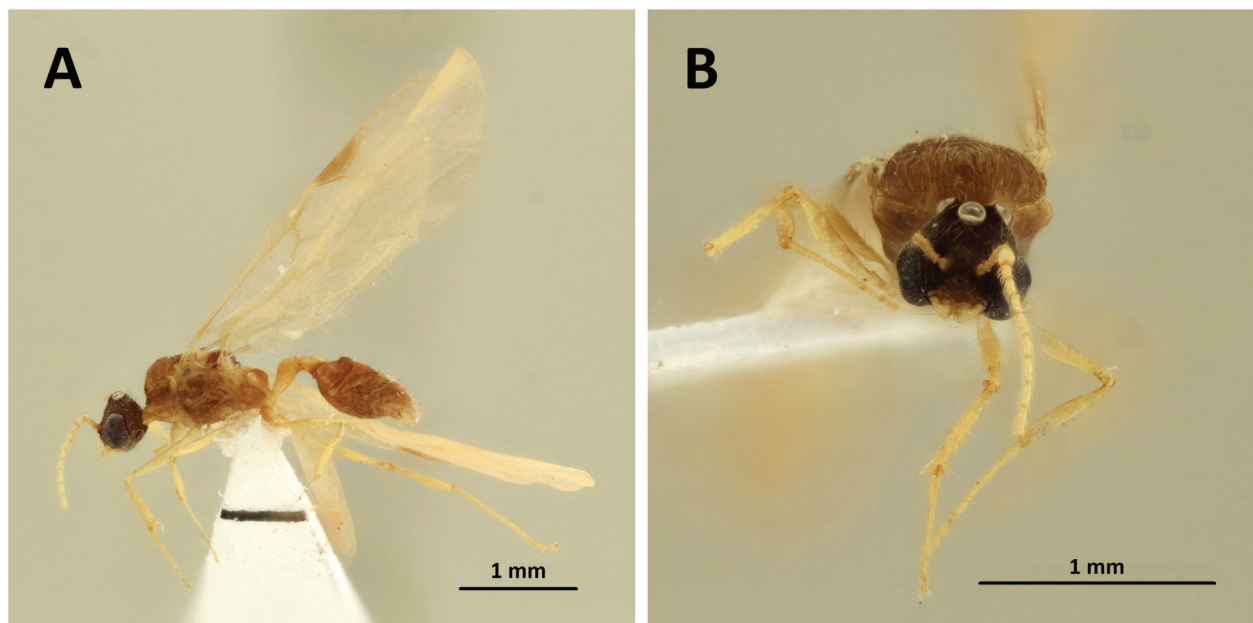
Subfamily	Genera	Species
Amblyoponinae	1 (2%)	1 (1%)
Dolichoderinae	5 (8%)	8 (5%)
Dorylinae	7 (12%)	14 (9%)
Ectatomminae	4 (7%)	9 (6%)
Formicinae	4 (7%)	10 (6%)
Myrmicinae	24 (40%)	88 (54%)
Ponerinae	12 (20%)	26 (16%)
Proceratiinae	2 (3%)	2 (1%)
Pseudomyrmecinae	1 (2%)	5 (3%)
<b>Total</b>	<b>60</b>	<b>162</b>



**Figure 3.** Select ant species collected in Cusuco National Park. Top: *Pheidole absurda* (A major worker), *Leptogenys imperatrix* (B worker); Bottom: *Labidus coecus* (C major worker), *Odontomachus yucatecus* (D wingless female).

**Table 3.** Ant species from Cusuco National Park with notable geographic distributions: known distribution restricted to Honduras (*P. cusuco* in CNP and just across the Guatemalan border), exotic to Honduras, and/or globally invasive. The first records for Honduras are also shown. \**P. cusuco* is in CNP and just across the Guatemalan border.

Species	First Honduran record	Regionally restricted	Exotic	Globally invasive
<i>Leptogenys bifida</i> Lattke, 2011		✓		
<i>Leptogenys honduriana</i> Mann, 1922		✓		
<i>Monomorium pharaonis</i> Linnaeus, 1758			✓	✓
<i>Octostruma leptocephs</i> Longino, 2013		✓		
<i>Pheidole cusuco</i> * Longino, 2019		✓		
<i>Pheidole natalie</i> Longino, 2019	✓			
<i>Pseudomyrmex pallens</i> Mayr, 1870	✓			
<i>Strumigenys</i> cf. <i>calamita</i>	✓			
<i>Solenopsis texana/carolinensis</i>	✓			
<i>Solenopsis invicta</i> Buren, 1972	✓		✓	✓
<i>Solenopsis geminata</i> Fabricius, 1804				✓
<i>Stenamma atribellum</i> Branstetter, 2013		✓		
<i>Stenamma cusuco</i> Branstetter, 2013		✓		
<i>Temnothorax altinodus</i> Prebus, 2021		✓		



**Figure 4.** *Pheidole balatro* male specimen (Specimen code: CNP-222-3-3) **A** lateral and **B** frontal view.

### Sampling elevation and methodology

Sampling efforts across the elevation range varied between LLAMA and MED (Fig. 2). There were also differences in the collection techniques used between the two datasets. LLAMA focused on a narrower elevation range between ~1210 and 1360 m a.s.l. (with the exception of three higher samples at ~1580 m and ~2030 m a.s.l.), using thorough sampling in a standardised framework. A large part of the ant diversity in CNP (127 species) was thus recorded in a narrower elevation range using four sampling techniques: leaf litter extraction, bait trapping, Malaise trapping, and vegetation beating. Project MED considered a broader range of elevation from ~1170–1960 m a.s.l., employing less exhaustive sampling. A different set of sampling techniques was employed (pitfall traps and more hand collections) at wider elevation ranges, including higher elevations. This resulted in the collection of a different subset of the ant fauna (78 species total; 41 newly reported). Additional species were even collected at the same narrow elevation range just by changing techniques (e.g., *Odontomachus yucatecus*).

### Species richness: sampling effort and efficiency

Leaf litter extraction obtained the greatest proportion of genera (26%) and species (31%) compared to other methods, with hand collection also comprising a large proportion of genera (20%) and species (21%) (Table 4). Baiting, Malaise, leaf litter extraction, and vegetation beating were predominantly conducted at relatively lower elevational ranges (Fig. 2). Pitfall traps and hand collection events were distributed more evenly across the elevation range in contrast, with a concentration of hand collection events at 1600 m a. s. l. To reduce interpretation bias on the efficiency of sampling techniques, Table 4 complements Fig. 2, illustrating sampling effort.

**Table 4.** Representation of subfamilies, genera, and species collected per respective sampling method (in respective elevational ranges), alongside asymptotic estimated species richness (with 95% confidence intervals). Percentage of total ant fauna collected by individual sampling methods, per taxonomic level. See Fig. 2 for complementary data regarding sampling intensities across the CNP elevation gradient.

Sampling method	Subfamily	Genera	Species	Estimated richness (95% CI)	Sample coverage
Baiting	6 (67%)	20 (33%)	42 (26%)	71 (51–139)	0.978
Hand collection	7 (78%)	30 (50%)	62 (39%)	114 (75–225)	0.835
Leaf litter extraction	7 (78%)	39 (65%)	92 (56%)	127 (105–183)	0.977
Malaise	7 (78%)	22 (37%)	30 (18%)	62 (40–128)	0.440
Pitfall	6 (67%)	23 (38%)	38 (23%)	65 (47–354)	0.916
Vegetation Beating	7 (78%)	16 (27%)	29 (18%)	50 (40–88)	0.815

Fig. 5 presents species accumulation curves, showing the captured species richness and the sampling intensity (observed and extrapolated). Additional sampling for each individual sampling technique is expected to result in the capture of more species, with a total asymptotic richness estimating 257 (95% confidence interval: 204–365) species when all sampling efforts are combined. The maximum rate of increase in species richness seems not to have been reached yet for vegetation beating and Malaise trapping. However, for leaf litter extraction, hand collection, pitfall trapping, and baiting, it appears that the maximum rate has been reached.

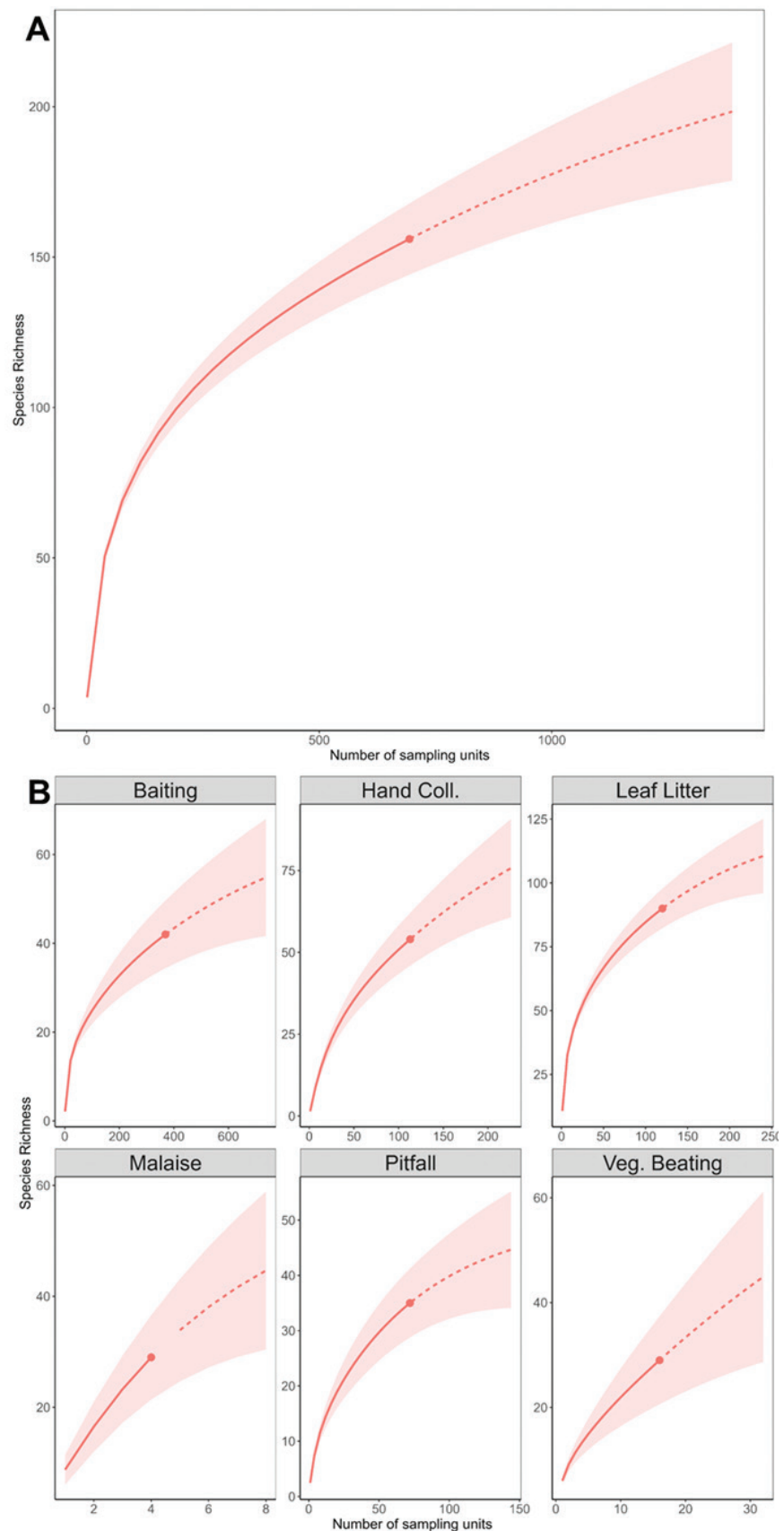
The relation between species richness and community sample coverage is presented in Fig. 6. As stated before, sample coverage concerns the probability that an individual of the entire ant community addressed per sampling technique belongs to a species that has been sampled before. High values show low probability ( $1 - \text{sample coverage}$ ) of sampling additional species for each technique; low values show high probabilities to find species that are yet unaccounted for. The total sampling considered in this study shows high community sample coverage with a value of 0.978. There are, however, differences between individual sampling techniques. Leaf litter extraction, pitfall trapping, and baiting show a community sample coverage of  $>0.9$  (0.977, 0.916, and 0.978, respectively), whereas this value is lower for hand collection, vegetation beating, and Malaise trapping (0.835, 0.815, and 0.44, respectively). The extrapolation shows the expected increase in species richness with greater sampling effort. The 95% confidence intervals are low for most techniques, except for Malaise trapping, which had a considerably lower effort compared with other sampling methods.

## Discussion

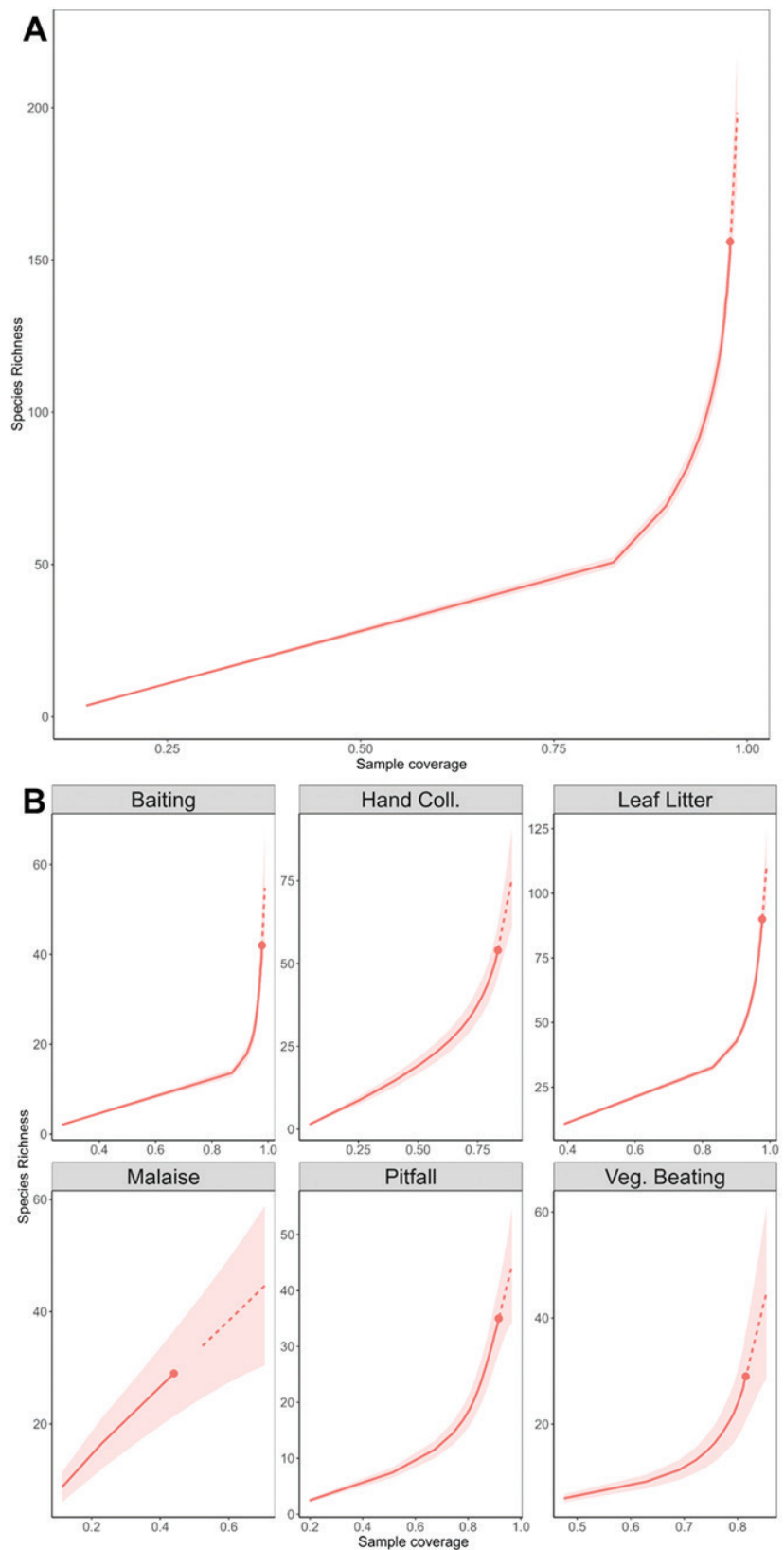
### Ant fauna of the CNP

Our research confirms that CNP supports a high richness of ant species, on par with many other taxa in the park (Martin et al. 2021). We present the first checklist for the ants of CNP, which is also the first species list published for ants in Honduras in over a century. With the inclusion of a genus-level identifi-





**Figure 5.** Species accumulation curves according to the number of samples taken. Shaded regions represent 95% confidence intervals **A** total species richness for the number of samples taken **B** species richness for the number of samples taken for each technique.



**Figure 6.** Species accumulation curves according to community sample coverage. Sample coverage is the probability for an individual in the entire ant community to belong to a species that has been sampled before. Shaded regions represent 95% confidence intervals **A** total species richness for the sample coverage **B** species richness for the sample coverage for each technique.

cation key for all Honduran ant taxa (Suppl. material 2), we hope this work will stimulate further ant research in Honduras.

Ant species composition showed patterns consistent with other ant faunal inventories (Yanoviak and Kaspari 2000; Patrick et al. 2012; Donoso 2017), with Myrmicinae, Formicinae, Ponerinae, and Dolichoderinae comprising over 80% of species collected (Table 2). Compared to other subfamilies, Dorylinae contributed a substantial number of genera (7), considering only 14 species were recorded.

Seven ant species recorded in CNP are spatially restricted to the region. According to current known occurrences, six species are regionally restricted to Honduras. Of these, four are restricted to CNP: *Octostruma leptoceps*, *Stenamma cusuco*, *S. atribellum*, and *Temnothorax altinodus*. Two other species restricted to Honduras belong to the genus *Leptogenys*, previously reported from La Ceiba (*Leptogenys bifida*) and Lombardia (*Leptogenys honduriana*). *Pheidole cusuco* was originally described by the CNP and has also been reported just across the Guatemalan border (Longino 2019). This indicates the importance of CNP as a potential refuge for these spatially restricted species, which has implications for their conservation. Conversely, two non-native ant species were recorded: *Monomorium pharaonis* (at the edge of the core zone) and *Solenopsis invicta* (in the buffer zone, a deforested area). Both species show globally invasive distribution patterns, as does *Solenopsis geminata*, which is native to the region (recorded in both the core and buffer zones). This suggests that the core zone of CNP remains largely unaffected by highly invasive ant species. Five species represent new records for Honduras: *Pheidole natalie*, *Strumigenys* cf. *calamita*, *Solenopsis invicta*, *Solenopsis texana/carolinensis*, and *Pseudomyrmex pallens*. The relatively high number of new country records identified in a single protected area indicates the understudied nature of the region, confirming the need for more field surveys.

The discovery of *Pheidole balatro* males is another noteworthy find. Given the large diversity of *Pheidole* ants, it is common practice not to describe newly discovered males through extensive morphological descriptions, which we have adopted here. The provided photograph and information in the results section could, however, provide a useful basis for comparison.

The genus *Procryptocerus* is understudied and requires greater taxonomic attention in order to better understand populations and species delimitations. Though we present *P. batesi* Forel, 1899, here, we were unable to exclude the possibility of *P. mayri* Forel, 1899, given that they are very morphologically similar.

### Species richness: sampling effort and efficiency

Results suggest that the recorded species richness (162) appears to be an underrepresentation of the actual species richness in CNP (asymptotic estimate 257; 95% CI: 204–365) (Figs 5, 6). The observed and extrapolated patterns in Fig. 5 can be interpreted as the relative efficiency of techniques for capturing different ant species. Since a plateau of species richness is not reached for any of the sampling techniques, all show much potential to record additional species with additional sampling. However, the rate of increase in species richness appears to have reached a maximum for most all techniques, meaning that greater sampling will be required to keep finding additional species.

Community sample coverage is relatively high overall, suggesting that our sampling is rather representative of the ant community in CNP. This holds true for the communities addressed by leaf litter extraction, pitfall trapping, and baiting (CSC >0.9). For these techniques, the probabilities of sampling additional species are relatively low. It seems that high community sample coverage only increases mildly with additional species. Species that have not been recorded yet at this point in the accumulation curve seem to play only a small role in the ant community and are likely to be rare. However, hand collection, vegetation beating, and Malaise trapping still seem to show underrepresentation of the community (CSC <0.9). The relative probabilities of sampling additional species are relatively higher when compared to the other three techniques. Hand collection and vegetation beating still show relatively high sample coverage (CSC = 0.8–0.9), with trends suggesting that some common species might still be added using these techniques. Malaise trapping shows a low sample coverage of <0.5 and thus might yield many more species, both common and rare. Sample coverage for hand collection might be lower because of less consistent sampling along the elevation range. For vegetation beating and Malaise trapping, this is probably due to lower sampling numbers (N = 4 and 16, respectively). It is interesting to consider that just four Malaise traps captured a similar number of ant genera as extensive pitfall trapping, although the collected genera are more typical of arboreal ant fauna, such as *Procryptocerus*, *Pseudomyrmex*, and *Crematogaster*, indicating an alternative faunal community sampled.

The elevational range addressed with each technique is to be considered. The species accumulation rate of leaf litter extraction, baiting, vegetation beating, and Malaise trapping is expected to increase when used along larger elevational ranges, especially when including higher altitudes, which will likely collect altitude specialist species and subclades (e.g., *Stenamma*).

### Knowledge gaps and research potential CNP

Despite the substantial species list accumulated from the two projects, there remains high potential to add more species. This study confirmed the presence of 162 ant species; however, a total of 250+ species is predicted to be present in CNP. Knowledge gaps are presented below, which could be considered in order to obtain a more complete ant species inventory for the park. First of all, it is important to note that there is a lack of specific canopy and subterranean sampling. Though Malaise trapping and leaf litter extraction may sample a subset of those communities, more species are likely present and have yet to be collected.

By expanding methodological approaches and sampling along a broader elevational range, we increased the number of ant species recorded from CNP. However, there are still some elevation zones that were not sampled using the primary survey methods (Fig. 2), and an unequal sampling effort was used along the elevation gradient. To address these sampling gaps, we recommend employing a variety of sampling techniques along the full elevational gradient with appropriate replication in order to ensure the inclusion of less prevalent species.

The middle elevation ranges were sampled most intensely; however, the higher (mountain peaks) and lower elevation ranges (e.g., buffer zone) remain undersampled. Leaf litter extraction results in the highest number of genera



and species recorded, followed by hand collection and pitfall trapping. Although hand collection and pitfall trapping were used along a broader elevational gradient than leaf litter extraction, the latter still shows a higher number of ant species captured. Sampling leaf litter at higher altitudes, in particular, should provide promising results. Baiting is still unexplored at higher altitudes. Additional Malaise trapping and vegetation beating are recommended in general, regardless of elevation range.

As a hotspot of biodiversity, numerous novel species have been previously described from CNP (Martin et al. 2021), including ants (Branstetter 2013; Longino 2013, 2019; Prebus 2021a). Further targeted surveys in CNP are expected to lead to the discovery of more ant species, especially at the higher, undersampled altitudes and buffer zones.

Sampling in a standardised framework would allow for a better understanding of species ecology and the taxonomy of the local ant fauna, which could then lead to improved knowledge of regional diversity and wider biogeographical patterns (especially for highly understudied groups such as those in the genera *Apterostigma*, *Procryptocerus*, and *Temnothorax*). The effect of anthropogenic habitat change could also be examined, given persistent habitat alterations across both core and buffer zones.

## Conclusion

CNP has a rich and diverse ant fauna with the potential to serve as a study site for addressing a multitude of research questions concerning ants. Other tropical mountain cloud forests in Honduras could hold similar ant species richness, with most of these being even more understudied and lacking any survey data. The materials we provide here could form a baseline for future work related to ants in other Honduran cloud forests, helping to guide research in these otherwise poorly explored yet highly threatened localities.

## Acknowledgements

We thank Operation Wallacea and Expediciones y Servicios Ambientales de Cusuco for their logistical support and the Instituto de Conservación Forestal for issuing research permits to support ongoing research in Cusuco. We also thank our local guides for their assistance and the numerous Operation Wallacea volunteers for their help with data collection. We are grateful to the Oxford University Museum of Natural History for providing additional specimens. Thanks should go to John T. Longino for his help in accessing the LLAMA data and for verifying several identifications. We would also like to extend our gratitude to Thomas J. Creedy for providing the pitfall trap specimens, as well as to Vivienne Martin for proof-reading the manuscript. Finally, we would like to thank Martijn Van Roie and Jan Mertens (BINCO: RePhoCus) for assisting with photography during the identification process. USDA is an equal opportunity provider and employer.

## Additional information

### Conflict of interest

The authors have declared that no competing interests exist.

## Ethical statement

No ethical statement was reported.

## Funding

LLAMA was funded by a National Science Foundation grant (DEB-0640015).

## Author contributions

According to the author order and the credit categories: Conceptualization (FCDW, MJ, and MTH); Methodology (FCDW, DO, and MTH); Validation (FCDW, MGB, WD, and MTH); Formal analysis (FCDW and MTH); Investigation (FCDW, DO, DDG, JS, RVO, and MTH); Resources (TEM, WD, and MJ); Data curation (FCDW and MTH); Writing – original draft (FCDW and MTH); Writing – review and editing (FCDW, DO, MGB, DDG, WD, MJ, TEM, JS, RVO, and MTH); Visualisation (FCDW, WD, and MTH); Supervision (FCDW and MJ); Project administration (FCDW).

## Author ORCIDs

Frederik C. De Wint  <https://orcid.org/0000-0003-0819-3903>

Michael G. Branstetter  <https://orcid.org/0000-0002-3734-6166>

Wouter Dekoninck  <https://orcid.org/0000-0002-7065-7310>

Merlijn Jocque  <https://orcid.org/0000-0002-7196-7476>

Jennifer Sudworth  <https://orcid.org/0009-0005-2994-5205>

Matthew T. Hamer  <https://orcid.org/0000-0001-6728-9046>

## Data availability

All of the data that support the findings of this study are available in the main text or Supplementary Information.

## References

- Anderson RS, Ashe JS (2000) Leaf litter inhabiting beetles as surrogates for establishing priorities for conservation of selected tropical montane cloud forests in Honduras, Central America (Coleoptera, Staphylinidae, Curculionidae). *Biodiversity and Conservation* 9(5): 617–653. <https://doi.org/10.1023/A:1008937017058>
- Baccaro FB, Feitosa RM, Fernández IO, Izzo TJ, Souza JD, Solar R (2015) Guia para os gêneros de formigas do Brasil. Editora INPA, Manaus.
- Bertelsmeier C (2021) Globalization and the anthropogenic spread of invasive social insects. *Current Opinion in Insect Science* 46: 16–23. <https://doi.org/10.1016/j.cois.2021.01.006>
- Bolton B (1995) A New General Catalogue of the Ants of the World. Harvard University Press. <https://www.hup.harvard.edu/books/9780674615144> [January 27, 2024]
- Bolton B (2003) Synopsis and Classification of Formicidae. *Memoirs of the American Entomological Institute*, 370 pp.
- Borowiec M (2016) Generic revision of the ant subfamily Dorylinae (Hymenoptera, Formicidae). *ZooKeys* 608: 1–280. <https://doi.org/10.3897/zookeys.608.9427>
- Branstetter M (2013) Revision of the Middle American clade of the ant genus *Stenammina* Westwood (Hymenoptera, Formicidae, Myrmicinae). *ZooKeys* 295: 1–277. <https://doi.org/10.3897/zookeys.295.4905>
- Brecko J, Mathys A, Dekoninck W, Laponce M, VandenSpiegel D, Semal P (2014) Focus stacking: Comparing commercial top-end set-ups with a semi-automatic low budget

- approach. A possible solution for mass digitization of type specimens. *ZooKeys* 464: 1–23. <https://doi.org/10.3897/zookeys.464.8615>
- Bubb P, May I, Miles L, Sayer J (2004) Cloud forest agenda. United Nations Environment Program World Conservation Monitoring Centre, Cambridge. [https://web.archive.org/web/20180423095416id\\_/https://www.ourplanet.com/wcmc/pdfs/cloudforests.pdf](https://web.archive.org/web/20180423095416id_/https://www.ourplanet.com/wcmc/pdfs/cloudforests.pdf)
- Camacho GP, Franco W, Branstetter MG, Pie MR, Longino JT, Schultz TR, Feitosa RM (2022) UCE Phylogenomics Resolves Major Relationships Among Ectaheteromorph Ants (Hymenoptera, Formicidae, Ectatomminae, Heteroponerinae): A New Classification For the Subfamilies and the Description of a New Genus. *Insect Systematics and Diversity* 6: 5. <https://doi.org/10.1093/isd/ixab026>
- Chao A, Gotelli NJ, Hsieh TC, Sander EL, Ma KH, Colwell RK, Ellison AM (2014) Rarefaction and extrapolation with Hill numbers: A framework for sampling and estimation in species diversity studies. *Ecological Monographs* 84(1): 45–67. <https://doi.org/10.1890/13-0133.1>
- Damron BN, Pinto-Da-Rocha R, Longhorn SJ (2018) Description of a new species of Eucynorta (Opiliones, Cosmetidae) from Cortés, Honduras. *Zootaxa* 4450(1): 125–134. <https://doi.org/10.11646/zootaxa.4450.1.9>
- Del Toro I, Ribbons R, Pelini S (2012) The little things that run the world revisited: A review of ant-mediated ecosystem services and disservices (Hymenoptera, Formicidae). *Myrmecological News* 17: 133–146.
- Donoso DA (2012) Additions to the taxonomy of the armadillo ants (Hymenoptera, Formicidae, Tatuidris). *Zootaxa* 3503(1): 61. <https://doi.org/10.11646/zootaxa.3503.1.5>
- Donoso DA (2017) Tropical ant communities are in long-term equilibrium. *Ecological Indicators* 83: 515–523. <https://doi.org/10.1016/j.ecolind.2017.03.022>
- Dunn RR, Agosti D, Andersen AN, Arnan X, Bruhl CA, Cerdá X, Ellison AM, Fisher BL, Fitzpatrick MC, Gibb H, Gotelli NJ, Gove AD, Guenard B, Janda M, Kaspari M, Laurent EJ, Lessard J, Longino JT, Majer JD, Menke SB, McGlynn TP, Parr CL, Philpott SM, Pfeiffer M, Retana J, Suarez AV, Vasconcelos HL, Weiser MD, Sanders NJ (2009) Climatic drivers of hemispheric asymmetry in global patterns of ant species richness. *Ecology Letters* 12(4): 324–333. <https://doi.org/10.1111/j.1461-0248.2009.01291.x>
- Economo EP, Narula N, Friedman NR, Weiser MD, Guénard B (2018) Macroecology and macroevolution of the latitudinal diversity gradient in ants. *Nature Communications* 9(1): 1778. <https://doi.org/10.1038/s41467-018-04218-4>
- Ellenberg H (1959) Typen tropischer Urwalder in Peru. *Schweizerische Zeitschrift für Forstwesen* 3: 169–187.
- Fayle TM, Yusah KM, Hashimoto Y (2014) Key to the Ant Genera of Borneo in English and Malay. <https://www.tomfayle.com/Ant%20key.htm>
- Fisher BL (1999) Improving inventory efficiency: A case study of leaf-litter ant diversity in Madagascar. *Ecological Applications* 9(2): 714–731. [https://doi.org/10.1890/1051-0761\(1999\)009\[0714:IIEACS\]2.0.CO;2](https://doi.org/10.1890/1051-0761(1999)009[0714:IIEACS]2.0.CO;2)
- Freeman BG, Scholer MN, Ruiz-Gutierrez V, Fitzpatrick JW (2018) Climate change causes upslope shifts and mountaintop extirpations in a tropical bird community. *Proceedings of the National Academy of Sciences of the United States of America* 115(47): 11982–11987. <https://doi.org/10.1073/pnas.1804224115>
- Guénard B, Weiser MD, Gómez K, Narula N, Economo EP (2017) The Global Ant Biodiversity Informatics (GABI) database: synthesizing data on the geographic distribution of ant species (Hymenoptera, Formicidae). *Myrmecological News* 24: 83–89. [https://doi.org/10.25849/MYRMECOL.NEWS\\_024:083](https://doi.org/10.25849/MYRMECOL.NEWS_024:083)

- Hansen MC, Wang L, Song X-P, Tyukavina A, Turubanova S, Potapov PV, Stehman SV (2020) The fate of tropical forest fragments. *Science Advances* 6(11): eaax8574. <https://doi.org/10.1126/sciadv.aax8574>
- Hinchcliffe D, Jones S, Vulinec K, Thompson P, Hoskins H, Creedy T, Jocque M, Green S, Lonsdale G, Phipps C, Burdekin O, Brown T (2017) Operation Wallacea – Cusuco National Park, Honduras 2016 & 2017: End of Season Report.
- Hölldobler B, Wilson EO (1990) *The Ants*. Harvard University Press, 732 pp. <https://doi.org/10.1007/978-3-662-10306-7>
- Hsieh TC, Ma KH, Chao A (2022) iNext: Interpolation and Extrapolation for Species Diversity. R package version 3.0.0. [http://chao.stat.nthu.edu.tw/wordpress/software\\_download/](http://chao.stat.nthu.edu.tw/wordpress/software_download/)
- ICF (2015) Anuario Estadístico Forestal de Honduras. Instituto Nacional de Conservación y Desarrollo forestal, Áreas Protegidas y Vida Silvestre, Honduras.
- Janicki J, Narula N, Ziegler M, Guénard B, Economo EP (2016) Visualizing and interacting with large-volume biodiversity data using client–server web-mapping applications: The design and implementation of antmaps.org. *Ecological Informatics* 32: 185–193. <https://doi.org/10.1016/j.ecoinf.2016.02.006>
- Jocque M, Garrison R (2022) Dragonflies of Cusuco National Park, Honduras; checklist, new country records and the description of a new species of *Palaemnema* Selys, 1860 (Odonata: Platystictidae). *Zootaxa* 5188(5): 453–476. <https://doi.org/10.11646/zootaxa.5188.5.3>
- Jones RW, O'brien CW, Ruiz-Montoya L, Gómez-Gómez B (2008) Insect Diversity of Tropical Montane Forests: Diversity and Spatial Distribution of Weevils (Coleoptera, Curculionidae) Inhabiting Leaf Litter in Southern Mexico. *Annals of the Entomological Society of America* 101(1): 128–139. [https://doi.org/10.1603/0013-8746\(2008\)101\[128:IDOTMF\]2.0.CO;2](https://doi.org/10.1603/0013-8746(2008)101[128:IDOTMF]2.0.CO;2)
- Kass JM, Guénard B, Dudley KL, Jenkins CN, Azuma F, Fisher BL, Parr CL, Gibb H, Longino JT, Ward PS, Chao A, Lubertazzi D, Weiser M, Jetz W, Guralnick R, Blatrix R, Lauriers JD, Donoso DA, Georgiadis C, Gomez K, Hawkes PG, Johnson RA, Lattke JE, MacGown JA, Mackay W, Robson S, Sanders NJ, Dunn RR, Economo EP (2022) The global distribution of known and undiscovered ant biodiversity. *Science Advances* 8(31): eabp9908. <https://doi.org/10.1126/sciadv.abp9908>
- Le Saout S, Hoffmann M, Shi Y, Hughes A, Bernard C, Brooks TM, Bertzky B, Butchart SHM, Stuart SN, Badman T, Rodrigues ASL (2013) Protected Areas and Effective Biodiversity Conservation. *Science* 342(6160): 803–805. <https://doi.org/10.1126/science.1239268>
- Liu C, Fischer G, Hita Garcia F, Yamane S, Liu Q, Peng YQ, Economo EP, Guénard B, Pierce NE (2020) Ants of the Hengduan Mountains: A new altitudinal survey and updated checklist for Yunnan Province highlight an understudied insect biodiversity hotspot. *ZooKeys* 978: 1–171. <https://doi.org/10.3897/zookeys.978.55767>
- Long A (1995) Restricted-range and threatened bird species in tropical montane cloud forests. *Tropical montane cloud forests*. Springer, New York, 47–75.
- Longino JT (2007) A taxonomic review of the genus *Azteca* (Hymenoptera, Formicidae) in Costa Rica and a global revision of the aurita group. *Zootaxa* 1491(1): 1–63. <https://doi.org/10.11646/zootaxa.1491.1.1>
- Longino JT (2013) A revision of the ant genus *Octostruma* Forel 1912 (Hymenoptera, Formicidae). *Zootaxa* 3699(1): 1. <https://doi.org/10.11646/zootaxa.3699.1.1>
- Longino JT (2019) *Pheidole* (Hymenoptera, Formicidae) of Middle American Wet Forest. *Zootaxa* 4599(1): 1–126. <https://doi.org/10.11646/zootaxa.4599.1.1>



- Longino JT, Branstetter MG (2019) The truncated bell: An enigmatic but pervasive elevational diversity pattern in Middle American ants. *Ecography* 42(2): 272–283. <https://doi.org/10.1111/ecog.03871>
- Longino JT, Branstetter MG, Colwell RK (2014) How Ants Drop Out: Ant Abundance on Tropical Mountains. *PLoS ONE* 9: e104030. <https://doi.org/10.1371/journal.pone.0104030>
- Mann WM (1922) 61 Ants from Honduras and Guatemala. US Government Printing Office. <https://doi.org/10.5479/si.00963801.61-2434.1>
- Martin T, Jones S, Hoskins H, McCann N, Batke S, Kelly D, Kolby J, Downing R, Zelaya S, Green S, Lonsdale G, Brown T, Waters S, Rodriguez-Vasquez F, McCravy K, D'Souza M, Crace D, Nunez-Mino J, Haelewaters D, Berkum P (2021) A review of the ecological value of Cusuco National Park: an urgent call for conservation action in a highly threatened Mesoamerican cloud forest. *Journal of Mesoamerican Biology*. <https://research.edgehill.ac.uk/en/publications/a-review-of-the-ecological-value-of-cusuco-national-park-an-urgent>
- Mendes HF, Andersen T, Jocqué M (2011) A new species of *Polypedilum* Kieffer from bromeliads in Parque Nacional Cusuco, Honduras (Chironomidae: Chironominae). *Zootaxa* 3062(1): 46. <https://doi.org/10.11646/zootaxa3062.1.5>
- Mertens JE, Van Roie M, Merckx J, Dekoninck W (2017) The use of low cost compact cameras with focus stacking functionality in entomological digitization projects. *ZooKeys* (712): 141. <https://doi.org/10.3897/zookeys.712.20505>
- Moreira X, Mooney KA, Zas R, Sampedro L (2012) Bottom-up effects of host-plant species diversity and top-down effects of ants interactively increase plant performance. *Proceedings. Biological Sciences* 279(1746): 4464–4472. <https://doi.org/10.1098/rspb.2012.0893>
- Myers N, Mittermeier RA, Mittermeier CG, Da Fonseca GAB, Kent J (2000) Biodiversity hotspots for conservation priorities. *Nature* 403(6772): 853–858. <https://doi.org/10.1038/35002501>
- Patrick M, Fowler D, Dunn RR, Sanders NJ (2012) Effects of Treefall Gap Disturbances on Ant Assemblages in a Tropical Montane Cloud Forest. *Biotropica* 44(4): 472–478. <https://doi.org/10.1111/j.1744-7429.2012.00855.x>
- Pinto R, Jocque M (2013) A new species of *Elpidium* (Crustacea, Ostracoda) from bromeliads in Cusuco National Park, Honduras. *ZooKeys* 313: 45–59. <https://doi.org/10.3897/zookeys.313.4904>
- Prebus MM (2021a) Phylogenomic species delimitation in the ants of the *Temnothorax salvini*. *Systematic Entomology* 46(2): 307–326. <https://doi.org/10.1111/syen.12463>
- Prebus MM (2021b) Taxonomic revision of the *Temnothorax salvini* clade (Hymenoptera, Formicidae), with a key to the clades of New World *Temnothorax*. *PeerJ* 9: e11514. <https://doi.org/10.7717/peerj.11514>
- Rudgers JA, Savage AM, Rúa MA (2010) Geographic variation in a facultative mutualism: Consequences for local arthropod composition and diversity. *Oecologia* 163(4): 985–996. <https://doi.org/10.1007/s00442-010-1584-6>
- Sabu TK, Shiju RT, Vinod K, Nithya S (2011) A Comparison of the Pitfall Trap, Winkler Extractor and Berlese Funnel for Sampling Ground-Dwelling Arthropods in Tropical Montane Cloud Forests. *Journal of Insect Science* 11(28): 1–19. <https://doi.org/10.1673/031.011.0128>
- Santos-Silva A, Mal N, Roie MV, Jocqué M (2018) A new species of *Derobrachus* Audinet-Serville, 1832 (Coleoptera, Cerambycidae, Prioninae) from a cloud forest in Honduras. *Zootaxa* 4422(3): 395–402. <https://doi.org/10.11646/zootaxa.4422.3.5>

- Santos-Silva A, Van Roie M, Jocqué M (2021) Longhorned woodboring beetles (Coleoptera, Cerambycidae) from Cusuco National Park, Honduras: New species, new records, and revalidation. *European Journal of Taxonomy* 764(1): 37–61. <https://doi.org/10.5852/ejt.2021.764.1469>
- Schmidt CA, Shattuck SO (2014) The Higher Classification of the Ant Subfamily Ponerinae (Hymenoptera, Formicidae), with a Review of Ponerine Ecology and Behavior. *Zootaxa* 3817(1): 1. <https://doi.org/10.11646/zootaxa.3817.1.1>
- Shattuck SO (1992) Generic revision of the ant subfamily Dolichoderinae (Hymenoptera, Formicidae). *Sociobiology*: 21.
- Solomon SE, Rabeling C, Sosa-Calvo J, Lopes CT, Rodrigues A, Vasconcelos HL, Bacci Jr M, Mueller UG, Schultz TR (2019) The molecular phylogenetics of *Trachymyrmex* Forel ants and their fungal cultivars provide insights into the origin and coevolutionary history of ‘higher-attine’ ant agriculture. *Systematic Entomology* 44(4): 939–956. <https://doi.org/10.1111/syen.12370>
- Stadtmüller T (1986) Cloud Forests in the Humid Tropics. A Bibliographic Review. The United Nations University, Tokyo, 81 pp.
- Tercel MPTG, Cuff JP, Symondson WOC, Vaughan IP (2023) Non-native ants drive dramatic declines in animal community diversity: A meta-analysis. *Insect Conservation and Diversity* 16(6): 733–744. <https://doi.org/10.1111/icad.12672>
- Ward PS (1985) The Nearctic species of the genus *Pseudomyrmex* (Hymenoptera, Formicidae). Las especies neárticas del género *Pseudomyrmex* (Hymenoptera, Formicidae). *Quaestiones Entomologicae* 21: 209–246.
- Ward PS, Fisher BL (2016) Tales of dracula ants: The evolutionary history of the ant subfamily Amblyoponinae (Hymenoptera, Formicidae). *Systematic Entomology* 41(3): 683–693. <https://doi.org/10.1111/syen.12186>
- Ward PS, Blaimer BB, Fisher BL (2016) A revised phylogenetic classification of the ant subfamily Formicinae (Hymenoptera, Formicidae), with resurrection of the genera *Colobopsis* and *Dinomyrmex*. *Zootaxa* 4072(3): 343. <https://doi.org/10.11646/zootaxa.4072.3.4>
- Wheeler WM (1907) A Collection of Ants from British Honduras. *Bulletin of the American Museum of Natural History*: 23.
- Wild AL (2007) Taxonomic revision of the ant genus *Linepithema* (Hymenoptera, Formicidae). *University of California Publications in Entomology* 126: 1–159.
- Williams JL, Lapolla JS (2016) Taxonomic revision and phylogeny of the ant genus *Prenolepis* (Hymenoptera, Formicidae). *Zootaxa* 4200(2): 201–258. <https://doi.org/10.11646/zootaxa.4200.2.1>
- Wong MKL, Economo EP, Guénard B (2023) The global spread and invasion capacities of alien ants. *Current Biology* 33(3): 566–571.e3. <https://doi.org/10.1016/j.cub.2022.12.020>
- Yanoviak SP, Kaspari M (2000) Community structure and the habitat templet: Ants in the tropical forest canopy and litter. *Oikos* 89(2): 259–266. <https://doi.org/10.1034/j.1600-0706.2000.890206.x>

## Appendix 1

**Table A1.** Ant species collected within Cusuco National Park, northwestern Honduras. The list is broken down by sub-family, collection method (Winkler sampling relates to specimens obtained through leaf litter extraction via Mini- and Maxi-Winkler), elevation range, and project collectors. \*Non-species level taxa; \*\*Subspecies level (some specimens were not identified to subspecies level; if the respective information for specimens with only species level identification deviates from that of the subspecies, it is shown between parentheses).

Species (per subfamily)	Collection	Elevation Range (m.a.s.l.)	Habitat	Project
<b>Amblyoponinae</b>				
<i>Prionopelta antillana</i> complex*	Winkler	1220	Mesophyll Forest	LLAMA
<b>Dolichoderinae</b>				
<i>Azteca alfari</i> (Emery, 1893)	Veg. Beating	1310	Mesophyll Forest	LLAMA
<i>Azteca</i> cf. <i>coeruleipennis</i>	Pitfall	1588	Mesophyll Forest	MED
<i>Azteca constructor/instablis</i>	Hand Coll.	1546	Mesophyll Forest	MED
<i>Bothriomyrmex paradoxus</i> (Dubovikoff & Longino, 2004)	Beating, Baiting, Hand Coll.	1220–1340	Mesophyll Forest	LLAMA
<i>Linepithema dispertitum</i> (Forel, 1885)	Veg. Beating	1310	Mesophyll Forest	LLAMA
<i>Tapinoma ramulorum</i>	Baiting, Veg. Beating	1210–1340	Mesophyll Forest	LLAMA
<i>Tapinoma</i> JTL-003	Baiting	1330	Mesophyll Forest	LLAMA
<i>Technomyrmex</i> JTL-001	Pitfall, Baiting, Malaise	1260–1336	Mesophyll Forest	LLAMA, MED
<b>Dorylinae</b>				
<i>Cheliomyrmex morosus</i> (Smith, 1859)	Hand Coll.	1270	No-Data	MED
<i>Cylindromyrmex meinerti</i> (Forel, 1905)	Veg. Beating	1300	Mesophyll Forest	LLAMA
<i>Eciton burchellii parvispinum</i> (Forel, 1899) **	Winkler, Hand Coll. (Pitfall, Hand Coll.)	1220–1628 (1364–1964)	Mesophyll Forest	LLAMA, MED
<i>Eciton mexicanum</i> (Roger, 1863)	Pitfall, Hand Coll.	1364–1637	Mesophyll Forest	MED
<i>Eciton vagans angustatum</i> (Roger, 1863) **	Pitfall, Hand Coll. (Pitfall)	1174–1336 (1174–1415)	Mesophyll Forest, Deforested	MED
<i>Labidus coecus</i> (Latreille, 1802)	Winkler, Pitfall	1407–1964	Mesophyll Forest	MED
<i>Labidus praedator</i> (Smith, 1858)	Pitfall, Hand Coll.	1197–1941	Mesophyll Forest	MED
<i>Leptanilloides gracilis</i> (Borowiec & Longino, 2011)	Malaise	1260	Mesophyll Forest	LLAMA
<i>Neivamyrmex halidai</i> (Shuckard, 1840)	Winkler	1290–1340	Mesophyll Forest	LLAMA
<i>Neivamyrmex sumichrasti</i> (Norton, 1868)	Pitfall, Hand Coll.	1197–1613	Mesophyll Forest	MED
<i>Syscia</i> JTL082	Winkler, Malaise	1260–2030	Cloud Forest, Mesophyll Forest, Pine-liquidambar Forest	LLAMA
<i>Syscia parietalis</i> (Longino & Branstetter, 2021)	Winkler, Malaise	1260–1330	Mesophyll Forest, Pine-liquidambar Forest	LLAMA
<i>Syscia persimilis</i> (Longino & Branstetter, 2021)	Winkler	1290–1300	Mesophyll Forest	LLAMA
<i>Syscia tolteca</i> (Forel, 1909)	Berlese	1550	Cloud Forest	LLAMA
<b>Ectatomminae</b>				
<i>Alfaria minuta</i> (Emery, 1896)	Winkler	1235–1340	Mesophyll Forest	LLAMA, MED
<i>Alfaria simulans</i> (Emery, 1896)	Winkler, Pitfall	1210–1599	Mesophyll Forest	LLAMA, MED
<i>Gnamptogenys interrupta</i> (Mayr, 1887)	Hand Coll.	1331	Mesophyll Forest	MED
<i>Gnamptogenys</i> JTL-010	Winkler	1260–1330	Mesophyll Forest	LLAMA
<i>Gnamptogenys mordax</i> (Smith, 1858)	Pitfall	1315	Mesophyll Forest	MED
<i>Gnamptogenys sulcata</i> (Smith, 1858)	Malaise	1260	Pine-liquidambar Forest	LLAMA
<i>Holcoponera porcata</i> (Emery, 1896)	Pitfall, Hand Coll.	1331–1718	Mesophyll Forest, Deforested	LLAMA, MED

Species (per subfamily)	Collection	Elevation Range (m.a.s.l.)	Habitat	Project
<i>Holcaponera strigata</i> (Norton, 1868)	Winkler, Pitfall, Veg. Beating, Baiting, Hand Coll., Malaise	1174–1718	Mesophyll Forest, Pine-liquidambar Forest, Coffee Plantation, Deforested	LLAMA, MED
<i>Typhlomyrmex</i> indet.*	Malaise	1260	Pine-liquidambar Forest	LLAMA
<b>Formicinae</b>				
<i>Acropyga exsanguis</i> (Wheeler, 1909)	Winkler	1330–1340	Mesophyll Forest	LLAMA
<i>Camponotus abscisus</i> (Roger, 1863)	Winkler, Veg. Beating, Baiting, Hand Coll., Malaise	1260–1838	Mesophyll Forest, Pine-liquidambar Forest	LLAMA, MED
<i>Camponotus albicoxis</i> (Forel, 1899)	Veg. Beating, Pitfall, Hand Coll., Malaise	1234–1613	Mesophyll Forest	LLAMA, MED
<i>Camponotus atriceps</i> (Smith, 1858)	Veg. Beating, Pitfall, Baiting, Hand Coll.	1270–1639	Mesophyll Forest	LLAMA, MED
<i>Camponotus</i> cf. <i>senex</i>	Hand Coll.	1190	Deforested Village	MED
<i>Camponotus cuneidorsus</i>	Veg. Beating, Baiting, Hand Coll.	1270–1613	Mesophyll Forest	LLAMA, MED
<i>Camponotus planatus</i> (Roger, 1863)	Veg. Beating	1330	Mesophyll Forest	LLAMA
<i>Camponotus sericeiventris</i> (Guérin-Méneville, 1838)	Hand Coll.	1197	Mesophyll Forest	MED
<i>Myrmelachista</i> indet.*	Hand Coll.	1331	Mesophyll Forest	MED
<i>Nylanderia</i> indet.*	Veg. Beating, Baiting, Winkler, Malaise, Pitfall, Hand Coll.	1220–1613	Mesophyll Forest, Deforested	LLAMA, MED
<b>Myrmicinae</b>				
<i>Acanthognathus ocellatus</i> (Mayr, 1887)	Malaise	1210	Mesophyll Forest	LLAMA
<i>Acromyrmex coronatus</i> (Fabricius, 1804)	Veg. Beating, Pitfall, Hand Coll.	1260–1639	Mesophyll Forest	LLAMA, MED
<i>Acromyrmex volcanus</i> (Wheeler, 1937)	Hand Coll.	~500	NO-DATA	MED
<i>Adelomyrmex</i> JTL-024	Berlese	1550	Cloud Forest	LLAMA
<i>Adelomyrmex silvestrii</i> (Menozi, 1931)	Winkler, Hand Coll.	1220–1613	Mesophyll Forest	LLAMA, MED
<i>Apterostigma pilosum</i> complex*	Winkler, Veg. Beating, Baiting	1210–1330	Mesophyll Forest	LLAMA
<i>Atta cephalotes</i> (Linnaeus, 1758)	Hand Coll.	498–1613	Mesophyll Forest, Deforested	MED
<i>Carebara intermedia</i> (Fernández, 2004)	Winkler, Baiting, Hand Coll.	1210–1330	Mesophyll Forest	LLAMA
<i>Carebara urichi</i> (Wheeler, 1922)	Winkler	1260–1340	Mesophyll Forest	LLAMA
<i>Cephalotes</i> cf. <i>multispinosus</i>	Hand Coll.	NA (lowland)	NO-DATA (forest)	MED
<i>Crematogaster crinosa</i> (Mayr, 1862)	Veg. Beating	1340	Mesophyll Forest	LLAMA
<i>Crematogaster montezumia</i> (Smith, 1858)	Malaise	1210	Mesophyll Forest	LLAMA
<i>Cyphomyrmex rimosus</i> s.l.	Malaise	1260	Pine-liquidambar Forest	LLAMA
<i>Cyphomyrmex salvini</i> (Forel, 1899)	Winkler, Pitfall, Baiting, Hand Coll.	1174–1340	Mesophyll Forest, Deforested	LLAMA, MED
<i>Eurhopalothrix hunhau</i> (Longino, 2013)	Berlese	1550	Cloud Forest	LLAMA
<i>Eurhopalothrix zipacna</i> (Longino, 2013)	Winkler, Baiting	1260–1340	Mesophyll Forest	LLAMA
<i>Hylomyrma versuta</i> (Kempf, 1973)	Winkler, Pitfall	1174–1340	Mesophyll Forest, Deforested	LLAMA, MED
<i>Megalomyrmex megadrifti</i> (Boudinot et al., 2013)	Winkler	1290–1330	Mesophyll Forest	LLAMA
<i>Monomorium pharaonis</i> (Linnaeus, 1758)	Hand Coll.	1613	Mesophyll Forest	MED



Species (per subfamily)	Collection	Elevation Range (m.a.s.l.)	Habitat	Project
<i>Mycetomoellerius squamulifer</i> (Emery, 1896)	Baiting	1290	Mesophyll Forest	LLAMA
<i>Mycetophylax andersoni</i> (MacKay & Serna, 2010)	Winkler, Baiting	1220–1340	Mesophyll Forest	LLAMA
<i>Octostruma balzani</i> complex	Winkler, Baiting	1210–1340	Mesophyll Forest	LLAMA
<i>Octostruma gymnogon</i> (Longino, 2013)	Winkler, Baiting	1260–1330	Mesophyll Forest	LLAMA
<i>Octostruma lepticeps</i> (Longino, 2013)	Winkler	1290	Mesophyll Forest	LLAMA
<i>Pheidole absurda</i> (Forel, 1886)	Hand Coll.	1190	Deforested Village	MED
<i>Pheidole balatro</i> (Longino, 2019)	Winkler, Veg. Beating, Pitfall, Baiting, Hand Coll.	1270–1620	Mesophyll Forest	LLAMA, MED
<i>Pheidole biconstricta</i> (Mayr, 1870)	Winkler, Baiting	1234–1290	Mesophyll Forest	LLAMA, MED
<i>Pheidole bilimeki</i> (Mayr, 1870)	Winkler, Veg. Beating, Baiting	1260–1330	Mesophyll Forest	LLAMA
<i>Pheidole branstetteri</i> (Longino, 2009)	Veg. Beating, Malaise	1210–1330	Mesophyll Forest	LLAMA
<i>Pheidole browni</i> (Wilson, 2003)	Winkler, Pitfall, Baiting, Hand Coll.	1210–1415	Mesophyll Forest	LLAMA, MED
<i>Pheidole cusuco</i> (Longino, 2019)	Winkler, Veg. Beating, Pitfall, Baiting	1225–1421	Mesophyll Forest	LLAMA, MED
<i>Pheidole deceptrix</i> (Forel, 1899)	Winkler, Baiting, Hand Coll.	1260–1330	Mesophyll Forest	LLAMA
<i>Pheidole guerrerana</i> (Wilson, 2003)	Winkler, Veg. Beating	1260–1330	Mesophyll Forest	LLAMA
<i>Pheidole gulo</i> (Wilson, 2003)	Winkler, Veg. Beating, Pitfall, Baiting, Hand Coll., Malaise	1210–1697	Mesophyll Forest	LLAMA, MED
<i>Pheidole harrisonfordi</i> (Wilson, 2003)	Winkler, Baiting	1210–1360	Mesophyll Forest	LLAMA
<i>Pheidole indagatrix</i> (Wilson, 2003)	Winkler, Baiting, Malaise	1260–1320m	Mesophyll Forest	LLAMA
<i>Pheidole insipida</i> (Forel, 1899)	Winkler, Veg. Beating, Baiting, Hand Coll., Malaise	1210–1388	Mesophyll Forest	LLAMA, MED
<i>Pheidole</i> JTL-209	Malaise	1210–1260	Mesophyll Forest	LLAMA
<i>Pheidole lagunculinor</i> (Longino, 2019)	Winkler, Baiting	1210–1360	Mesophyll Forest	LLAMA
<i>Pheidole natalie</i> (Longino, 2019)	Winkler, Baiting	1270–1340	Mesophyll Forest	LLAMA
<i>Pheidole rectispina</i> (Wilson, 2003)	Baiting	1290	Mesophyll Forest	LLAMA
<i>Pheidole tschinkeli</i> (Wilson, 2003)	Winkler, Veg. Beating, Baiting, Hand Coll., Malaise	1210–1360	Mesophyll Forest	LLAMA, MED
<i>Pheidole ursus</i> (Mayr, 1870)	Winkler, Veg. Beating, Pitfall, Baiting, Hand Coll.	1210–1594	Mesophyll Forest, Deforested	LLAMA, MED
<i>Procryptocerus batesi</i> (Forel, 1899)	Winkler, Veg. Beating, Hand Coll., Malaise	400–1648	Mesophyll Forest, Coffee Plantation	LLAMA, MED
<i>Rhopalothrix andersoni</i> (Longino & Boudinot, 2013)	Winkler	1300	Mesophyll Forest	LLAMA
<i>Rogeria innotabilis</i> (Kugler, 1994)	Winkler	1210–1340	Mesophyll Forest	LLAMA
<i>Rogeria</i> JTL-009	Malaise	1260	Pine-liquidambar Forest	LLAMA
<i>Solenopsis geminata</i> (Fabricius, 1804)	Hand Coll.	1190–1407	Mesophyll Forest, Deforested Village	MED
<i>Solenopsis invicta</i> (Buren, 1972)	Hand Coll.	1190	Deforested Village	MED

Species (per subfamily)	Collection	Elevation Range (m.a.s.l.)	Habitat	Project
<i>Solenopsis terricola</i> (Menozzi, 1931)	Winkler, Pitfall	1331–1599	Mesophyll Forest	MED
<i>Solenopsis texana/carolinensis</i>	Hand Coll.	1668	Deforested	MED
<i>Stenamma atribellum</i> (Branstetter, 2013)	Hand Coll., Berlese	1550–2030	Cloud Forest	LLAMA, MED
<i>Stenamma brujita</i> (Branstetter, 2013)	Winkler, Baiting, Berlese	1210–1550	Cloud Forest, Mesophyll Forest	LLAMA
<i>Stenamma crypticum</i> (Branstetter, 2013)	Winkler	2030	Cloud Forest	LLAMA
<i>Stenamma cusuco</i> (Branstetter, 2013)	Winkler	1280–1330	Mesophyll Forest	LLAMA
<i>Stenamma felixi</i> (Mann, 1922)	Winkler, Baiting, Hand Coll., Malaise	1260–1613	Mesophyll Forest	LLAMA, MED
<i>Stenamma hojarasca</i> (Branstetter, 2013)	Winkler	1220–1340	Mesophyll Forest	LLAMA
<i>Stenamma ignotum</i> (Branstetter, 2013)	Winkler, Berlese	1300–1550	Cloud Forest, Mesophyll Forest	LLAMA
<i>Stenamma manni</i> (Wheeler, 1914)	Pitfall	1472–1845	Mesophyll Forest	LLAMA, MED
<i>Stenamma murella</i> (Branstetter, 2013)	Hand Coll.	1677	Mesophyll Forest	MED
<i>Stenamma ochrocnemis</i> (Branstetter, 2013)	Winkler	2030	Cloud Forest	LLAMA
<i>Stenamma pelophilum</i> (Branstetter, 2013)	Baiting, Hand Coll.	1290–1320	Mesophyll Forest	LLAMA
<i>Stenamma picopicucha</i> (Branstetter, 2013)	Winkler	2030	Cloud Forest	LLAMA
<i>Stenamma saenzae</i> (Branstetter, 2013)	Winkler	1210–1340	Mesophyll Forest	LLAMA
<i>Strumigenys biolleyi</i> (Forel, 1908)	Winkler, Baiting, Hand Coll.	1260–1613	Mesophyll Forest	LLAMA, MED
<i>Strumigenys brevicornis</i> (Mann, 1922)	Winkler	1210–1340	Mesophyll Forest	LLAMA
<i>Strumigenys cassicuspis</i> (Bolton, 2000)	Winkler	1290	Mesophyll Forest	LLAMA
<i>Strumigenys cf. calamita</i>	Winkler	1599	Mesophyll Forest	MED
<i>Strumigenys cf. myllorhapha</i>	Winkler	1331	Mesophyll Forest	MED
<i>Strumigenys elongata</i> (Roger, 1863)	Winkler	1300	Mesophyll Forest	LLAMA
<i>Strumigenys excisa</i> (Weber, 1934)	Winkler	1220	Mesophyll Forest	LLAMA
<i>Strumigenys gundlachi</i> (Roger, 1862)	Winkler, Baiting, Malaise	1210–1340	Mesophyll Forest, Pine-liquidambar Forest	LLAMA, MED
<i>Strumigenys humata</i> (Lattke & Goitía, 1997)	Winkler	1220–1330	Mesophyll Forest	LLAMA
<i>Strumigenys</i> JTL-028	Winkler	1290–1300	Mesophyll Forest	LLAMA
<i>Strumigenys</i> JTL-pyr020	Winkler	1290	Mesophyll Forest	LLAMA
<i>Strumigenys microthrix</i> (Kempf, 1975)	Winkler	1280	Mesophyll Forest	LLAMA
<i>Strumigenys paradoxa</i> (Bolton, 2000)	Winkler	1290–1340	Mesophyll Forest	LLAMA
<i>Strumigenys rogata</i> (Bolton, 2000)	Winkler	1260–1290	Mesophyll Forest	LLAMA
<i>Strumigenys subedentata</i> (Mayr, 1887)	Pitfall, Malaise	1260–1263	Mesophyll Forest, Pine-liquidambar Forest	LLAMA, MED
<i>Strumigenys timicala</i> (Bolton, 2000)	Winkler	1330	Mesophyll Forest	LLAMA
<i>Temnothorax altinodus</i> (Prebus, 2021)	Veg. Beating	1290	Mesophyll Forest	LLAMA
<i>Temnothorax aztecus</i> (Wheeler, 1931)	Winkler, Veg. Beating	1220–1310	Mesophyll Forest	LLAMA
<i>Temnothorax cf. longinoi</i>	Hand Coll.	1364	Mesophyll Forest	MED
<i>Temnothorax</i> med01	Hand Coll.	1838	Mesophyll Forest	MED
<i>Temnothorax</i> med02	Winkler	1838	Mesophyll Forest	MED
<i>Temnothorax</i> med03	Hand Coll.	1331	Mesophyll Forest	MED
<i>Temnothorax terraztecus</i> (Prebus, 2021)	Winkler	1220	Mesophyll Forest	LLAMA
<b>Ponerinae</b>				
<i>Anochetus mayri</i> (Emery, 1884)	Winkler	1300–1330	Mesophyll Forest	LLAMA

Species (per subfamily)	Collection	Elevation Range (m.a.s.l.)	Habitat	Project
<i>Belonopelta deletrix</i> (Mann, 1922)	Winkler, Malaise	1260–1330	Mesophyll Forest, Pine-liquidambar Forest	LLAMA
<i>Cryptopone gilva</i> (Roger, 1863)	Winkler, Hand Coll.	1599–1838	Mesophyll Forest	MED
<i>Hypoponera nitidula</i> (Emery, 1890)	Winkler, Pitfall, Baiting	1210–1415	Mesophyll Forest	LLAMA, MED
<i>Hypoponera parva</i> (Forel, 1909)	Winkler	1210–1330	Mesophyll Forest	LLAMA
<i>Leptogenys</i> BEB003	Malaise	1210	Mesophyll Forest	LLAMA
<i>Leptogenys bifida</i> (Lattke, 2011)	Hand Coll.	1331	Mesophyll Forest	MED
<i>Leptogenys</i> cf. <i>foveonates</i>	Pitfall	1263	Mesophyll Forest	MED
<i>Leptogenys honduriana</i> Mann, 1922	Pitfall	1174–1501	Mesophyll Forest, Deforested	MED
<i>Leptogenys imperatrix</i> (Mann, 1922)	Pitfall, Hand Coll.	1197–1718	Mesophyll Forest	MED
<i>Leptogenys</i> JTL-023	Winkler	1290–1330	Mesophyll Forest	LLAMA
<i>Leptogenys tiobil</i> (Lattke, 2011)	Pitfall	1597	Mesophyll Forest	MED
<i>Neoponera apicalis</i> (Latreille, 1802)	Hand Coll.	498–1331	Mesophyll Forest	MED
<i>Neoponera crenata</i> (Roger, 1861)	Pitfall, Malaise	1197–1260	Mesophyll Forest	LLAMA, MED
<i>Neoponera curvinodis</i> (Forel, 1899)	NO-DATA	NO-DATA	NO-DATA	MED
<i>Neoponera lineaticeps</i> (Mayr, 1866)	Veg. Beating, Pitfall, Hand Coll., Malaise	1210–1630	Mesophyll Forest, Pine-liquidambar Forest	LLAMA, MED
<i>Odontomachus haematodus</i> (Linnaeus, 1758)	Pitfall, Hand Coll.	498–1639	Mesophyll Forest	MED
<i>Odontomachus laticeps</i> (Roger, 1861)	Winkler, Baiting	1290–1330	Mesophyll Forest	LLAMA
<i>Odontomachus yucatecus</i> (Brown, 1976)	Hand Coll.	1331–1639	Mesophyll Forest	MED
<i>Pachycondyla harpax</i> (Fabricius, 1804)	Winkler, Pitfall, Baiting, Hand Coll.	498–1612	Mesophyll Forest	LLAMA, MED
<i>Pachycondyla purpurascens</i> (Forel, 1899)	Winkler, Pitfall, Baiting, Hand Coll.	400–1613	Mesophyll Forest, Coffee Plantation, Deforested	LLAMA, MED
<i>Platythyrea prizo</i> (Kugler, 1977)	Malaise	1210–1260	Mesophyll Forest, Pine-liquidambar Forest	LLAMA
<i>Ponera exotica</i> (Smith, 1962)	Malaise	1260	Pine-liquidambar Forest	LLAMA
<i>Rasopone mesoamericana</i> (Longino & Branstetter, 2020)	Winkler, Pitfall	1220–1514	Mesophyll Forest	LLAMA, MED
<i>Rasopone politognatha</i> (Longino & Branstetter, 2020)	Winkler, Pitfall	1290–1514	Mesophyll Forest	LLAMA, MED
<i>Thaumatomyrmex ferox</i> complex*	Winkler, Hand Coll.	1210–1340	Mesophyll Forest	LLAMA
<b>Proceratiinae</b>				
<i>Discothyrea horni</i> complex*	Winkler	1210–2030	Cloud Forest, Mesophyll Forest	LLAMA
<i>Proceratium mancum</i> (Mann, 1922)	Winkler	1300	Mesophyll Forest	LLAMA
<b>Pseudomyrmecinae</b>				
<i>Pseudomyrmex ejectus</i> (Smith, 1858)	Malaise	1260	Mesophyll Forest	LLAMA
<i>Pseudomyrmex elongatulus</i> complex	Hand Coll.	1364–1838	Mesophyll Forest	MED
<i>Pseudomyrmex pallens</i> (Mayr, 1870)	Hand Coll.	1190	Deforested Village	MED
<i>Pseudomyrmex</i> PSW-159	Veg. Beating, Baiting	1220–1330	Mesophyll Forest	LLAMA
<i>Pseudomyrmex</i> PSW-53	Veg. Beating, Baiting	1210–1330	Mesophyll Forest	LLAMA

## Supplementary material 1

### Ant species collected within Cusuco National Park, northwestern Honduras

Authors: Frederik C. De Wint, Dominik Oorts, Michael G. Branstetter, Dario De Graaf, Wouter Dekoninck, Merlijn Jocque, Thomas E. Martin, Jennifer Sudworth, Ronja Van Osselaer, Matthew T. Hamer

Data type: xlsx

Explanation note: The list is broken down by subfamily, collection method (Winkler sampling relates to specimens obtained through leaf litter extraction via Mini- and MaxiWinkler), elevation range and project collectors. \*Non-species level taxa; \*\*Sub-species level (some specimens were not identified to subspecies level; if respective information for specimens with only species level identification deviates from that of the subspecies, it is shown between parentheses).

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/neotropical.19.e119775.suppl1>

## Supplementary material 2

### Identification key to ant genera of Honduras

Authors: Frederik C. De Wint

Data type: pdf

Explanation note: Dichotomous keys are included to identify all subfamilies and genera currently found in Honduras, based on the worker caste. Records from this work were supplemented with records from the AntWeb database (AntWeb reference: AntWeb. Version 8.87. California Academy of Science, online at <https://www.antweb.org>. Accessed 4 March 2023.). These keys are based on other keys as referred to in each subfamily section below. Respective works can still be used complementary to this key for clarification purposes, i.e. images and terminology.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/neotropical.19.e119775.suppl2>

### Supplementary material 3

#### Pictures of select ant species

Authors: Frederik C. De Wint

Data type: pdf

Explanation note: Pictures of select ant species collected by MyrmEcoDex (2018–2019). This for visualisation purposes accompanying the manuscript, checklist and identification key.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/neotropical.19.e119775.suppl3>

### Supplementary material 4

#### Specimen data of collections within Cusuco National Park, northwestern Honduras

Authors: Frederik C. De Wint, Dominik Oorts, Michael G. Branstetter, Dario De Graaf, Wouter Dekoninck, Merlijn Jocque, Thomas E. Martin, Jennifer Sudworth, Ronja Van Osselaer, Matthew T. Hamer

Data type: xlsx

Explanation note: The file presents the data of all specimens collected in Cusuco National Park. This is the data used as a base for the manuscript, resulting in the respective graphs.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/neotropical.19.e119775.suppl4>